

# *Dynamical Control of the Shape and Size of Stereocilia and Microvilli*

*J.Prost, J.F.J*

*Hair cells, stereocilia; microvilli*

*Kachar's law*

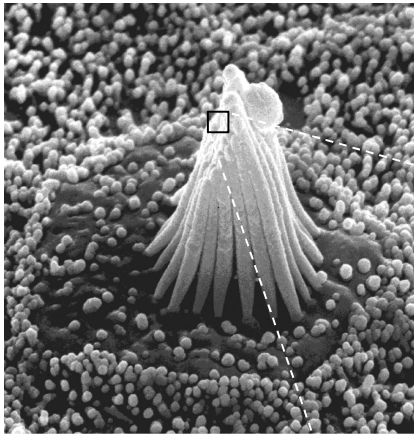
*Tip and Membrane dynamics*

*Stereocilia shape:      depolymerization  
   membrane shape*

*Penetration in the cortical layer*

*Lamellipodium motion*

# *Hair cells*



200 nm



*Ciliated cells of the inner ear*

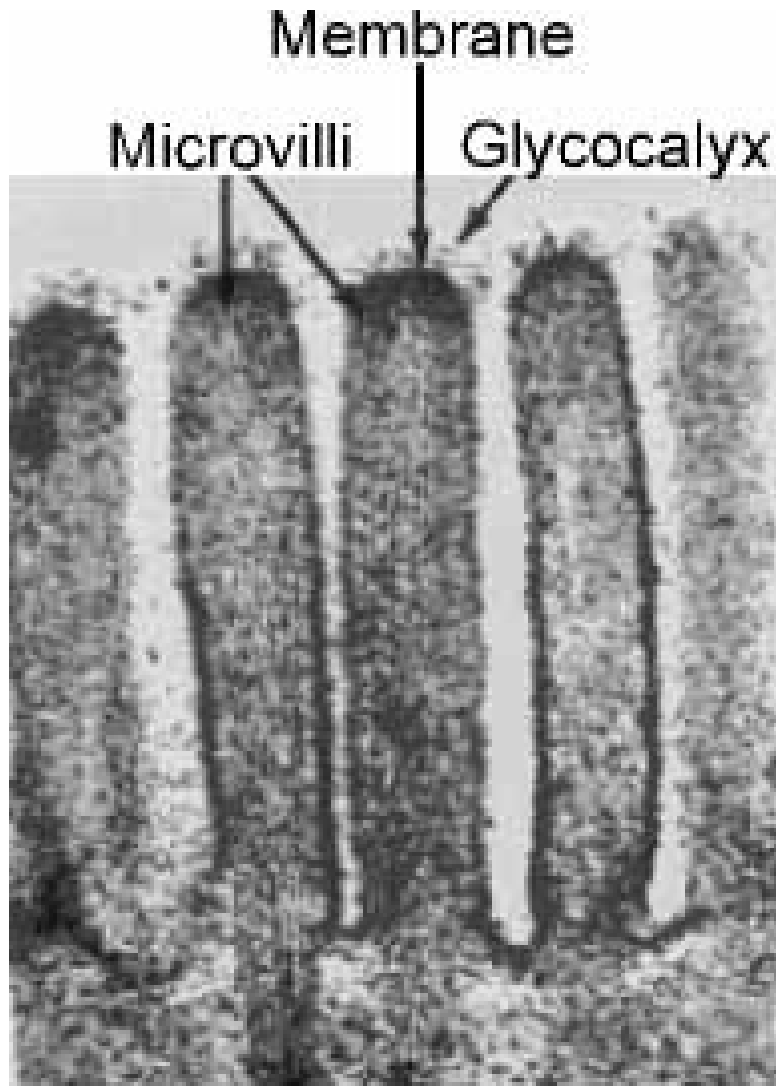
*Bundle of actin cilia*

*Graded sizes in a bundle*  
*Connections via tip-links*

*Bundles of varying sizes*

*Shape and size regulation?*

## *Other ciliated cells*



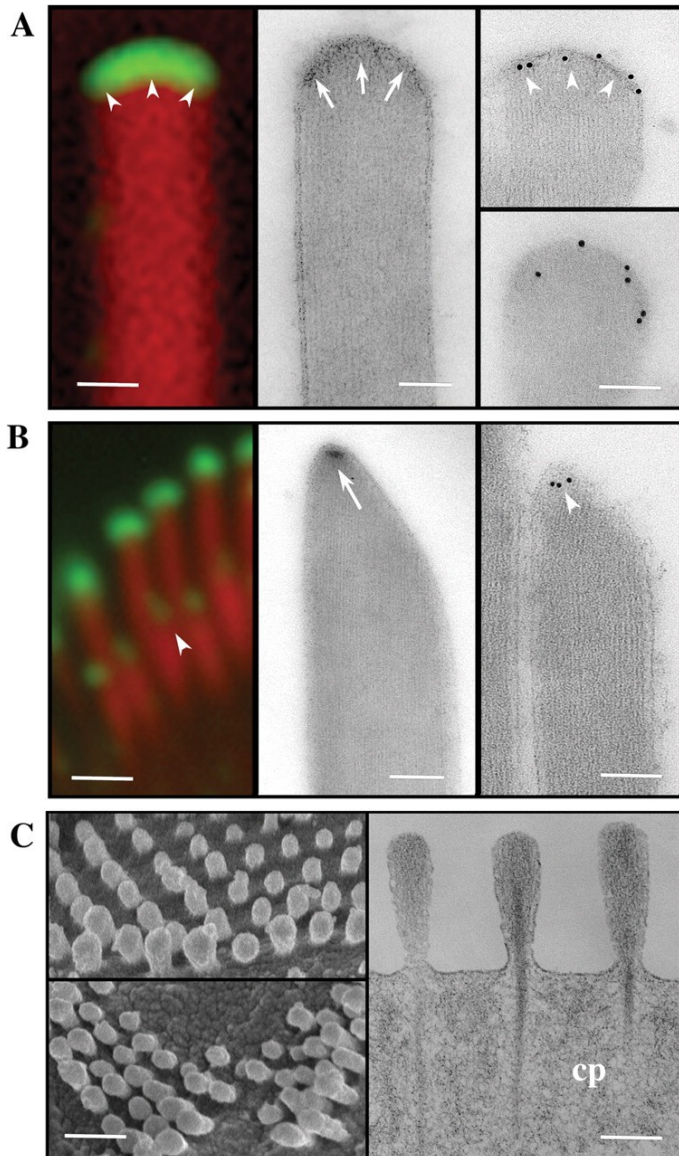
*Intestinal epithelial brush border cells*

*Microvilli (actin structures)*

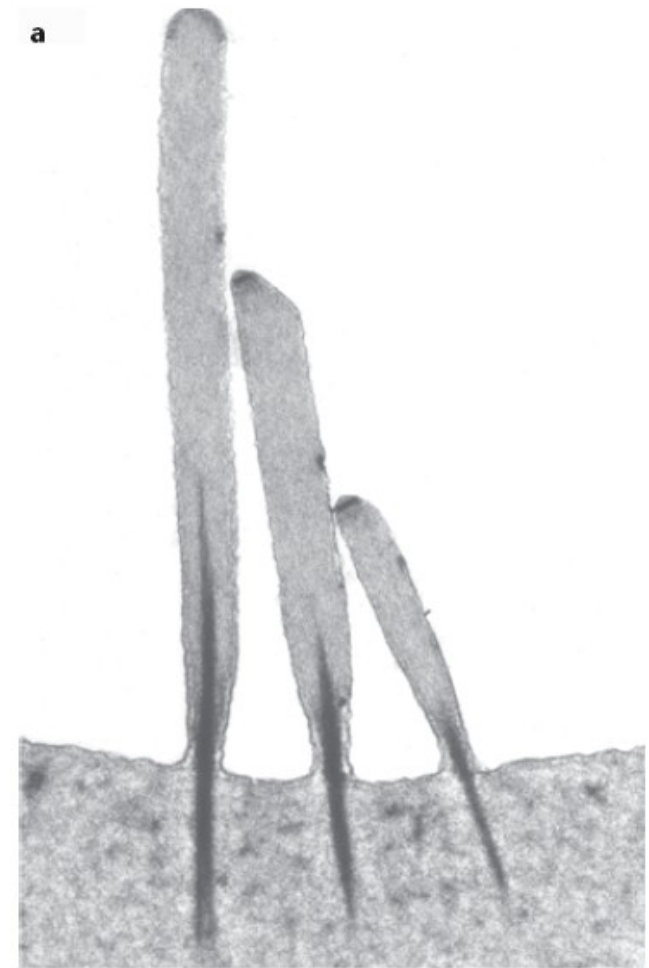
*Spines and boutons in neurons*

*Filipodia-lamellipodia*

# *Stereocilia shape in hairbundle*

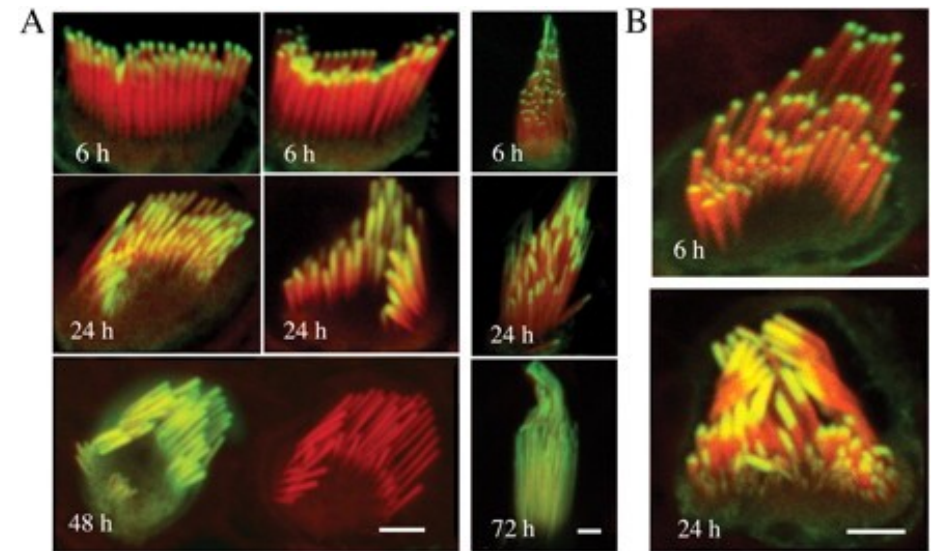
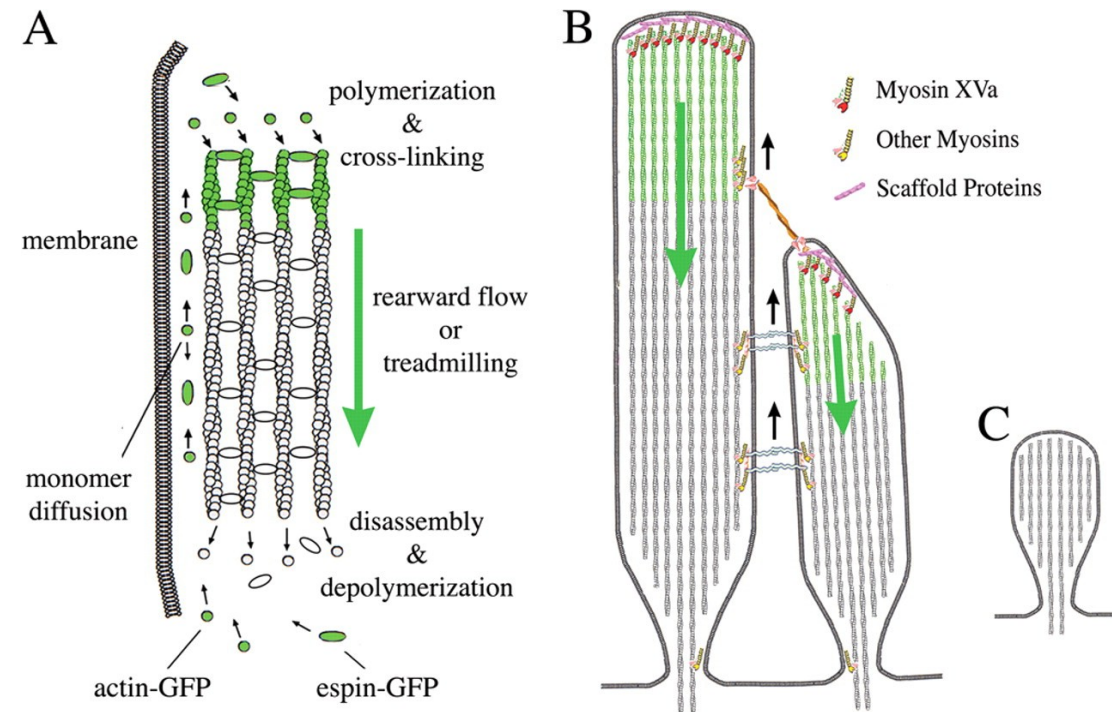


*Radzinska et al*



*Fettiplace and Hackney*

# *Actin and espin in stereocilia* Radzinska et al.

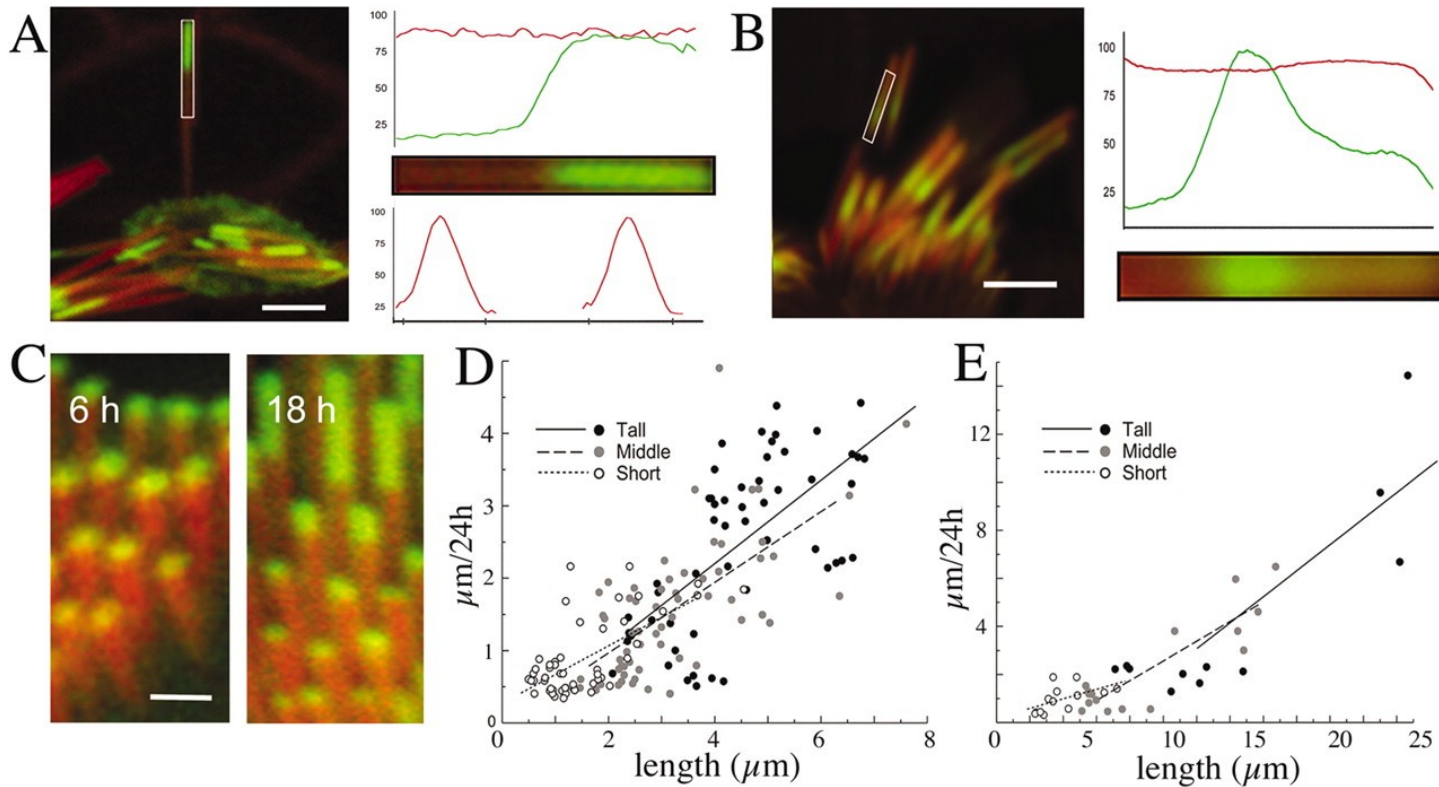


*Stereocilium structure*

*Actin and espin incorporation  
organ of corti and vestibule*



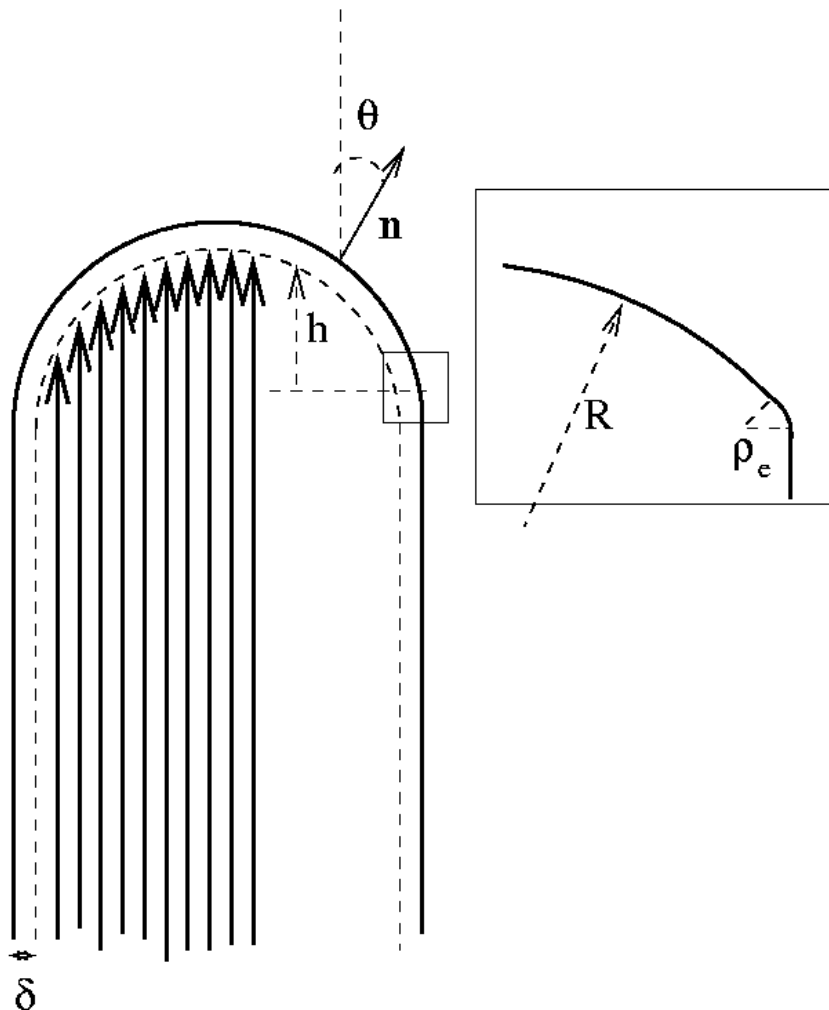
# *Kachar's treadmilling laws Radzinska et al.*



*Treadmilling velocity proportional to stereocilium length  
constant treadmilling time*

*Thicker stereocilia have a larger treadmilling velocity*

# Membrane dynamics and polymerization front



*Free energy*

$$F_m = \int ds_m \left\{ \sigma + \frac{1}{2} \kappa H^2 \right\} - \int P dv$$

$$F_i = \int ds_m \frac{1}{2} k (\delta - \delta_0)^2$$

*Ignore orientation dependence of  $\delta_0$  and  $\sigma$*

*Linear dynamic equation*

$$\mathbf{v}_n = -\lambda_m \frac{\delta F}{\delta \mathbf{r}_n}$$

$$\frac{\partial h}{\partial t} + \mathbf{v}_T = -\lambda_a \frac{\delta F}{\delta h} + \mathbf{v}_p^0$$

## *Steady state treadmilling*

*Membrane shape equation* 
$$\frac{\delta F_m}{\delta r_n} = \frac{v_p^0 - v_T}{\lambda_a} + \frac{1}{2} \frac{H (v_T - v_p^0)^2}{k \lambda_a^2}$$

*Effective tension* 
$$\sigma_{\text{eff}} = \sigma + \frac{1}{2} \frac{(v_T - v_p^0)^2}{k \lambda_a^2}$$

*Effective pressure* 
$$P_{\text{eff}} = P + \frac{v_p^0 - v_T}{\lambda_a}$$

$P_{\text{eff}} \sim 3 \cdot 10^3 \text{ Pa}$ , Total force 100 pN, Force per filament  $3 \cdot 10^{-1} \text{ pN}$

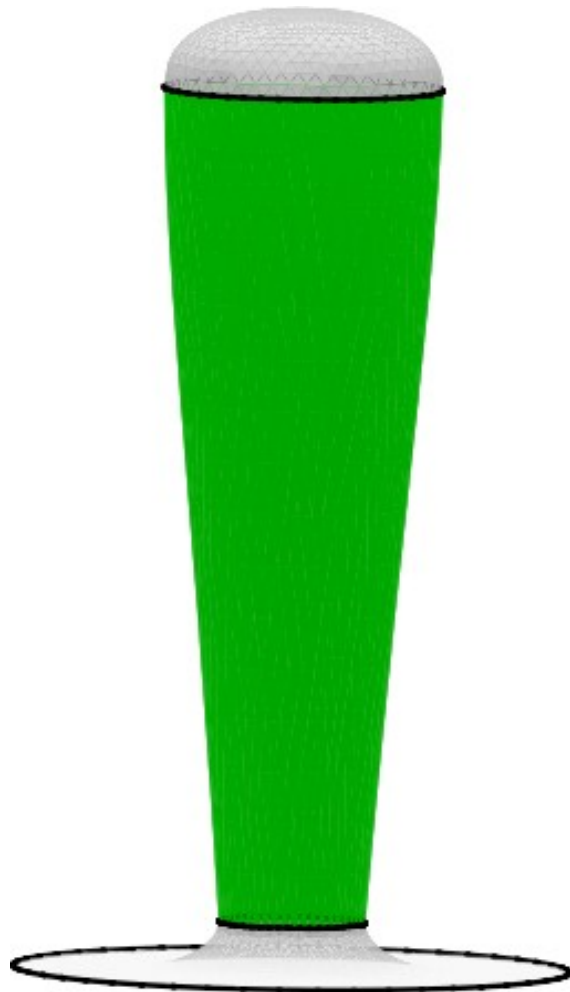
*Treadmilling velocity* 
$$v_T = v_p^0 - \lambda_a \left( \frac{2\sigma_{\text{eff}}}{r_0} + \frac{\kappa}{r_0^3} \right)$$

*Thicker cilia have a large treadmilling velocity Gale et al.*

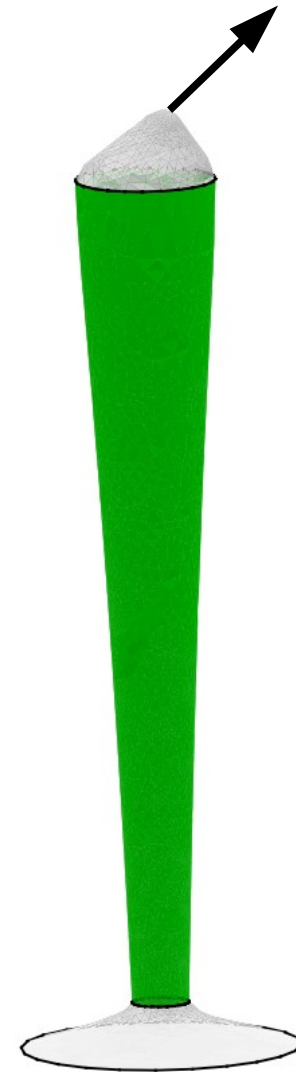
*Mean field theory, ignores non-linear effects*



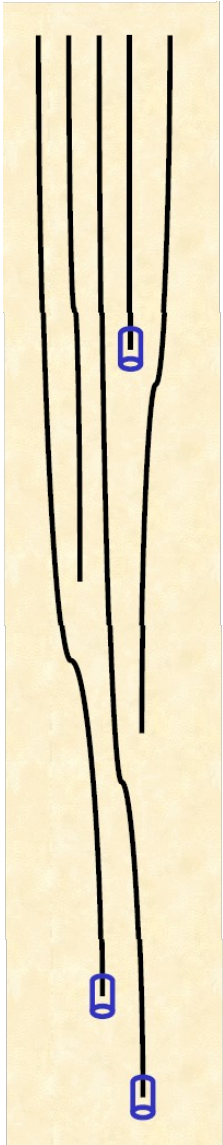
*Tip-link force C.Barbotta*



*Tip-link force 10 pN*



# *Shape of the stem: depolymerization*



*Polymerization at the barbed end*

*At the tip  $z=0$  with a rate  $k_p$*

*Capping protein of the pointed end (espin)*

*prevents depolymerization*

*uncaps with a rate  $k_u$*

*Depolymerization at the pointed end*

*only uncapped filaments*

*rate  $k_d$*

# Depolymerization kinetics

## Rate equations

$$\frac{\partial p^c(n)}{\partial t} = -k_u p^c(n) + k_p p^c(n-1) - k_p p^c(n)$$

$$\frac{\partial p^u(n)}{\partial t} = k_u p^c(n) + k_p p^u(n-1) - k_p p^u(n) + k_d p^u(n+1) - k_d p^u(n)$$

## Stereocilium shape

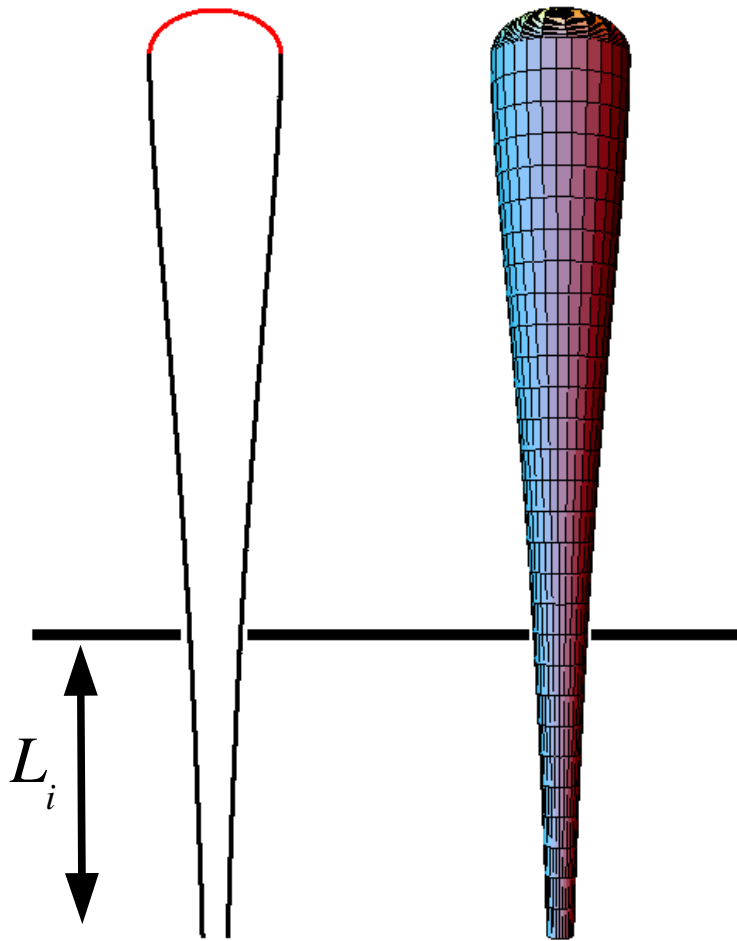
*Constant inplane density*

$$r(z) = r_0 \left[ \frac{k_d - k_p}{k_d - k_u - k_p} \left( \frac{k_p}{k_p + k_u} \right)^{z/a} - \frac{k_u}{k_d - k_u - k_p} \left( \frac{k_p}{k_d} \right)^{z/a} \right]^{1/2}$$

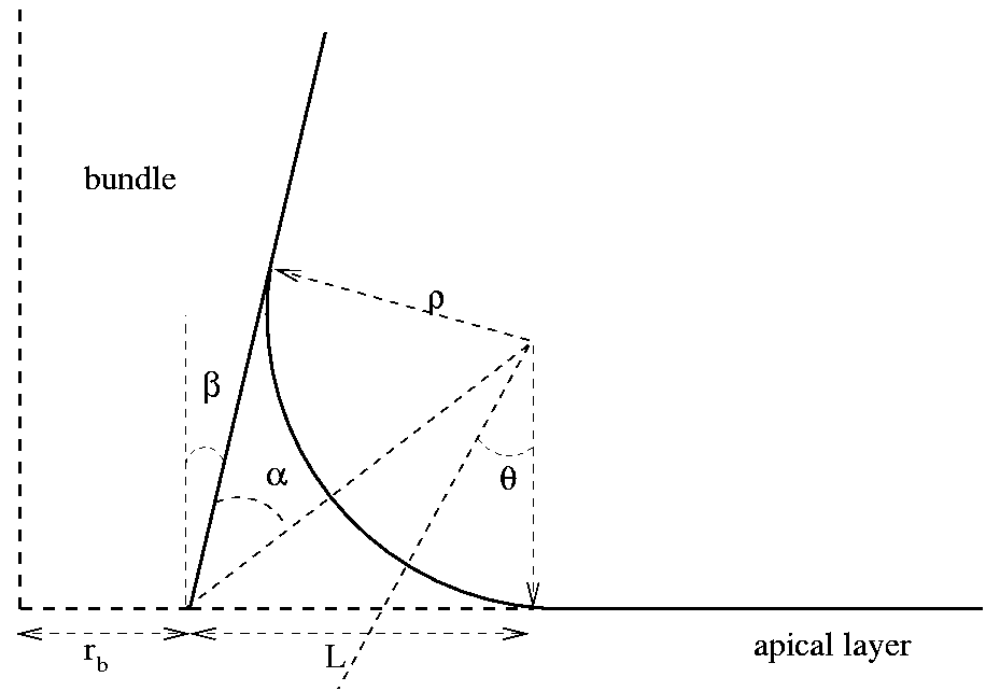
*exponential decay  
over  $\lambda = v_T k_u^{-1}$*

*Kachar's law*  $L_T = 2 v_T k_u^{-1} \log \frac{r_0}{a} \left( \frac{k_d - k_p}{k_d - k_p - k_u} \right)^{1/2}$  *time scale fixed by uncapping*

# Shape of stereocilia



## Membrane shape



Curvature radius  $\rho \sim (\kappa / 2 \tilde{\sigma})^{1/2}$

## *Emerging part of stereocilia*

*Force balance between*

*Membrane force*  $f_m = 2\pi r_b \sigma$

*Viscous friction between actin fascicle and cortical layer*  $f_f = \alpha \eta L_i v_T$

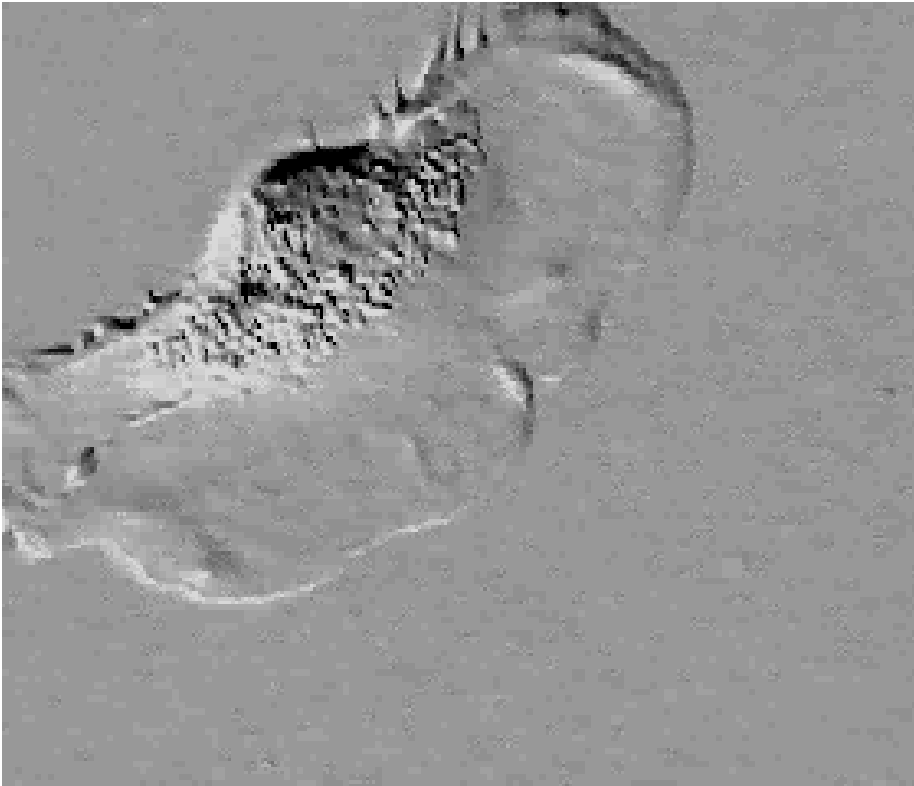
*Immersed length*  $\frac{L_i}{2\lambda} \exp\left(-\frac{L_i}{2\lambda}\right) = \frac{a\sigma}{(\alpha/\pi)\lambda v_T \eta} = \epsilon$

*large  $\epsilon$  totally immersed fascicle*

*small  $\epsilon$  finite immersed length smaller than  $2\lambda$*

*Maximum immersed length  $L_i < L_T/3$*

# *Keratocyte motion*



*Flat region*

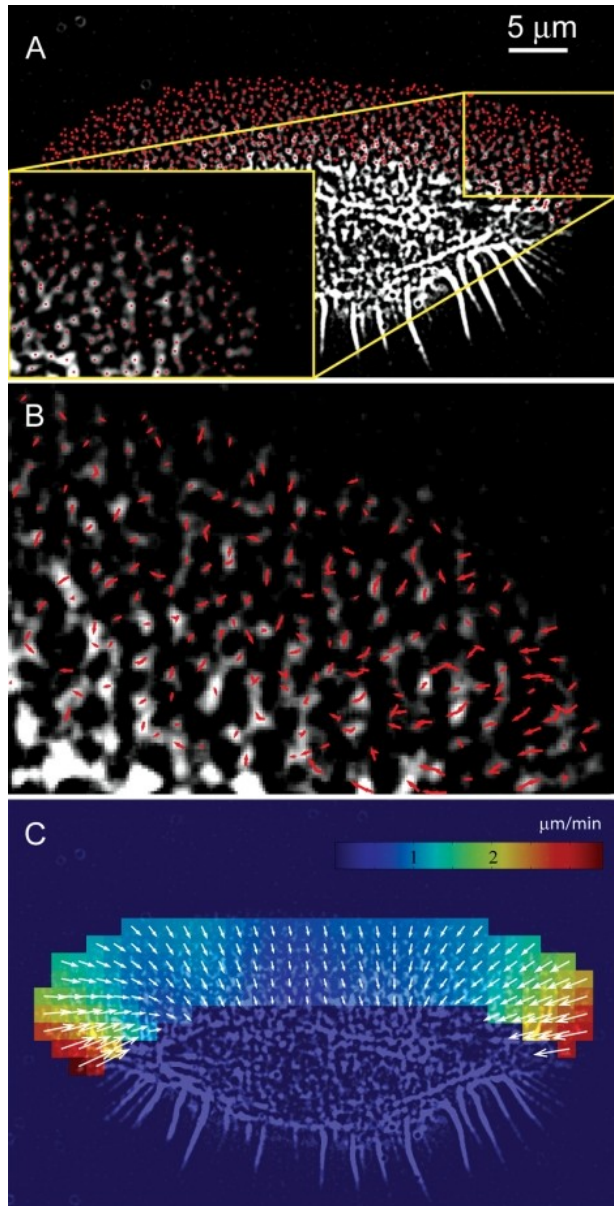
*Lamellipodium at the front*  
*only actin*  
*fast retrograde flow*

*Lamella at the back*  
*actin and myosin*  
*slow retrograde flow*

*Advancing velocity 10  $\mu\text{m}/\text{min}$ .*



# *Keratocyte motion*



*Velocity field obtained by speckle microscopy Vallotton et al.*

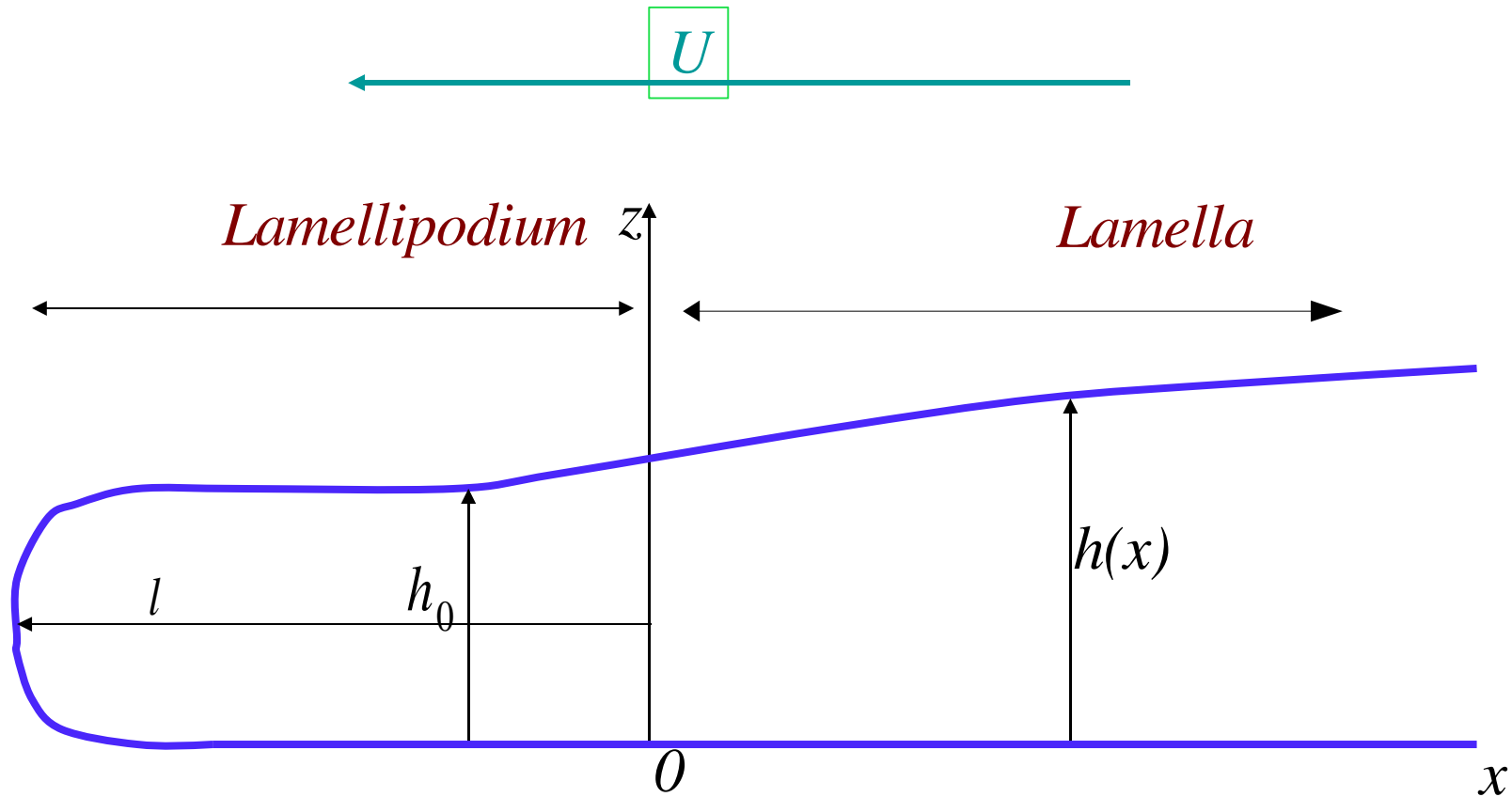
*Advancing velocity 10 μm/min*

*Retrograde flow 1 μm/min*

*Surface stress distribution  $\sigma_{xz} = 4 \cdot 10^2$  N/m<sup>2</sup> Oliver et al.*

*Actin viscosity  $\eta = E\tau \sim 10^5$  Pa.s Käs et al.*

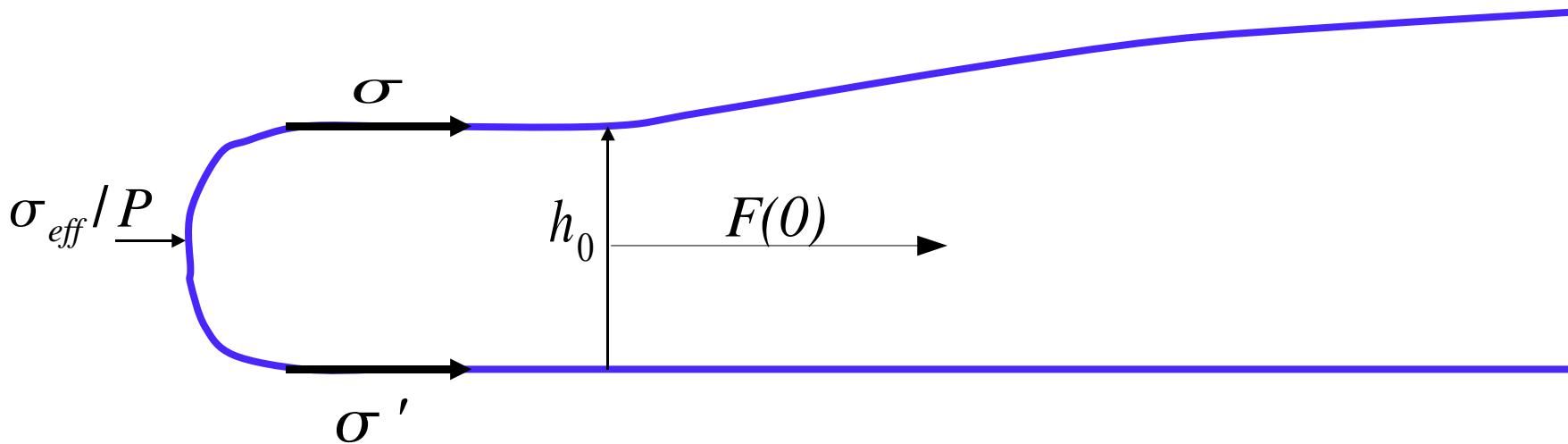
# Lamellipodium motion



*Actin viscoelasticity: solid-like region at the front (lamellipodium)  
liquid-like region at the back (lamella)*

*Size of lamellipodium  $l = (U + v(0)) \tau$*

## *Solid like motion of lamellipodium*



*Force balance on the tip (moving frame)*

*Effective polymerization pressure*

$$P_{eff} = \frac{v_p^0 - (U + v(0))}{\lambda_a}$$

*Force balance on the lamellipodium*

$$F(0) = \sigma + \sigma' + F_{ext} - \xi v(0)l$$

*Force balance on the membrane*

$$P_{eff} h_0 = \sigma + \sigma' + F_{ext}$$

# *Liquid like motion of the lamella*

## *Active gel theory*

*Constitutive equation*  $2\eta \frac{\partial v}{\partial x} = \sigma_{xx} + \zeta \Delta \mu$

*Viscous friction on the substrate*  $\sigma_{xz}(z=0) = \xi v$

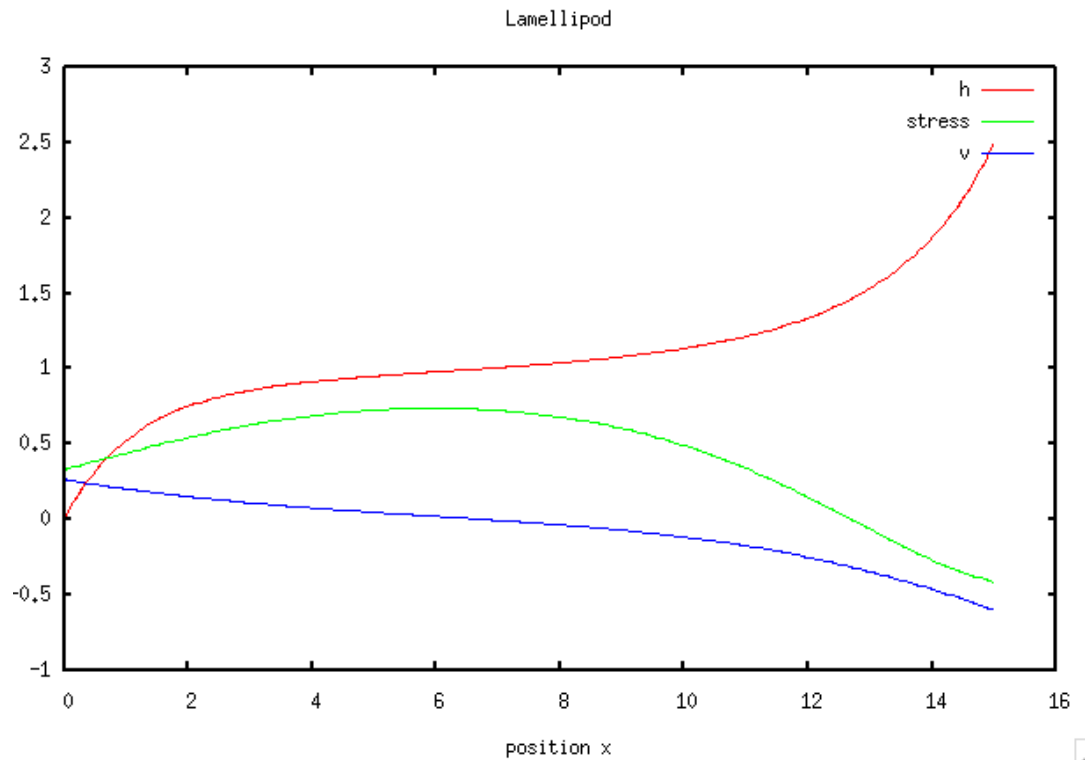
*Force balance*  $\frac{\partial(\sigma_{xx} h)}{\partial x} = \frac{\partial F}{\partial x} = \xi v$

*Mass conservation*  $h(x)(U + v(x)) = cst$

## *Matching with the solid region*

$$h_0, v(0), -F(0) = \sigma_{xx} h_0$$

# Lamellipodium profile



*Lamellipodium thickness*  $h = h_0 \frac{v_d}{u}$

*Advancing velocity*  $u = v_d - \left( \frac{h_0}{4 \eta \xi} \right)^{1/2} \zeta \Delta \mu$  *active effects 10%*

*Force dipole due to active stresses*  $Q = \zeta \Delta \mu L_x L_y h \sim -6 \cdot 10^{-13} \text{ J}$

## *Concluding remarks*

*Kachar's law under the assumption that depolymerization is controlled by a capping protein*

*Qualitative shape of the stereocilium reproduced*

*Immersed length of the stereocilium*

*One parameter undetermined: radius of stereocilium*

*Mean field theory: polymerization fluctuations at the tip*

*Role of molecular motors: myosin XV*

*myosin VI*

*Application to lamellipodium motion*