

Fish growth and degree-days: Advice for selecting base temperatures in both within- and among-lake studies.

A Thesis  
SUBMITTED TO THE FACULTY OF  
UNIVERSITY OF MINNESOTA  
BY

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IN PARTIAL FULFILLMENT OF THE REQUIREMENTS  
FOR THE DEGREE OF  
MASTER OF SCIENCE

Advised by Paul A. Venturelli

August 2013

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## Acknowledgements

Isaac Newton once said, “if I have seen further, it is by standing on the shoulders of giants.” The metaphor taken to mean that new discoveries are only possible by building on the discoveries of those that have come before. I would like to extend the meaning of this metaphor and say that whatever offering this thesis has to give; it is the cumulative result of all my fortunes in life, of which I have been the recipient of many.

I would like to begin by recognizing my advisor, Dr. Paul Venturelli, who saw potential in an astray recent Bachelor of Arts graduate with shared curricular and extracurricular interests. Paul has been endlessly attentive to my needs as a student, known and unknown, and has been tremendously supportive and dedicated to my professional and intellectual development. I could not have asked for a more dependable, understanding and responsive mentor.

I would also like to thank Dr. Nigel Lester of the Ontario Ministry of Natural Resources for not only collaborating with Paul and me, but for also providing invaluable incite and direction. Nigel has taught me how to build from first principles a logical progression of ideas that lead to new understandings. I will surely take this with me in all my future endeavors. I would also like to thank my committee members, Dr. Roger Moon and Dr. Don Pereira for guiding project objectives and providing academic advice. Furthermore, I would also like to thank the University of Minnesota and the Conservation Biology Program therein for providing funding for this project and my further education.

I would like to express additional gratitude to the following at the Ontario Ministry of Natural Resources (OMNR), Minnesota Department of Natural Resources (MNDNR) and Minnesota State Climatology Office: Pete Addison, James Cross, Susan Mann, Trevor Middel (all of the OMNR); Gerry Albert, Maggie Gorsuch, Tom Heinrich, Eric Jensen, Tony Kennedy, Mike McInerny, Nick Schlessler, Al Stevens, Ben Vondra, Matt Ward, Duane Williams (all of the MNDNR) and Greg Spoden (Minnesota State Climatology Office). The data (fish, waterbody and temperature) and professional opinions provided by these people and agencies were instrumental in the development and realization of this project.

Also, thank you to all the graduate students of which I have shared these last few years. The collective experience of this group is astonishing and the knowledge and wisdom I have garnered invaluable. I will forever cherish the experiences, laughs and stories we have all shared. Specifically to those with whom I have shared an office, I would like to say thanks for all your listening ears, enjoyable conversations and constructive feedback. Marcus Beck, Will French, Hans Martin and Beth Rigby, you made living in a windowless office space with decades old furniture, suspiciously asbestos laden ceiling dividers and terrifying ventilation ‘booms’, incredible. Also to my lab mates Grace Loppnow and Jason Papenfuss, thank you for joining me in being Paul’s first group of graduate students. Grace, I wouldn’t trade our six weeks in the field for anything. Jason, I will miss talking shop, watching you mold cups and bowls out of clay and hearing you play the guitar.

Finally, I would like to thank my grandmother, Dolores Dageford, my parents, Bob and Jill Chezik, my sister Adrienne Chezik and my extended family (living and deceased), for all their love, support and encouragement. My successes are manifestations of all your selfless hard work. I am so very fortunate to have you in my life.

## **Dedication**

This thesis is dedicated to my parents, Robert J. Chezik and Jill E. Chezik. They have been with me every step of the way with endless love and support. I could not have done this without them.

## Abstract

Ectotherm growth and development are largely a function of temperature. Within the range of thermal tolerance, growth and development increases linearly over a mid-range of temperatures. Degree-days (DD; °C·days) are a method of quantifying the thermal experience of an organism over the linear range and are an increasingly popular method for describing growth and development in fish. To limit degree-day calculations to temperatures within the linear range, a lower temperature threshold ( $T_0$ ) is incorporated that defines the onset of growth. However, there is currently no convention for choosing  $T_0$  and the implications of choosing an inappropriate  $T_0$  are largely unknown. This uncertainty has resulted in a wide range of  $T_0$  values currently in use both among- and within-species. In this thesis I explore i) how  $T_0$  affects the ability of DD to explain within-population variation in fish growth, and ii) the effect of  $T_0$  on apparent growth among populations in thermally dissimilar environments. To address these objectives, I first use simulated data to determine the theoretical relationship between  $T_0$  and growth. I then demonstrate these theoretical relationships using immature length-at-age data from 8 species and 85 waterbodies in Minnesota USA and Ontario Canada. My results show that DD at low  $T_0$  are highly correlated and that these highly correlated low  $T_0$  values explain variation in growth equally well within a single population. Furthermore, growth rates among populations of thermally dissimilar habitat become increasingly dissimilar as  $T_0$  deviates from the  $T_0$  that minimizes the variation in growth rate among populations. This effect is especially apparent when the among-population range in thermal habitats is large. These findings suggest that, although  $T_0$  matters little when accounting for variation in growth within a single population, an inappropriate  $T_0$  can lead to the appearance of among-population differences in growth. When choosing  $T_0$ , I recommend estimating  $T_0$  using the Charnov and Gillooly (2003)<sup>1</sup> “10°C rule” and then rounding to the nearest  $T_0$  standard (0, 5, 10 and 15°C). Choosing a  $T_0$  standard in this way will

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<sup>1</sup> Charnov, E.L., and Gillooly, J.F. 2003. Thermal time: body size, food quality and the 10 degree C rule. *Evol. Ecol. Res.* **5**(1): 43-51.

minimize the effect of  $T_0$  error on growth bias in among-population studies. In general, standardization i) simplifies  $T_0$  identification, ii) facilitates comparative studies, and iii) promotes the use of DD in future studies.

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# Chapter 1

## **General introduction: Growth, temperature and degree-days in fish science**

Fish growth can be defined as the investment of surplus energy into the production of somatic or reproductive tissue. Although growth is often defined in these simple terms of energy transfer, as a life history trait growth may be the single most important predictor of fish survival and fitness (Jonsson and Jonsson 2011). For instance, size is positively correlated with juvenile survival (e.g., Sogard 1997), adult female egg abundance (e.g., Morita and Takashima 1998) and male reproductive success (e.g., Magnhagen and Kvarnemo 1989). These growth-dependent mechanisms act to regulate population dynamics (Lorenzen and Enberg 2001) and determine when fish recruit into a fishery. Therefore, by understanding growth, we gain insight into the “health”, quality and economic stability of our fisheries (Hubert and Quist 2010).

Fish are poikilotherms meaning that their growth, and the underlying processes therein, are predominately determined by the temperature of their environment when food is not limiting (Diana 1995). The inability of fish to maintain thermal homeostasis, as their thermal environment fluctuates, is largely a result of their relatively low metabolic rate, lack of insulation and countercurrent lamellar blood flow (Beitinger 2000). In general, fish growth is linear over a midrange of temperatures (Sharpe and DeMichele 1977) because the rates of the chemical reactions that result in metabolism are temperature dependent (Jobling 1994). In this way, fish growth and ambient temperature are inextricably linked (e.g., Weatherley and Gill 1987).

Because ambient temperature controls fish growth it is considered an ecological resource (Magnuson et al. 1979), and can be quantified using a degree-day approach. The degree-day, first introduced as thermal time by René Antoine Ferchault de Réaumur in 1735 (Rittner 2009), quantifies the thermal opportunity for growth by aggregating temperatures relevant to growth. A single degree-day (DD; °C·days) is calculated as

$$(1.1) \quad DD = \left[ \frac{T_{Max} + T_{Min}}{2} \right] - T_o$$

where  $T_{Max}$  and  $T_{Min}$  are the maximum and minimum daily ambient temperature, respectively, and  $T_o$  is the temperature below which growth or development does not occur (often referred to as the base or threshold temperature). When positive degree-day values are summed over some period of interest, the resultant cumulative degree-days act as an index of the metabolically relevant thermal energy that was experienced over that period.

The degree-day assumes growth to be linear with and solely a function of temperature (Legg 1998a). Although fish growth is non-linear at the extremes of their thermal limits (e.g., Edsall 1993), the majority of growth typically takes place in the linear region. Also, it is well understood that there are temperature independent regulators of growth (i.e., population density, genetics, pH, trophic state, etc.), and a complete growth model would include such variables. On the other hand, assuming temperature is the only contributor to growth allows degree-days to uniquely account for temperature dependent processes. In this way, unexplained variation can be attributed to factors that are independent of temperature (e.g., population density, Venturelli et al. 2010). Ultimately, neither assumption impinges on the degree-days ability to account for temperature dependent growth.

The degree-day greatly outperforms calendar time when describing concurrent patterns of growth and development because it directly measures the thermal opportunity for growth, which can vary over time and across latitude or altitude. For example, in agronomy when predicting the fifth-leaf stage of wheat, a calendar-based model is accurate within nine days whereas a degree-day model is accurate to within two days (Miller et al. 2001). Similar increases in precision have been shown in fish science. For instance, Colby and Nepzy (1981) demonstrated that variation in interspecific growth rates among walleye (*Sander vitreus*) populations was best described using degree-days rather than calendar time. In marine systems, Neuheimer and Taggart (2007) showed that length-at-age in 17 stocks of North Atlantic cod (*Gadus morhua*) was significantly

correlated with degree-days, accounting for 93% of the variation in length-at-age. Calendar time failed to detect a significant correlation in these same data.

Although the degree-day approach is superior to calendar time when describing growth, degree-days continue to be underutilized in fish science, accounting for only 5% of temperature-related growth studies between 1980 and 2006 (Neuheimer and Taggart 2007). It is unclear why fish science has been reluctant to adopt the degree-day given that the relationship between temperature and development in fish is well understood (e.g., Morrow and Mauro 1950, Paloheimo and Dickie 1966, Fry 1971, Kitchell et al. 1977) and the degree-day approach has been widely used in agronomy for 270+ years (e.g., Réaumur 1735, cited in Bonhomme 2000).

Two likely reasons why degree-days have been underutilized in fish science include, i) a general reluctance to depart from the convention of calendar time and ii) prohibitive limitations inherent in collecting water temperature data. The latter is understandable if the efficacy of the degree-day were dependent on precisely measuring the thermal experience of fish. If so, not only would temperature need to be recorded at multiple depths but also in several locations in order to account for within lake variability (Stevens et al. 1978). Furthermore, because fish exhibit behavioral thermo-regulation (Neill and Magnuson 1974), individual locality data would also be essential. These limitations in water temperature may explain why early degree-day studies were restricted primarily to egg development and aquaculture (Wallich 1901), where temperature is easily monitored and controlled.

It is now understood that air temperature is highly correlated with surface water temperature and acts as a suitable surrogate when water temperature data are incomplete or absent (e.g., Kumar et al. 2013, Livingstone and Lotter 1998, Macan and Maudsley 1966). The availability of air temperature data allows for and has resulted in comparative growth studies among populations and over large areas (e.g., Rypel 2012a,b). A review of the literature shows that while there is a general trending increase in growth studies using degree-days, comparative studies are becoming more common and almost exclusively use air temperature data (Fig. 1.1). At the same time, within population studies are not increasing as rapidly and continue to rely heavily on water temperature data. Although

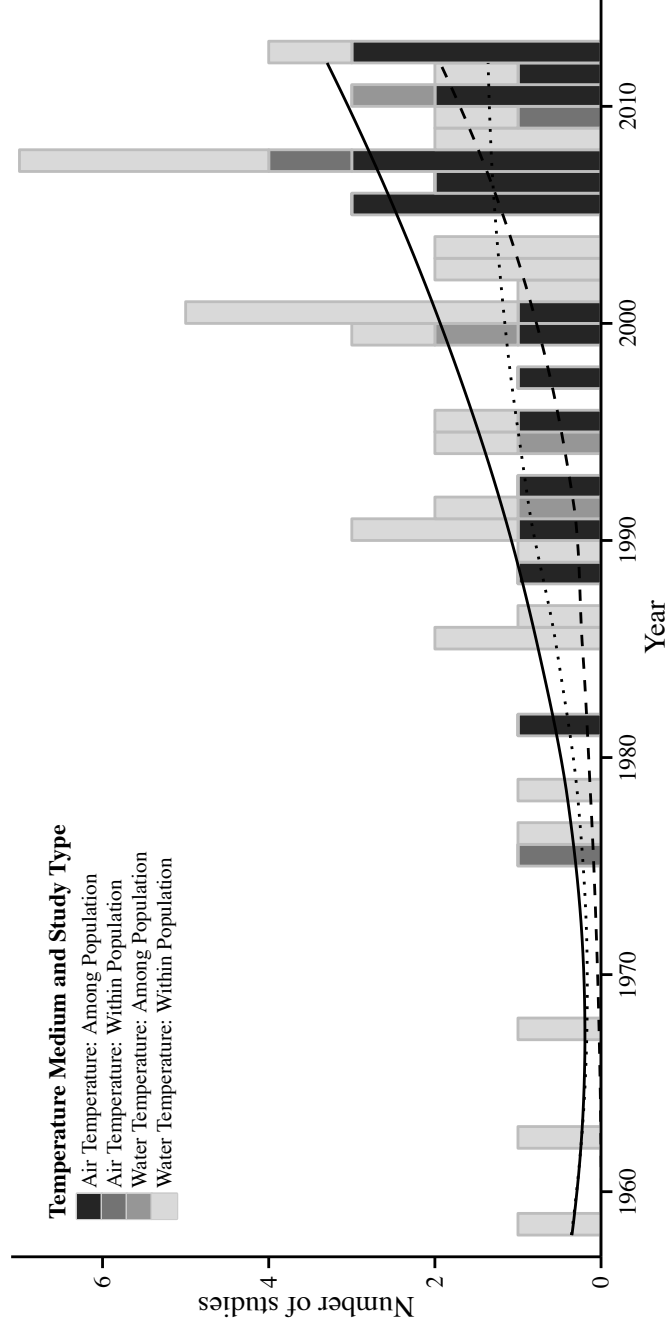
there is no evidence suggesting the efficacy of water temperature to be greater than that of air, this dichotomy in study type suggests water temperature is preferred when available while air temperature is an acceptable alternative. Between the adoption of air temperature as a water temperature surrogate and water temperature records becoming more complete, the degree-day is becoming much more popular. Given its utility and efficacy it is likely to become a standard tool for describing fish growth and development.

Although the degree-day is becoming more widely used in fish science, little has been done to understand its shortcomings and pitfalls, nor have standards or methods been developed to maximize its utility. For instance, it is unclear what temperatures act as lower and upper bounds to growth and whether the precision of these values affect the degree-days efficacy. Furthermore, although degree-days and annual immature growth exhibit a linear relationship, this has yet to be shown for within season immature growth. Simple questions of whether air temperature is truly a good surrogate for water temperature or whether degree-days estimate the growth of warm-water and cool-water species equally well have yet to be understood. By taking the time now, during the degree-day's infancy in fish science, we can avoid developing unsubstantiated conventions that leave datasets incompatible, results inaccurate, and conclusions incorrect.

My thesis takes the first steps in defining the bounds to describing fish growth when using degree-days. It does this by looking critically at the effect of  $T_0$  when accounting for variation in growth and when comparing growth among populations. Currently there is no common method in use for defining  $T_0$ , rather these values are often chosen with either no justification or are generalized to the minimum temperature for a given physiological process. In entomology, several methods have been developed to empirically estimate  $T_0$  values (Arnold 1959), but these have received limited use in fish science. This is concerning because aggregating degree-days before or after the initiation of growth results in an inaccurate description of growth-rate. Minimizing the statistical noise that results from our methods enhances our ability to detect real differences over time or among populations and limits type II errors (i.e., false positives). In chapter 2, I

explore how the description of the growing season changes with  $T_0$ , and the effect of  $T_0$  on explained variation in growth. Chapter 3 builds on the previous chapter by understanding the effect of  $T_0$  on apparent growth rates among lakes over a thermal gradient. In both chapters simple conceptual models are used to develop clear predictions and hypotheses, which are then tested using empirical length-at-age data aggregated from databases in Minnesota and Ontario.

Chapters 2 and 3 have been prepared as manuscripts for publication in the Canadian Journal of Fisheries and Aquatic Sciences (CJFAS). At the time of this writing, Chapter 2 had been accepted with minor revisions but those revisions had yet to be submitted and therefore will be referred to as Chezik et al. (*in review*). Publication of these chapters by the University of Minnesota is allowed under the ‘author rights’ provided by CJFAS. These rights state, “authors may reuse all or part of their manuscript in other works created by them for noncommercial purposes, provided the original publication in an NRC Research Press journal is acknowledged through a note or citation”. Although Chapter 2 is not yet published the DOI for this paper will be 10.1138/cjfas-2013-0295 and can be found at the CJFAS website upon acceptance. Co-authors on these manuscripts include Nigel P. Lester and Paul A. Venturelli. I will use the plural pronouns “we” or “our” instead of the singular “I” or “my” in reference to co-authorship.



**Figure 1.1** The frequency of freshwater studies over time using degree-days to i) account for growth within a single population or localized region and ii) compare growth among thermally disparate populations, using either water or air temperature. Overall, the number of growth studies using degree-days is increasing (-) along with the use of air temperature as a surrogate for water temperature (--), recently surpassing water temperature based studies (•••). All trends described by a loess smoother ( $\alpha = 1$ ).

## Chapter 2

### **Fish growth and degree-days I: Selecting a base temperature for a single population study.**

#### **Synopsis**

Degree-days (DD) are an increasingly popular method for explaining variation in fish growth and development. By including a base temperature ( $T_0$ ) the DD formula limits calculations to temperatures that are relevant to growth. However, a review of growth studies shows multiple  $T_0$  values in use for a given fish species. To determine how  $T_0$  affects the ability of DD to explain within-population growth variation, we first show that the ability of DD to describe a growing season is robust to low values of  $T_0$ . We then analyzed immature length data from 8 species and 85 waterbodies in North America to show that there is a broad range of  $T_0$  values that effectively explain growth variation. Based on these results, we argue that precise  $T_0$  estimates are unwarranted for most single-population studies, and recommend standard  $T_0$  values (0, 5, 10, 15°C). Standardization facilitates comparative studies and promotes the use of DD in future research. To this end, we provide equations for converting annual DD at a given  $T_0$  to annual DD at a standard  $T_0$ .

## 2.1 Introduction

The metabolic rate of fish and other ectotherms is largely determined by the ambient temperature of their environment (Hazel and Prosser 1974) and increases linearly over a midrange of temperatures (Sharpe and DeMichele 1977). The amount of mid-range ambient thermal energy that an ectotherm has experienced can be quantified using a degree-day approach (also known as thermal time, e.g., Trudgill et al. 2005). The degree-day for a single day (DD; °C·days) is calculated as

$$(2.1) \quad DD = \left[ \frac{T_{Max} + T_{Min}}{2} \right] - T_o,$$

where  $T_{Max}$  and  $T_{Min}$  are the maximum and minimum daily ambient temperature respectively and  $T_o$  (often referred to as the base or threshold temperature) is the temperature below which growth or development is non-linear and effectively zero.

When non-negative DD values are summed over some period of interest, the resultant cumulative degree-days at a given  $T_o$  ( $CDD_{T_o}$ ; °C·days) is an index of the metabolically relevant thermal energy that was experienced over that period. This index is useful for describing concurrent patterns of growth and development (e.g., Neuheimer et al. 2012, Venturelli et al. 2010).

Although degree-days have been used for decades to describe growth and development in plants (e.g., Réaumur 1735, cited in Bonhomme 2000) and insects (e.g., Seamster 1950, Thorup 1963), this approach is rare in fish science, accounting for only 5% of all temperature-related growth studies between 1980 and 2006 (Neuheimer and Taggart 2007). Relationships between temperature and development in fish are well understood (e.g., Morrow and Mauro 1950, Paloheimo and Dickie 1966, Fry 1971, Kitchell et al. 1977), but for decades the degree-day approach was limited to egg development (e.g., Wallich 1901). Degree-days were first applied to growth by Le Cren (1958), but it wasn't until the late 1970's that this application began to increase in popularity (Fig. 1.1). This increase appears to have been facilitated by the use of air temperatures as a surrogate for surface water temperatures (e.g., Ney and Smith 1975, Colby and Nepszy 1981). Air temperatures are highly correlated with surface water temperatures (e.g., Macan and Maudsley 1966, Reuter et al. 2009) and air temperature

records are typically more extensive and complete. Therefore, air temperatures permit growth studies over large spatial and temporal scales (e.g., Purchase et al. 2005, Wagner et al. 2007, Latta et al. 2008) and when local time series of water temperature are incomplete or unavailable (e.g., Rennie et al. 2009, David 2012, Rypel 2012a,b).

The degree-day approach to describing fish growth is increasing in popularity, but a review of the freshwater literature shows a diversity of methods for estimating  $T_0$  and a diversity of  $T_0$  values in use for a given species (Table 2.1). More often than not,  $T_0$  values are used without justification. When justification is provided, it is usually that  $T_0$  was i) based on the minimum temperature for some physiological process (growth, feeding, spawning, etc.), ii) estimated via model fitting (e.g. bioenergetics, regression), or iii) taken from other (sometimes unrelated) fishes or even plants (Table 2.1). Among freshwater species, reported values of  $T_0$  range from  $-3.6^{\circ}\text{C}$  for smelt (*Osmerus eperlanus*) to  $18^{\circ}\text{C}$  for spotted gar (*Lepisosteus oculatus*). Within species,  $T_0$  can range from  $0$  to  $14^{\circ}\text{C}$  (yellow perch, *Perca flavescens*; European perch, *P. fluviatilis*) and take as many as 7 different values (roach, *Rutilus rutilus*; Table 2.1). This diversity appears to be due, in part, to a tendency to estimate  $T_0$  anew rather than to adopt an existing estimate (Table 2.1).

There are at least four potential reasons to be concerned about the inconsistency with which  $T_0$  values for fish growth are being estimated (or simply adopted). First, different approaches to estimating or justifying  $T_0$  are likely to lead to different values, even for the same population. For example,  $T_0$  for roach in a Dutch lake was  $11.5^{\circ}\text{C}$  in the lab,  $10.2^{\circ}\text{C}$  when fitting a growth model to field data, and  $17.5^{\circ}\text{C}$  when using this model to predict growth in the field (Table 2.1). Second, the same estimation method can yield different  $T_0$  values for different populations (e.g., goodness of fit applied to roach; Table 2.1), which suggests that we must estimate  $T_0$  for every population. Third, different  $T_0$  values can lead to differences in the ability of degree-days to explain or predict growth. These differences arise because  $T_0$  determines the extent to which degree-days integrate thermal energy that is relevant to growth. This point argues for the adoption of a single, robust estimation method across studies, and cautions against adopting  $T_0$  values from different species. Finally, variation in  $T_0$  hinders the comparison and interpretation

of degree-days and growth rates among populations and studies. For example, whereas Abbey and Mackay (1991) reported a yellow perch growth rate of 0.02 mm/°C·Day using  $T_0$ , Power and van den Heuvel (1999) reported 0.09 mm/°C·Day when using  $T_{13.5}$ .

In this study we address potential concerns over variation in  $T_0$  among within-lake studies of degree-days and growth. Specifically, we use simulated and empirical data to i) determine how variation in  $T_0$  affects the ability of degree-days to account for within-lake variation in immature growth, and ii) identify the scope for standardized  $T_0$  values in future within-lake growth studies. Our interest is in  $T_0$  from an applied rather than a theoretical perspective. In theory, there is a precise, species- or population-specific temperature below which growth rate is effectively zero (Charnov and Gillooly 2003) and it is therefore appropriate to set  $T_0$  to this value. However, if the ability of degree-days to explain growth is robust to  $T_0$ , then, from an applied perspective, it is appropriate to abandon precise  $T_0$  estimates in favour of a standardized value or set of values.

## 2.2 The effect of $T_0$ on degree-days

A logical first step in determining how the degree-days ability to explain growth changes with  $T_0$  is to determine how degree-days themselves change with  $T_0$ . To this end, we assumed that annual temperature cycles are well approximated by a sine curve (Arnold 1959, Shuter et al. 1983). Annual degree-days above some  $T_0$  ( $ADD_{T_0}$ ; °C·days) are then given by the area under this curve (Baskerville and Emin 1969):

$$(2.2) \quad ADD_{T_0} = \frac{365}{\pi} \left[ w \int_{\theta}^{\pi} \sin t \, dt - \int_{\theta}^{\pi} (T_0 - T_{Mean}) dt \right],$$

where  $t$  is day in year, 365 is the cycle period,  $w = \frac{T_{Max} - T_{Min}}{2}$ ,  $T_{Mean}$  is mean annual temperature and  $\theta = \frac{T_0 - T_{Mean}}{w}$  (Fig. 2.1a). To determine the effect of  $T_0$  on  $ADD_{T_0}$ , we used equation 2.2 to calculate  $ADD_{T_0}$  for all integer values of  $T_0$  in the range 0 to 30°C, and for four values of  $w$  (15, 20, 25 and 30 °C). Increasing the threshold ( $T_0$ ) decreased the summable area under the curve (i.e.,  $ADD_{T_0}$ ) and increasing  $w$  increased available thermal energy (i.e., decreased latitude). For each  $w$ , we then expressed each value of  $ADD_{T_0}$  as a proportion of  $ADD_{T_0-1}$  (Fig. 2.1d). Results suggest that  $ADD_{T_0} \approx ADD_{T_0-1}$  for a range of  $T_0$  values  $\geq 1^\circ\text{C}$ , and that the extent of this plateau increases with  $w$  (Fig. 2.1d).

In other words, a 1°C change at low  $T_o$  results in relatively similar  $ADD_{T_o}$  values while a 1°C change at high  $T_o$  results in relatively different  $ADD_{T_o}$  values. We observed a similar pattern in both air (Fig. 2.1b and Fig. 2.1e) and water (Fig. 2.1c and Fig. 2.1f) temperature data from the Gulf Coast to Alaska. Overall, these results suggest that our ability to describe a growing season using degree-days is largely robust to  $T_o$  regardless of medium, provided that  $T_o$  is not too large (or  $w$  is not too small).

### **2.3 The effect of $T_o$ on explained variation in growth**

If our ability to describe a growing season using degree-days is robust to  $T_o$ , then our ability to use degree-days to explain growth within a population should also be robust to  $T_o$ . To test this hypothesis, we compiled fish and temperature data from Ontario Canada and Minnesota USA, and regressed cohort length at age of each population against cumulative degree-days for a range of  $T_o$  values. We used the coefficient of determination ( $r^2$ ) of each length|degree-day regression as a measure of the effect of  $T_o$  on explained variation in species-specific growth within a waterbody. This method has been developed independently by Baras and Philippart (1999) and Neuheimer and Taggart (2007), and was applied to a population of walleye by Venturelli et al. (2010). We used the goodness of fit ( $r^2$ ) approach and empirical rather than simulated growth data to test our hypothesis because i) the  $r^2$  approach allowed us to both identify and evaluate the performance of each  $T_o$  value, ii) empirical growth data integrate both direct (i.e., metabolic) and indirect (i.e., productivity-based) temperature effects, and iii) we are ultimately interested in the ability of degree-days to explain growth in natural environments.

#### **2.3.1 Methods**

##### *i) Fish data*

We created a database of individual fish records that combined the Ontario Ministry of Natural Resources (OMNR) fall walleye index netting (FWIN) database (1993-2008) with the Minnesota Department of Natural Resources (MNDNR) statewide database (2001-2011) and supplemental data (2001-2011) from Minnesota's ten large lakes (Winnibigoshish, Rainy, Vermillion, Leech, Cass, Pepin, Mille Lacs, Kabetogama, Lake

of the Woods, and Upper Red Lake). Minnesota data were collected between March and December using a variety of methods (e.g., electrofishing, gillnetting, trap-netting, trawling, trot-lining, seining, angling). Ontario data were collected between September and November using multi-mesh gillnets set overnight at multiple depths (Morgan 2002).

For each fish we were interested in the following information: waterbody, sample date (day, month and year), species, age (years), total length (L; mm), maturity and aging structure. We focused on immature fish so that our analyses of somatic growth were not confounded by the allocation of surplus energy to reproduction. This approach can create a small bias in the growth trajectory if it removes the fast-growing (i.e., early-maturing) fish from the oldest cohort(s). We also focused on fish that were aged using a reliable aging structure or sampled at the start or end of the growing season. We avoided unreliable aging structures because the accuracy of the degree-day assignment depended on the accuracy of age estimates (see next section). We assigned reliability to different aging structures using a combination of expert opinion (Mike McInerny MNDNR 23070 N. Lakeshore Dr. Glenwood, MN 55041, Susan Mann OMNR 479 Government Rd. PO Box 730 Dryden ON P8N2Z4) and the literature (Campbell and Babaluk 1979, Graeb and Willis 2010, Schneider 2001, Soupier et al. 1997, Trippel and Harvey 1991). For example, we assumed that only age-0 fish were accurately aged in the field, and that only walleye  $\leq$ age-5 were accurately aged using scales. We also used expert opinion (MNDNR) to define the growing season as June through August, inclusive. We avoided records that were sampled during this period because there is currently no validated protocol for combining whole- and partial-season degree-days in a single analysis.

Our final database contained 47,486 fish records representing 8 species from 64 waterbodies in Ontario Canada and 21 waterbodies in Minnesota USA (Table 2.2). Walleye comprised the majority of the database (86% of all fish) followed by other percoid species (14%).

#### ii) *Temperature data*

We assigned  $CDD_{T_0}$  values to each fish by summing the annual degree-days that each fish experienced prior to capture. This approach required time series estimates of  $ADD_{T_0}$  for each waterbody. For waterbodies in Minnesota, we calculated  $ADD_{T_0}$  time series

(2001-2011) using 100 km<sup>2</sup> resolution maps of kriging-interpolated air temperature data ( $T_{\text{Min}}$  and  $T_{\text{Max}}$ ). These data were collected from automated weather stations owned and operated by the National Weather Service, and assembled and processed by the Minnesota State Climatology Office-MNDNR. We calculated  $\text{ADD}_{T_0}$  by applying equation 2.1 to daily  $T_{\text{Min}}$  and  $T_{\text{Max}}$  values from the grid point closest to the centroid of the waterbody of interest, and then summing across non-negative DD values in that year. Because our objective was to determine the effect of  $T_0$  on explained variation in growth, we repeated these degree-day calculations for each integer value of  $T_0$  in the range 0-30°C (the range of water temperatures that a temperate, freshwater fish is likely to experience in a given growing season). Thus, we generated a unique time series of  $\text{ADD}_{T_0}$  values for all waterbodies and  $T_0$  values. For waterbodies in Ontario, we obtained  $\text{ADD}_{T_0}$  time series (1981-2008) using the Historical Climate Analysis Tool (Cross et al. 2012), which automatically applied the above protocol to 1 km<sup>2</sup> resolution maps of kriging-interpolated  $T_{\text{Min}}$  and  $T_{\text{Max}}$  data.

We used age, together with capture year and month, to determine the  $\text{CDD}_{T_0}$  that each fish experienced prior to capture. Because fish are aged to a January 1 birthday (Holden and Raitt 1974), we used different summation rules for different capture months. For fish captured during the period January to May (i.e., after their common birthday but before the start of a growing season), we summed  $\text{ADD}_{T_0}$  across the previous  $n$  growing seasons, where  $n$  is the age of the fish in years. Thus, if a 2 year-old fish was captured in January 2005, we calculated  $\text{CDD}_{T_0}$  as the sum of  $\text{ADD}_{T_0}$  values from 2003 and 2004. For fish captured during the period September to December (i.e., before their common birthday but after the previous growing season), we calculated  $\text{CDD}_{T_0}$  by summing  $\text{ADD}_{T_0}$  across the previous  $n+1$  growing seasons, beginning with the current year of capture. Thus, if a 2 year-old fish was captured in September 2005, we summed  $\text{ADD}_{T_0}$  values from 2003 to 2005.

### iii) *Statistical analyses*

Our analyses included populations for which we had  $\geq 5$  cohorts distributed over  $\geq 3$  age classes that each contained  $\geq 5$  individuals (sexes combined). We set these minima to ensure that growth was accurately described and not unduly influenced by outliers. For

each population, we calculated mean length-at-age for each cohort. Here ‘cohort’ refers to an age-class observed during a sampling event rather than a group of fish that hatched in the same year. This means if we had multiple years of data for a given population, a single age class typically had several mean length estimates (e.g., Fig 2.2). We assumed the length ( $L$ ) of immature fish could be described by a linear model

$$(2.3) \quad L = m \cdot CDD_{T_o} + b$$

where  $CDD_{T_o}$  is the thermal age for a given  $T_o$ . We used ordinary least squares to estimate parameters  $m$  and  $b$  and the coefficient of determination ( $r^2$ ) for a range of  $T_o$  values (0 to 30°C) – see example in Fig. 2.2. We then plotted  $r^2$  against  $T_o$  to examine how goodness of fit varied with temperature threshold.

Statistically speaking, the relative goodness of fit of different models should be evaluated using AIC because  $r^2$  is i) an inappropriate method for comparing models when the range of the explanatory variable varies among models (which was the case in our study), and ii) biased towards models with large ranges in the explanatory variable (Maindonald & Braun 2007). However, whereas both approaches produced qualitatively (and almost quantitatively) identical results,  $r^2$  also quantifies the strength of the relationship and there is precedence for this approach (Baras and Philippart 1999, Neuheimer and Taggart 2007, Venturelli et al. 2010).

### 2.3.2 Results

Across species and waterbodies (Fig. 2.3), the  $r^2$  method revealed a wide range of  $T_o$  values that were effective at describing growth. For all eight species in our database,  $r^2$  values were similar for small values of  $T_o$ . Beyond this plateau  $r^2$  dropped off quickly or became erratic. Erratic behaviour at high  $T_o$  was common for species or waterbodies that had low sample sizes (i.e., few cohorts or fish per cohort). This plateau in  $r^2$  was evident in all 81 walleye populations in our analysis and had a median value of 14°C (range 6-21°C) (Fig. 2.4). Although Fig. 2.1 suggests that the length of the  $r^2$  plateau increases with the length of the growing season and  $w$  in equation 2.2, we found no evidence in walleye that  $r^2$  increased over the mean  $ADD_0$  range (2157-3733°C·days) (df=79,

$r^2=0.04$ ,  $p\text{-value}=0.062$ ). Visual inspection of each regression and associated diagnostic plots indicated the assumptions of the linear model were met.

## 2.4 Discussion

In the Introduction, we described four concerns over the inconsistency with which  $T_0$  values are being estimated (or adopted) in within-lake growth studies: i) different approaches applied to the same population are likely to lead to different  $T_0$  values, ii) identical approaches applied to different populations are likely to lead to different  $T_0$  values, iii) different  $T_0$  values can lead to differences in the ability of degree-days to explain or predict growth, and iv) variation in  $T_0$  hinders the comparison and interpretation of degree-days and growth rates among populations and studies. Our results suggest that, from an applied perspective, concerns i-iii are largely unfounded; within a given waterbody, there is a broad range of  $T_0$  values that are almost equally effective at explaining variation in growth. This pattern was consistent across the 8 freshwater species (Fig. 2.3) and among the 81 walleye populations in this study (Fig. 2.4). It was also apparent in 21 other freshwater and marine species (Baras and Phillippart 1999, supplemental material for Neuheimer and Taggart 2007). This broad range of effective  $T_0$  values has little to do with limitations on growth at temperatures approaching  $0^\circ\text{C}$ . Rather, it stems from an inherent property of the degree-day approach, namely that  $\text{ADD}_{T_0}$  values become increasingly similar as  $T_0$  approaches  $0^\circ\text{C}$  (Fig. 2.1). These results suggest that there is little benefit in precisely estimating  $T_0$  when describing growth within a waterbody. Indeed, given that  $T_0$  estimates are likely to vary across methods, populations, and species (Table 2.1), increased precision is costly in that it complicates comparisons across populations and studies (concern iv).

The lack of a need to precisely estimate  $T_0$  for a given waterbody and the benefit of being able to compare results across populations and studies provides ample scope for adopting standard  $T_0$  values when describing fish growth. To this end, we recommend standardized  $T_0$  values of 0, 5, 10, and  $15^\circ\text{C}$  for growth studies involving freshwater fishes in temperate and arctic waterbodies. These standards are widely used in agronomy and entomology (Pruess 1983) and are common in engineering (e.g., heating and cooling

degree-days). As such, air temperature-based degree-day data for each of these standards are widely available. These standards should apply equally well to marine fishes (at least in temperate and arctic waters; supplemental material for Neuheimer and Taggart 2007) and processes other than growth (e.g., maturation) because the correlation between  $ADD_{T_0}$  is an inherent property of the degree-day.

Standardization does not imply that a single threshold model is the best way to account for variation in growth. The bioenergetics literature clearly shows that growth in fish is a complex function of temperature rather than the simple step function that is assumed by the degree-day approach: growth rate typically increases as temperature approaches an optimal value and then declines as temperature exceeds the optimum (e.g., Kitchell et al 1977, Hanson et al. 1997 – Wisconsin Bioenergetics model). Although a single threshold model can approximate the rise in growth with temperature, it does not account for a reduction at high temperature. Thus, the single threshold degree-day approach assumes that more heat is always better. However, standardization is appropriate for the degree-day approach because there is a range of  $T_0$  values that effectively explain growth in a given species, and these ranges of values overlap considerably among species (Fig. 2.3). We provide multiple standardized values so as to accommodate the disparate thermal preferences among species (e.g. cold-, cool-, and warm-water species; Magnuson et al. 1979) and simplify the task of assigning a single  $T_0$  to a given species or community.

To facilitate the use of 0, 5, 10, and 15°C as  $T_0$  standards in fish science, we used air-based annual degree-day data from 303 waterbodies in Ontario for the period 1981-2008 ( $n = 8484$  waterbody years) to derive the following linear conversion equation:

$$(2.4) \quad ADD_{T_{0,y}} = i + s \cdot ADD_{T_{0,x}}$$

$$(2.5) \text{ where } i = a_4 \cdot T_{0,x} + a_5,$$

$$(2.6) \quad s = \begin{cases} a_1 \cdot T_{0,x} + a_2, & T_{0,y} = 0 \\ \frac{1}{a_1 + a_2 \cdot T_{0,x}^{a_3}}, & T_{0,y} > 0 \end{cases} ,$$

$ADD_{T_{0,x}}$  is observed annual degree-days,  $T_{0,x}$  is the  $T_0$  value being converted from,

$ADD_{T_{0,y}}$  is predicted annual degree-days at the  $T_{0,y}$  standard of interest (0, 5, 10, or 15°C),

and  $a_1$ -  $a_5$  are constants (Table 2.3). Because we estimated parameter values using  $ADD_{T_{o,y}}|ADD_{T_{o,x}}$  relationships for which  $r^2$  was  $\geq 0.90$ , each standard  $T_{o,y}$  model has an  $\widehat{ADD}_{T_{o,y}}|ADD_{T_{o,y}}$   $r^2$  of  $0.91 \pm 0.006$  provided that  $T_{o,x}$  and  $ADD_{T_{o,y}}$  are within prescribed range limits (see Table 2.3). Without further study we recommend against using equation 2.4 to convert among degree-days based on water temperatures. Although air and water temperatures are both highly correlated at a given location and well approximated by a sine curve (Fig. 2.1), these sine curves and the  $ADD_{T_o}$  values that result from them are different because annual air and water profiles do not share the same  $T_{Mean}$ ,  $w$  or cycle period.

The argument for adopting standardized base temperatures when applying degree-days to fish growth is essentially this: if one is interested in using degree-days to describe growth, and the ability of degree-days to describe growth is robust to  $T_o$  (as we have shown), then the benefits of standardizing  $T_o$  far outweigh the costs. We acknowledge that standardization can result in a marginal reduction in explanatory and predictive power and will certainly preclude certain types of analyses (e.g., among-species comparisons of the extent to which degree-days explain growth). However, standardization eliminates the need to identify  $T_o$  values for each population and study, facilitates comparative studies, and promotes the use of degree-days in future research. When the use of a standard  $T_o$  value is untenable, our advice is to follow published precedent. Following precedent for identifying  $T_o$  values relevant to growth should ensure a modicum of standardization. If  $T_o$  must be estimated with precision then we recommend setting  $T_o$  to the minimum temperature for growth. Methods for estimating  $T_o$  in this way are described in relevant references in Table 2.1; see also Charnov and Gillooly (2003), Gagné and Rodríguez (2008) and Legg et al. (1998a).

In this study, we showed that the ability to describe fish growth in a single population is robust to  $T_o$  and that there is scope for adopting standard  $T_o$  values that are already in use in agronomy and entomology. However, it remains to be tested whether these results hold when describing fish growth in multiple populations or over large spatial scales (e.g., marine stocks). Until this work is complete, we caution that precise  $T_o$

might be necessary in certain studies. We also encourage research to determine the extent to which our results hold at more southern latitudes, and under what circumstances we should include thermal maxima in degree-day calculations. Similarly, further research is needed to determine why cisco growth in Leech Lake was best explained using a  $T_o$  of  $\sim 20^\circ\text{C}$  (Fig. 2.3b). Given that cisco exhibit optimal growth at  $18.1^\circ\text{C}$  and prefer  $8.7$  to  $16.1^\circ\text{C}$  (Hasnain 2013), this result is likely a statistical anomaly stemming from a relatively small sample size. Regardless, it appears that growth in this population is relatively poorly described by air temperature derived degree-days, and we wonder if this trend is evident in other populations and cold-water species. In general, there is also a need to develop protocols for combining whole- and partial-season degree-days in a single analysis, and for converting among  $T_o$  values when degree-days are based on water temperature. Clearly there is still much to learn about the use of degree-days in fish science and we encourage more research to address these and other questions.

**Table 2.1** A summary of published  $T_0$  values used in growth studies of freshwater fishes within a single waterbody or localized region. Adoption frequency refers to the number of times that an estimated  $T_0$  value was adopted in later studies involving the same species.

| scientific name               | common name         | age(s) <sup>a</sup> | $T_0$ (°C)     | temperature <sup>b</sup> | justification <sup>c</sup> | reference                                     | adoption frequency |
|-------------------------------|---------------------|---------------------|----------------|--------------------------|----------------------------|---|--------------------|
| <i>Abramis brama</i>          | common bream        | 0                   | 12.8           | W                        | 1                          | Mooij et al. (1994)                           | 2                  |
| <i>Aplodinotus grunniens</i>  | freshwater drum     | 1-10                | 9.0            | W                        | 2                          | Braaten and Guy (2002)                        | 0                  |
| <i>Barbus barbus</i>          | barbel              | 0                   | 13.5           | W                        | 3                          | Baras and Phiippart (1999)                    | 0                  |
| <i>Carassius carassius</i>    | crucian carp        | 0-2                 | 12.0           | W                        | 4                          | Copp et al. (2008)                            | 1                  |
| <i>Carpionides carpio</i>     | river carpsucker    | 1-8                 | 10             | W                        | 2                          | Braaten and Guy (2002)                        | 0                  |
| <i>Coregonus clupeaformis</i> | lake whitefish      | 2-3                 | 5              | A                        | 9                          | Rennie et al. (2009)                          | 0                  |
| <i>Esox lucius</i>            | northern pike       | juvenile<br>unknown | 7<br>10        | W<br>W                   | 2<br>5                     | Bry et al. (1991)<br>Frost and Kipling (1967) | 0<br>3             |
| <i>Gobio gobio</i>            | gudgeon             | 0                   | 12             | W                        | 9                          | Cowx (2000)                                   | 0                  |
| <i>Gymnocephalus cernua</i>   | ruffe               | 0                   | 7.6            | W                        | 1                          | Mooij et al. (1994)                           | 0                  |
| <i>Lepomis gibbosus</i>       | pumpkinseed sunfish | 0                   | 14             | W                        | 2                          | Murphy et al. (2012)                          | 0                  |
| <i>Lepomis macrochirus</i>    | bluegill sunfish    | 0                   | 14             | W                        | 2                          | Murphy et al. (2011)                          | 0                  |
| <i>Leuciscus leuciscus</i>    | common dace         | 0                   | 12             | W                        | 9                          | Cowx (2000)                                   | 0                  |
|                               |                     | 0                   | 12             | W                        | 3                          | Mills and Mann (1985)                         | 4                  |
| <i>Lota lota</i>              | burbot              | 1-3                 | 0 <sup>d</sup> | W                        | 9                          | Kjellman and Eloranta (2002)                  | 0                  |
| <i>Macrhybopsis meeki</i>     | sicklefin chub      | 1-2                 | 10             | W                        | 2                          | Braaten and Guy (2002)                        | 0                  |
| <i>Micropterus salmoides</i>  | largemouth bass     | 0                   | 0              | W                        | 9                          | Schlosser et al. (2000)                       | 0                  |
| <i>Morone americana</i>       | white perch         | 0                   | 15             | W                        | 2                          | O'Gorman and Burnett (2001)                   | 0                  |
| <i>Notropis atherinoides</i>  | emerald shiner      | 1-2                 | 7              | W                        | 2                          | Braaten and Guy (2002)                        | 0                  |
| <i>Oncorhynchus clarkii</i>   | cutthroat trout     | 0                   | 5              | W                        | 2                          | Coleman and Fausch (2007a)                    | 1                  |
| <i>Osmerus eperlanus</i>      | smelt               | 0                   | -3.6           | W                        | 1                          | Mooij et al. (1994)                           | 1                  |
|                               |                     | 1-2                 | 5              | A                        | 7                          | Power and Atrill (2007)                       | 0                  |
| <i>Perca flavescens</i>       | yellow perch        | 0                   | 0              | W                        | 2                          | Mills et al. (1989)                           | 0                  |
|                               |                     | 0                   | 4.4            | A                        | 9                          | Ney and Smith (1975)                          | 0                  |
|                               |                     | 0                   | 10             | A                        | 9                          | Ney and Smith (1975)                          | 0                  |
|                               |                     | 2-4                 | 14             | W                        | 5                          | Henderson (1985)                              | 0                  |
|                               |                     | 0                   | 13.5           | W                        | 5                          | Tardif et al. (2005)                          | 0                  |
| <i>Perca fluviatilis</i>      | European perch      | 1                   | 0 <sup>d</sup> | W                        | 9                          | Borcharding and Magnhagen (2007)              | 1                  |
|                               |                     | 0                   | 9.8            | W                        | 1                          | Mooij et al. (1994)                           | 4                  |

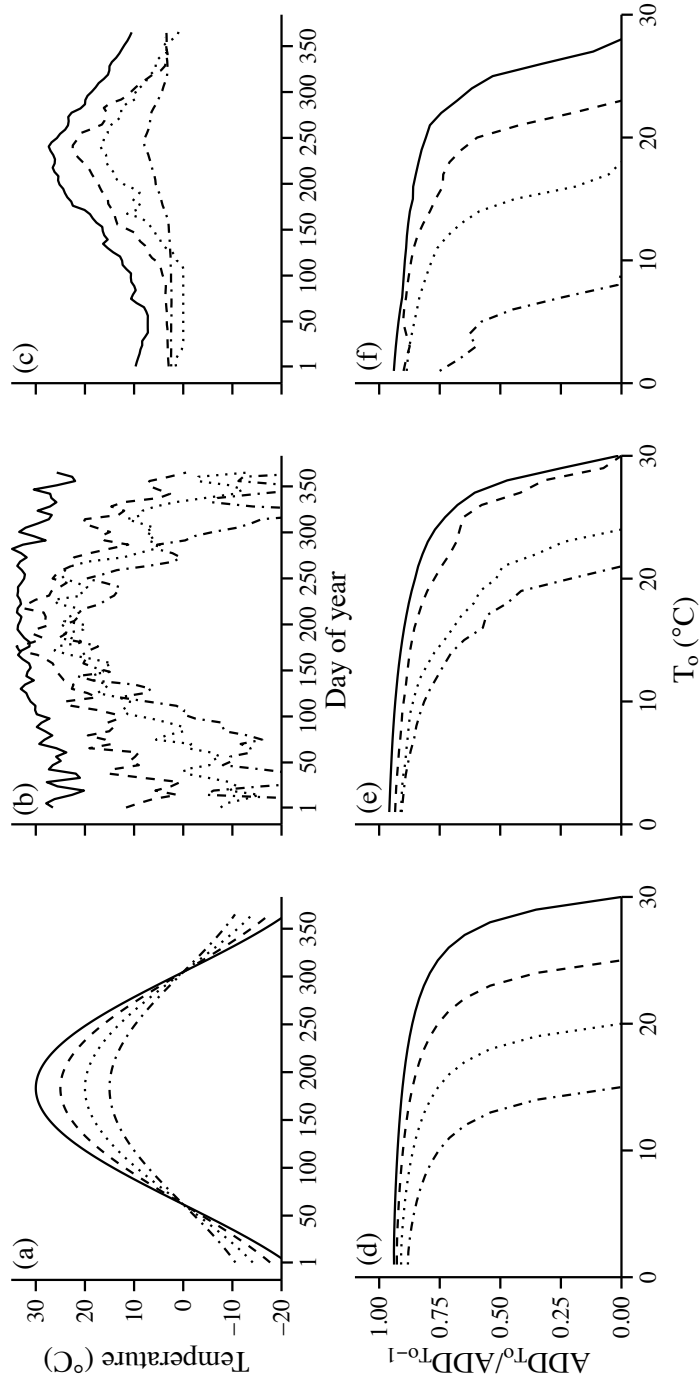


**Table 2.2** Species-specific summary of fish data from both the OMNR and MNDNR.  $ADD_0$  range is the observed range of annual degree-days above 0°C across all cohorts and waterbodies, and cohorts is the number of unique age classes in a given sampling year summed across all sampling years and waterbodies.

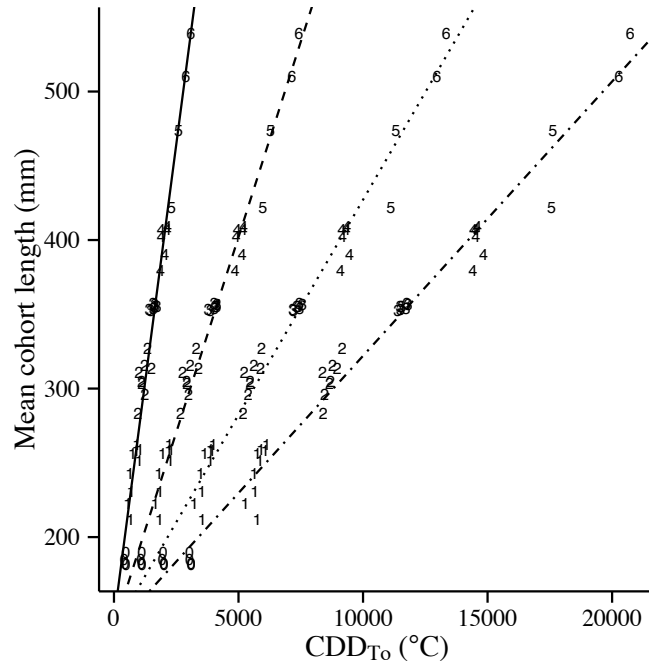
| scientific name               | common name   | waterbodies | $ADD_0$ range | cohorts | fish  |
|-------------------------------|---------------|-------------|---------------|---------|-------|
| <i>Catostomus commersonii</i> | white sucker  | 1           | 2523-3083     | 6       | 54    |
| <i>Coregonus artedii</i>      | cisco         | 1           | 2617-3132     | 5       | 135   |
| <i>Esox lucius</i>            | northern pike | 1           | 2482-3084     | 5       | 113   |
| <i>Lepomis macrochirus</i>    | bluegill      | 2           | 2584-3637     | 11      | 116   |
| <i>Morone chrysops</i>        | white bass    | 1           | 3153-3863     | 6       | 62    |
| <i>Perca flavescens</i>       | yellow perch  | 8           | 2517-3567     | 93      | 3652  |
| <i>Sander canadensis</i>      | sauger        | 6           | 2295-4110     | 86      | 2680  |
| <i>Sander vitreus</i>         | walleye       | 81          | 1886-4110     | 976     | 40674 |

**Table 2.3** Parameter values and range limits for using equation 2.4 to convert annual degree-days at a given  $T_o$  ( $ADD_{T_o,x}$ ) to annual degree-days at one of four  $T_o$  standards ( $ADD_{T_o,y}$ ). Parameter values were estimated for each standard  $T_o$  by fitting curves to describe how the slope and intercept of the  $ADD_{T_o,y}|ADD_{T_o,x}$  relationship changed with  $T_o$  (CurveExpert Professional 1.6.5). Only  $ADD_{T_o,y}|ADD_{T_o,x}$  regressions with an  $r^2 > 0.90$  were included when describing these relationships. The  $T_{o,x}$  range limit indicates the range of  $T_o$  values from which conversion is appropriate and the  $ADD_{T_o,y}$  range limit indicates the range of  $ADD$  values to which conversion is appropriate. Conversion outside of either of these ranges is likely to result in greater prediction error ( $ADD_{T_o,y} \overline{SE}$ ). We estimated prediction error and its standard deviation via bootstrap validation of equation 2.4 ( $1 \times 10^4$  iterations).

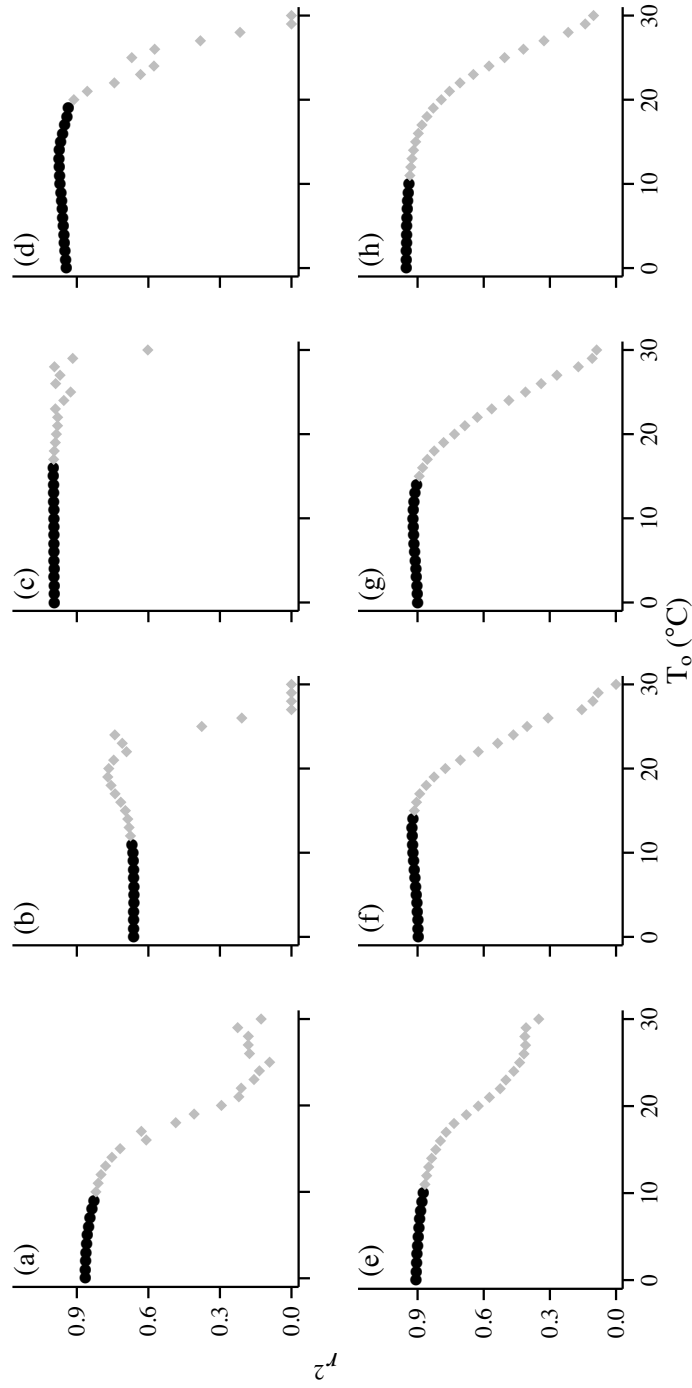
| Standard<br>$T_o$ (°C) | Slope parameter values |       |       | Intercept parameter values |         | $T_{o,x}$ range<br>limit (°C) | $ADD_{T_o,y}$ range<br>limit (°C-days) | $ADD_{T_o,y} \overline{SE}$<br>and ( $\sigma_{\overline{SE}}$ ) |
|------------------------|------------------------|-------|-------|----------------------------|---------|-------------------------------|--|---|
|                        | $a_1$                  | $a_2$ | $a_3$ | $a_4$                      | $a_5$   |                               |  |   |
| 0                      | 0.05                   | 0.99  | -     | 126.00                     | -32.44  | 0-8                           | 1809-3725                              | 46 (0.61)   |
| 5                      | 1.31                   | -0.08 | 0.81  | 96.04                      | -446.21 | 0-12                          | 922-2425                               | 37 (0.32)   |
| 10                     | 1.66                   | -0.05 | 1.13  | 68.61                      | -681.79 | 2-16                          | 45-1464                                | 26 (0.36)   |
| 15                     | 3.77                   | -0.36 | 0.75  | 35.98                      | -544.76 | 8-19                          | 43-743                                 | 17 (0.24)   |



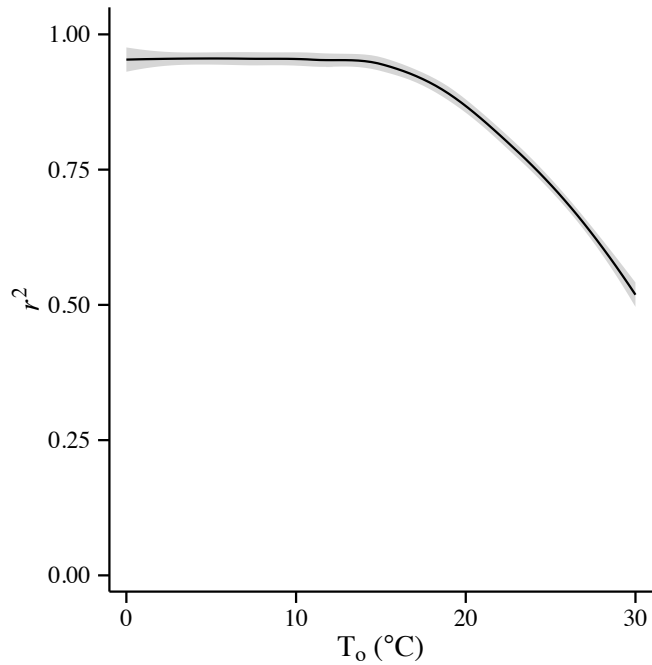
**Figure 2.1** Annual temperature curves and associated values of  $ADD_{T_0-1}$  expressed as a proportion of  $ADD_{T_0-1}$ . Data in (a) and (d) are from a sine curve (equation 2.2) with  $w$  values of 15 (·-·), 20 (···), 25 (---) and 30°C (-). Data in (b) and (e) are from daily 2009  $T_{Min}$  and  $T_{Max}$  values at weather stations (NOAA, unpublished data, National Climatic Data Center) in Bettles Alaska (·-·), Duluth Minnesota (···), Springfield Missouri (---) and Everglade City Florida (-). Data in (c) and (f) are from daily mean temperature values taken above the thermocline within Shadow Bay of Lake Chaeukuktuli Alaska, 2008 (·-·) (Darrell S. Kaufman, unpublished data, Climatic records from lakes in southern Alaska), Lake Superior Minnesota (NOAA, unpublished data, Index of publications and tech reports), 1992 (···), Pillager Lake Minnesota, 2011 (Andrew Carlson MnDNR, unpublished data, 1601 Minnesota Drive, Brainerd, MN 56401) (---) and Lake Powell Utah, 2011 (-) (Wayne Gustavson, unpublished data, water-data.com).



**Figure 2.2** Example mean cohort length-at-age given  $CDD_{T_0}$  regression analysis at  $T_0 = 0$  (·-·), 5 (···), 10 (---), and 15°C (-). Data are from walleye in Cass Lake, MN. Number points show the age in years of each cohort. Each regression had 36 degrees of freedom, p-values < 0.05 and  $r^2$  values  $\geq 0.97$ .



**Figure 2.3** The coefficient of determination ( $r^2$ ) for the relationship between total length (mm) and  $CDD_{T_0}$  over a range of  $T_0$  values for (a) white sucker in Lake of the Woods ON (see Table 2.2), (b) cisco in Leech Lake MN (see Table 2.2), (c) northern pike in La Cloche Lake ON (see Table 2.2), (d) bluegill in Pumushe Lake MN (63 individuals across 6 cohorts), (e) white bass in Lac Qui Parle MN (see Table 2.2), (f) yellow perch in Lake Mille Lacs MN (2299, 26), (g) sauger in Lake Pepin MN (859, 21) and (h) walleye in Lake Pepin MN (616, 26). Filled circles show the approximate extent of each  $r^2$  plateau.



**Figure 2.4** The coefficient of determination ( $r^2$ ) for the relationship between total length (mm) and  $CDD_{T_o}$  over a range of  $T_o$  values, averaged over 81 walleye populations in Minnesota ( $n=20$ ) and Ontario ( $n=61$ ). The solid line is LOESS fit ( $\alpha = 0.65$ ) of all data combined and the shaded area is the 95% confidence interval.

## Chapter 3

### **Fish growth and degree-days II: Selecting a base temperature for an among population study.**

#### **Synopsis**

The degree-day (DD) is a method of describing the thermal opportunity for growth and development in fish, and is becoming increasingly popular when comparing growth over large spatial scales. The degree-day equation includes a lower temperature threshold ( $T_0$ ) so that calculations are constrained to temperatures that are relevant to growth. However, there is currently no convention for choosing  $T_0$ , and unknown are the effects of an incorrect  $T_0$  on how growth is perceived. We used model simulations to demonstrate an effect of  $T_0$  error on apparent growth that increased both with the size of the error and the among-population difference in thermal habitat. We then showed the same relationships in an analysis of length-at-age data from 81 walleye (*Sander vitreus*) populations in North America. Our results show that apparent differences in among-population growth can be an artifact of  $T_0$  rather than a significant biological phenomenon, especially when populations span a wide thermal range. To minimize this artifact, we recommend carefully assigning species to one of four  $T_0$  standards (0, 5, 10, 15°C).

### 3.1 Introduction

The relationship between fish growth and temperature is well documented and highly correlated (Brett 1969, Fry 1971) yet many growth models do not incorporate a thermal component (Neuheimer and Taggart 2007). The degree-day ( $DD_{T_0}$ ;  $^{\circ}\text{C}\cdot\text{days}$ ) is an index of the thermal opportunity for growth and development. Degree-day calculations incorporate a lower threshold temperature ( $T_0$ ) so that they are limited to temperatures relevant to growth. However, Chezik et al. (*in review*) showed that precise  $T_0$  estimates are largely unnecessary when explaining annual growth in a single population because degree-days at different  $T_0$  values tend to be similar (especially as  $T_0$  approaches  $0^{\circ}\text{C}$ ).

Although  $T_0$  matters little when describing growth in a single population,  $T_0$  may become important when describing growth over a wide thermal range. For example, we can imagine a simple thought experiment in which growth rates at the thermal extremes of a species' range are identical, but can appear to be different depending on  $T_0$  (Fig. 3.1). We begin by imagining separate annual temperature curves that describe the thermal environment at these extremes. We also imagine that fish at these extremes share a growth rate  $\left(\frac{mm}{^{\circ}\text{C}\cdot\text{Day}}\right)$  and do not grow below some limiting temperature (i.e., 'true'  $T_0$ ). If we calculate degree-days at 'true'  $T_0$ , fish at either extreme have identical growth rates. However, if we calculate degree-days assuming a  $T_0$  above 'true'  $T_0$ , fish in the cooler environment appear to be growing faster (i.e.,  $\frac{mm}{^{\circ}\text{C}\cdot\text{Day}}$  is greater).

In our thought experiment, the apparent difference in growth rate at  $T_0 > \text{'true' } T_0$  stems from differences between the annual temperature curves of the cooler and warmer environments. In both environments, 'true' degree-days are underestimated at high  $T_0$  because degree-days decrease with increasing  $T_0$ . As the number of degree-days decrease, apparent growth rates  $\left(\frac{mm}{^{\circ}\text{C}\cdot\text{Day}}\right)$  increase because growth remains constant. However, this effect is exaggerated in the cooler environment because degree-days in cooler environments decrease with increasing  $T_0$  more rapidly than in warmer environments (Fig. 3.1e). Therefore, because  $T_0$  in our thought experiment is  $> \text{'true' } T_0$ , fish in the cooler environment appear to grow faster than fish in the warmer environment.

This thought experiment suggests that  $T_o$  can influence our interpretation of growth rates among populations that are thermally dissimilar.

If  $T_o$  can exaggerate differences in apparent growth rates among populations then this may have implications for studies that use degree-days to describe and compare growth over a wide thermal range (Table 3.1). For example, many species exhibit counter-gradient growth, which is an increase in growth rate with latitude (e.g., Power and McKinley 1997, Conover et al. 2009, Chavarie et al. 2010). This pattern has been attributed to genetically based phenotypic adaptations to climate (Rypel 2012) but it could also be a statistical artifact that resulted from an inappropriate  $T_o$ . Current methods for identifying  $T_o$  do not consider and have not accounted for the effect of  $T_o$  on apparent growth. In fact, many studies provide little or no justification for their use of  $T_o$  and when justification is provided, it is usually that  $T_o$  is the approximate minimum temperature associated with some physiological process (e.g., growth). These various methods have resulted in a wide range of  $T_o$  values (0-18°C) currently in use among species, with as many as four different values applied to a single species (e.g., yellow perch, *Perca flavescens*, 0, 5, 10 and 13.5°C) (Table 3.1).

In this study, we determine the effect of  $T_o$  on apparent growth rates among populations in thermally dissimilar environments. This work is a companion to Chezik et al. (*in review*), which describes how degree-days vary with  $T_o$ , and how this variation affects the ability of degree-days to account for growth in a single population. Here we explore how the relationship between degree-days and  $T_o$  varies among waterbodies in different climates, and determine how this variation affects differences in apparent growth. We explore these relationships by using both theoretical and empirical data to compare among-population growth rates across thermal ranges and for different values of  $T_o$ .

### **3.2 Simulated effects of $T_o$ and thermal difference on apparent growth rates**

In this section, we simulate the thought experiment in Fig. 3.1 using hypothetical data from four populations; we then simulate an extensive database to determine how apparent growth rates might vary both with  $T_o$  and the degree of thermal difference among

populations. The populations in these simulations share i) a maximum annual air temperature of 35°C, ii) a growing season of 160 days, iii) an immature growth rate of  $0.03 \frac{mm}{^{\circ}C \cdot Day}$  at a  $T_o$  of 5°C, and iv) no discernable length at hatch. This approach ignored interactions between growing season, mean annual temperature and maximum annual temperature in favor of a simplified model for exploring how among-population differences in the availability of thermal energy affect apparent growth. We used a sine curve (Arnold 1959) to generate annual temperature profiles for each simulated waterbody and then calculated annual degree-days ( $ADD_{T_o}$ ; °C·days) by integrating the area under each curve (Baskerville and Emin 1969, Chezik et al. *in review*). Temperature curves and  $ADD_{T_o}$  values varied by their assigned mean annual temperature determined in each simulation. We then predicted immature length-at-age as

$$(3.1) \quad \hat{L} = \frac{dl}{add_5} \cdot [(ADD_5 \cdot Age) + ADD_5],$$

where,  $\frac{dl}{add_5}$  is the change in length per degree-day at a  $T_o$  of 5°C, *Age* is age in years assuming a common birthday of January 1, and  $ADD_5$  is the annual degree-days experienced at a  $T_o$  of 5°C. In this way each simulation was made up of thermally unique waterbodies with fish that share a common growth rate.

To simulate the effect of  $T_o$  on apparent immature growth, we imagined three studies that compared growth in four waterbodies over a large thermal range. These four waterbodies had mean annual temperatures of -7, 5, 17, and 29°C, respectively. The three studies were identical except that each used a different  $T_o$  to calculate degree-days (-5, 5, and 15 °C). Our simulation showed that the study that assumed a  $T_o$  of 5°C would have concluded that all populations shared the same growth rate, but that the other studies would have concluded that growth rates either increased ( $T_o = 15^{\circ}C$ ) or decreased ( $T_o = -5^{\circ}C$ ) over the same thermal range (Fig. 3.2). Because growth in our simulation began at 5°C (i.e., ‘true’  $T_o = 5^{\circ}C$ ), apparent differences in immature growth were entirely an artifact of choosing an inappropriate  $T_o$ . As in our simple thought experiment, these artifacts stem from the interaction between an inappropriate  $T_o$  and among-waterbody differences in annual temperature curves. Specifically, an inappropriate  $T_o$  biases degree-day calculations in all waterbodies, but this bias is smallest in the warmest waterbody and

largest in the coolest waterbody. These degree-day differences translate into apparent differences in the rate at which fish grow in different thermal environments.

Given that apparent growth rates depend on  $T_o$  and among-waterbody differences in annual temperature (Fig. 3.2), it stands to reason that the effect of an inappropriate  $T_o$  on apparent immature growth increases with both the among-waterbody difference in annual temperature and the difference between ‘inappropriate’  $T_o$  and ‘true’  $T_o$ . To test these hypotheses, we first created 200 waterbodies with mean annual temperatures that were uniformly spread over a narrow thermal range (7.3-7.9°C). Each temperature curve was approximated by a sine curve (Baskerville and Emin 1969, Chezik et al. *in review*) and ADD values were calculated for each  $T_o$  in the range -20 to 20°C. We used equation 3.1 to generate immature length-at-age data for each temperature curve (i.e., waterbody) and we incorporated temperature-independent variation in length-at-age by randomly sampling normal distributions centered on each calculated length-at-age, assuming a standard deviation of 0.08 mm. This standard deviation was based on observed variation in walleye length-at-age (see next section) and was therefore realistic. We incorporated temperature independent length-at-age variation because growth is rarely exclusively dependent on temperature in natural systems.

For each waterbody, we estimated immature growth rate by regressing length onto cumulative degree-days (CDD; °C·days) (i.e., the summation of all experienced  $ADD_{T_o}$ ) at each  $T_o$  value. We then identified the  $T_o$  that minimized the coefficient of variation (CV) in growth rates among waterbodies. Our rationale for using the CV was that, whereas growth rates at ‘true’  $T_o$  are identical in deterministic simulations (Fig. 3.2), growth rates at ‘true’  $T_o$  are likely to be most similar in stochastic simulations. However, because variation in length-at-age can result in the CV being minimized at a  $T_o$  other than ‘true’  $T_o$ , we allowed the CV method to identify biologically meaningless (i.e., negative)  $T_o$  values (Yang et al. 1995, Legg 1998a). To determine the distribution of  $T_o$  values at which CV was likely to be minimized, we generated length-at-age data 1000 times for each waterbody, and for each iteration determined the  $T_o$  at which CV was minimized. We then repeated this entire process for 200 waterbodies that had mean annual temperatures spread uniformly between 0 and 15°C. The result was two distributions of

$T_o$  values at which the CV of growth was minimized, one for a wide thermal range and one for a narrow thermal range.

These preliminary simulations show that, on average, CV was minimized when  $T_o$  was equal to ‘true’  $T_o$  (Fig. 3.3a). This relationship was especially pronounced for the large thermal range. The large thermal range also resulted in a much narrower distribution of  $T_o$  values over which CV was minimized (Fig. 3.3b). This inverse relationship between thermal range and the width of the  $T_o$  distribution resulted from the relative importance of among-waterbody thermal variation and variation in length-at-age. When waterbodies were distributed over a small thermal range, among-waterbody variation in growth was primarily a result of variation in length-at-age (i.e., noise) such that the CV was minimized over a wide range of  $T_o$  values. When waterbodies were distributed over a large thermal range, among-waterbody variation in growth was primarily a result of differences in mean annual temperature such that the CV was minimized at only a few  $T_o$  values.

Finally, we created 38 additional thermal ranges that had a median annual temperature of 7.5°C and were uniformly distributed in width between the narrowest thermal range (7.3-7.9°C) and the widest thermal range (0-15°C). As described above, each thermal range contained 200 waterbodies and length-at-age in each waterbody was randomly generated 1000 times. When growth was simulated in this way, we found that as thermal range increased, there was a rapid decrease in the distribution of  $T_o$  values at which CV was minimized (Fig. 3.4a) and gradual increase in the effect of  $T_o$  on apparent growth (Fig. 3.4b). Figure 3.4b shows clearly that increased differences in growth rates over a wide thermal range are a result of a bias that was introduced by selecting an inappropriate  $T_o$ . This bias increased with the magnitude of the difference between inappropriate  $T_o$  and ‘true’  $T_o$ . The results of this simulation suggest that evidence for counter-gradient growth among populations can be a result of  $T_o$  rather than a biological phenomenon. For instance, in this simulation, using a  $T_o$  of 15 rather than the ‘true’ value of 5°C inflated growth rates by 5% over a thermal range of 900 °C·days.

### **3.3 Observed effects of $T_o$ and thermal difference on apparent growth rates**

Although our simulations show that an inappropriate  $T_0$  can artificially generate among-waterbody differences in growth rate, especially over large thermal ranges, unknown is the extent to which this effect is observable in nature. To determine if among-waterbody variation in observed growth is sensitive to  $T_0$  and thermal range, we analyzed a database of immature walleye (*Sander vitreus*) length-at-age (40,674 individual records) and air temperature data from 81 waterbodies throughout Minnesota, US, and Ontario, Can. See Chezik et al. (*in review*) for a detailed description of this database. In brief, individual fish records were collected by the Minnesota Department of Natural Resources (2001-2011) and the Ontario Ministry of Natural Resources (1993-2008). Fish records included, waterbody, sample date (day, month, year), age (years), total length (L; mm) and maturity. For waterbodies in Minnesota, air temperature data ( $T_{Max}$  and  $T_{Min}$ ) were kriging-interpolated onto  $100\text{km}^2$  grids and temperatures estimated by determining the grid point nearest the centroid of each lake. Degree-days were calculated at a range of  $T_0$  (0-20°C) and summed to produce annual degree-days ( $ADD_{T_0}$ ) for each year of interest. For waterbodies in Ontario,  $ADD_{T_0}$  values were obtained using the Historic Climate Analysis Tool (Cross et al. 2012). We assigned  $CDD_{T_0}$  values to each fish by summing the annual degree-days experienced prior to capture.

Our analysis of observed length-at-age data was similar to our analysis of simulated length-at-age data. For each waterbody, we regressed mean length-at-age onto cumulative degree-days at each  $T_0$  in the range 0-20°C and determined growth rate ( $\frac{mm}{^\circ\text{C}\cdot\text{day}}$ ) using ordinary least squares. We used a simple linear model because linear immature growth is predicted by theory (Quince et al. 2008) and was observed in this database (Chezik et al. *in review*). We then defined 17 unique thermal ranges, each containing  $\geq 15$  waterbodies. To define these thermal ranges, we first determined the thermal character of each waterbody by i) identifying all unique years experienced by the cohorts in a given waterbody, and ii) calculating mean  $ADD_0$  across these years. We then identified the 15 waterbodies that had mean  $ADD_0$  values that were closest to the median  $ADD_0$  (2645 °C·days) across all waterbodies in the database. This step established a narrow thermal range (2550-2720 °C·days) that included a ‘thermal group’ of 15

waterbodies. We began with 15 waterbodies to ensure a large enough sample to meet the needs of all statistical methods (see next paragraph). We then increased the thermal limits of the initial range in increments of 100°C·days. Additional thermal groups were identified as the number of included waterbodies increased. This process resulted in 17 thermal ranges, the largest of which ranged from 2090-3710 °C·days at  $T_o$  of 0°C and included all waterbodies. For each thermal range, we generated a distribution of  $T_o$  values at which the CV of growth was minimized by sampling, without replacement, 85% of waterbodies 1000 times. This approach assumed that sample variance was approximately equal to ‘population’ variance. This procedure was nearly identical to that of our simulation in the previous section, except the number of waterbodies (i.e., growth rates) in each thermal group was variable and waterbodies were unevenly distributed over the thermal range.

Our results show that the CV in growth rate among waterbodies increased with  $T_o$  (Fig. 3.5a). This effect was more pronounced over the entire thermal range than it was over the narrowest thermal range (Fig. 3.5a). The distribution of  $T_o$  over the entire thermal range had a median value of 3°C with a range of 0 to 6°C (Fig. 3.5b). Over the narrowest thermal range the median value of  $T_o$  was 0°C (range 0 to 16°C). Furthermore, as thermal range decreased, CV in growth tended to be minimized at a  $T_o$  of 0°C (Fig. 3.5b). This trend was a result of limiting our empirical statistical analysis to biologically relevant (i.e., non-negative)  $T_o$  values. As noted in Yang et al. (1995)  $T_o$  identification using statistical analysis may sometimes select negative values that make little biological sense. This limitation resulted in a standard deviation of 0 for many thermal groups (Fig. 3.6a). A result more similar to figure 3.4a is expected if negative values were included. Finally, when using a  $T_o$  of 15°C, the difference in growth rate between the 1<sup>st</sup> and 3<sup>rd</sup> quartiles significantly increased with thermal range (Fig. 3.6b). The difference in the growth rate interquartile range was as much as  $0.12 \frac{mm}{^\circ C \cdot day}$  higher than the smallest thermal group. We did not detect a significant effect of  $T_o$  on apparent growth rates when  $T_o$  was 0, 5, or 10°C.

### 3.4 Discussion

Our results show that apparent growth rates vary with both  $T_o$  and thermal range (Fig. 3.4), and that these relationships are observable in empirical data (Fig. 3.6). The apparent variability in growth rate increases as assumed  $T_o$  deviates from the  $T_o$  that minimizes the among-waterbody variation in growth rate (presumably ‘true’  $T_o$ ), and this effect becomes more pronounced as the thermal range among waterbodies increases (Fig. 3.4b). For instance, we found that variation in the growth rates of immature walleye increased with thermal range when  $T_o$  was 15°C, but not when  $T_o$  was 0, 5, or 10°C. This result suggests that a hypothetical study that used a  $T_o$  of 15°C to compare growth rates among the populations in this database would have concluded that walleye are latitudinally adapted and exhibit counter-gradient growth. Given the results of our simulation (Fig. 3.4b) and the absence of counter-gradient growth at lower values of  $T_o$  (Fig. 3.6b), it is entirely possible that this conclusion would be false. Overall, our analyses suggest that it is possible to falsely detect counter-gradient growth, and that care must be taken to select a  $T_o$  that minimizes this statistical bias.

The effect of assumed  $T_o$  on apparent growth rates is greater when assumed  $T_o$  is less than ‘true’  $T_o$  than when assumed  $T_o$  is greater than ‘true’  $T_o$  (Fig. 3.3a, Fig. 3.5a). In our simulations, the effect of  $T_o$  on apparent growth stemmed from both error in  $T_o$  and among-waterbody differences in temperature curves. When both of these conditions were met they resulted in waterbody-specific biases in apparent growth. However, because annual degree-days become more similar as  $T_o$  values get smaller (Chezik et al. (*in review*)), biases in degree-days, and therefore the bias in apparent growth, were larger when  $T_o$  was overestimated by some amount than when  $T_o$  was underestimated by the same amount.

Given that the effect of assumed  $T_o$  on apparent growth rates is greater when assumed  $T_o$  is larger than ‘true’  $T_o$ , a risk-averse strategy is to err low when selecting a  $T_o$  for an among-population study. This will reduce the risk of introducing bias that may otherwise result when simply relying on precedent or convention. For example, Power and van den Heuvel (1999) compared the growth of yellow perch in northern Alberta to a population in southern Manitoba using a  $T_o$  of 13.5°C. This  $T_o$  is routinely used to

describe European perch (*P. fluviatilis*) growth in Lake Windermere (Le Cren 1958), but may be sufficiently high to introduce a bias when comparing growth over a wide thermal range ( $\Delta\text{ADD}_5 \sim 950 \text{ }^\circ\text{C}\cdot\text{days}$ ). To test for such a bias, we repeated the Power and van den Heuvel (1999) analysis for a range of  $T_0$  values (5-12°C and 13.5°C). Our results (Table 3.2) show that the difference between growth models (i.e., growth rates) is greatest at 13.5°C and statistically insignificant when  $T_0$  is 8, 9 or 10°C (Fig. 3.7). Power and van den Heuvel (1999) attributed the evidence for counter-gradient growth at  $T_0 = 13.5^\circ\text{C}$  to “latitudinal or trophic factors”; however, our re-analysis suggests that this evidence is either exaggerated or solely a result of an inappropriately large  $T_0$ . Although this study may be an example of  $T_0$  detecting growth differences when none exist, it is encouraging that many comparative growth studies tend to use low values of  $T_0$  that are less prone to artificially inflating growth rates (Table 3.1). However, we suspect that 18°C (the minimum temperature for growth of larval gar, *Lepisosteus oculatus*) is too high for comparing growth of individuals age 0-16 over 2240-7570  $\text{ADD}_5$  (David 2012).

If error in  $T_0$  can bias among-population growth studies, especially over wide thermal ranges, it follows that  $T_0$  should be estimated with care. Unfortunately, many  $T_0$  values currently in use are rarely justified (Table 3.1) and others may bias results (e.g., Power and van den Heuvel 1999). Our results suggest that  $T_0$  can be estimated as the  $T_0$  that minimizes the CV of growth over a large thermal range; however, data are rarely available to perform such an analysis. Similarly, although there are methods for estimating  $T_0$  in single populations, results often vary among methods and populations (reviewed by Chezik et al. *in review*).

A relatively simple approach of estimating  $T_0$  is to determine the mean development temperature and subtract 10°C (Charnov and Gillooly 2003). To demonstrate, we used the Wisconsin Bioenergetics model (Hanson et al. 1997) to describe how walleye and yellow perch growth rates changed with temperature (Fig. 3.8). Given that mean developmental temperature in these species occurs at 15 and 18°C, respectively, the “10°C rule” predicts that  $T_0$  is 5 and 8°C, respectively. These results are consistent with our analysis of walleye data (i.e., 3°C, Fig. 3.5b), and with our re-analysis of yellow perch data showing that among-population variation in perch growth was

minimized at 9°C. Although the method needs to be explored further, it does show promise as a means of assigning a species to one of the  $T_0$  standards (0, 5, 10, and 15°C) advocated by Chezik et al. (*in review*). Assigning species to a standard  $T_0$  (e.g., 5°C for walleye and 10°C for yellow perch) would minimize the risk of using  $T_0$  values that bias growth rate comparisons, simplify comparisons among datasets and studies, and facilitate the use of degree-days in future studies.

In this study, we have shown that an inappropriate  $T_0$  can bias growth comparisons among waterbodies. This bias leads to apparent differences in growth rates among populations that are solely an artifact of  $T_0$ . This effect of  $T_0$  on apparent growth is exaggerated by large thermal differences among waterbodies. Because these thermal differences are predominately observed over a latitudinal gradient, an inappropriate  $T_0$  can result in the appearance of counter-gradient growth. To minimize the risk of introduced bias, we suggest first estimating  $T_0$  using the “10°C rule” (Charnov and Gillooly 2003) method and then setting  $T_0$  to the nearest standard value. Doing so will limit the introduction of a bias that may obscure differences among populations and both simplify and encourage the use of degree-days in future studies. Although, there is no evidence that this effect of  $T_0$  is observed in marine systems, until further study we suggest taking similar precautions when choosing  $T_0$ . The results of this study should pertain to spatially isolated marine populations that inhabit thermally dissimilar environments. Populations that are not geographically isolated and therefore can behaviorally thermo-regulate, are unlikely to exhibit an affect of  $T_0$  on apparent growth rates.

**Table 3.1** A summary of published  $T_o$  values used in comparative growth studies of freshwater fishes over a thermal gradient (i.e.,  $ADD_5$  range).

| scientific name                | common name      | location(s)                          | $T_o$ (°C) <sup>a</sup> | temperature<br>medium | justification <sup>b</sup> | range<br>range<br>(°C-days) | reference                     |
|--------------------------------|------------------|--------------------------------------|-------------------------|-----------------------|----------------------------|-----------------------------|-------------------------------|
| <i>Acipenser fluvescens</i>    | lake sturgeon    | Canada (MB, ON, QC, SK), US (WI)     | 5                       | A                     | 1                          | 650-2250                    | Power & McKinley (1997)       |
| <i>Esox lucius</i>             | northern pike    | Minnesota<br>North America & Eurasia | 10                      | A                     | 2                          | 550-4010 <sup>d</sup>       | Jacobson (1992)               |
| <i>Ictalurus punctatus</i>     | channel catfish  | Illinois                             | 10                      | A                     | 3                          | 1120-2730 <sup>e</sup>      | Rypel (2012b)                 |
| <i>Lepisosteus oculatus</i>    | spotted gar      | US (AL, AR, IL, KY, LA, MI, MS, TX)  | 10                      | A                     | 2                          | 4080-6080 <sup>d</sup>      | Shoup et al. (2007)           |
| <i>Lepomis macrochirus</i>     | bluegill sunfish | Illinois                             | 18                      | A                     | 1                          | 2240-7570 <sup>f</sup>      | David (2012)                  |
| <i>Micropterus dolomieu</i>    | smallmouth bass  | Illinois                             | 10                      | A                     | 2                          | 4080-6080 <sup>d</sup>      | Shoup et al. (2007)           |
| <i>Micropterus salmoides</i>   | largemouth bass  | Canada & US                          | 10                      | A                     | 4                          | 1160-4660 <sup>d</sup>      | Beamesderfer & North (1995)   |
|                                |                  | Canada & US                          | 10                      | A                     | 4                          | 1640-3950 <sup>e</sup>      | Dunlop & Shuter (2006)        |
|                                |                  | Canada & US                          | 10                      | A                     | 4                          | 1160-4660 <sup>d</sup>      | Beamesderfer & North (1995)   |
|                                |                  | Illinois                             | 10                      | A                     | 2                          | 4080-6080 <sup>d</sup>      | Shoup et al. (2007)           |
| <i>Morone saxatilis</i>        | striped bass     | Canada & US                          | 10                      | A                     | 5                          | 1030-7510 <sup>f</sup>      | McCauley & Kilgour (1990)     |
| <i>Oncorhynchus clarkii</i>    | cutthroat trout  | E. USA & E. Canada                   | 10                      | A                     | 3                          | 1400-5420 <sup>d</sup>      | Rypel (2012a)                 |
| <i>Perca flavescens</i>        | yellow perch     | North Central Colorado               | 0 <sup>c</sup>          | A                     | 2                          | 900-1010 <sup>d</sup>       | Coleman & Fausch (2007b)      |
|                                |                  | Alberta                              | 0 <sup>c</sup>          | W                     | 2                          | 1020-1120 <sup>d</sup>      | Abbey & Mackay (1991)         |
|                                |                  | Ontario                              | 10                      | A                     | 1                          | 1030-2330 <sup>d</sup>      | Chong (2000)                  |
|                                |                  | Ontario                              | 0 <sup>c</sup>          | W                     | 2                          | 660-1910 <sup>d</sup>       | Post & McQueen (1994)         |
|                                |                  | Alberta, Manitoba & Saskatchewan     | 13.5                    | W                     | 6                          | 1050-1710                   | Power & van den Heuvel (1999) |
|                                |                  | Ontario                              | 5                       | A                     | 2                          | 1160-2070                   | Purchase et al. (2005)        |
|                                |                  | Ontario                              | 5                       | W                     | 7                          | 1160-2070                   | Rennie et al. (2010)          |
| <i>Phoxinus phoxinus</i>       | Eurasian minnow  | Ontario                              | 5                       | A                     | 3                          | 470-710                     | Mills (1988)                  |
| <i>Pomoxis nigromaculatus</i>  | black crappie    | Southern England & Central Finland   | 15.5                    | A                     | 8                          | 2630-3500 <sup>f</sup>      | McInerney & Cross (1999)      |
| <i>Richardsonius balteatus</i> | reside shiner    | Southern Minnesota                   | 8                       | A                     | 1                          | 1670-3400 <sup>d</sup>      | Houston & Belk (2006)         |
| <i>Salvelinus alpinus</i>      | arctic char      | Utah, Idaho, Wyoming                 | 0                       | A                     | 1                          | 30-1960 <sup>e</sup>        | Chavarie et al. (2010)        |
| <i>Sander vitreus</i>          | walleye          | Canada & NE US                       | 5                       | A                     | 2                          | 970-5680 <sup>d</sup>       | Colby & Nepszy (1981)         |
|                                |                  | Canada & US                          | 5                       | A                     | 2                          | 1000-4500                   | Bozek et al. (2011)           |
|                                |                  | Wisconsin                            | 0                       | A                     | 8                          | 1140-1920 <sup>d</sup>      | Sass & Kitchell (2005)        |
|                                |                  | Ontario & Quebec                     | 5                       | A                     | 1                          | 1200-2300                   | Venturelli et al. (2010)      |
| Multiple species               | Multiple species | Michigan                             | 10                      | A                     | 3                          | 1420-3120 <sup>e</sup>      | Wagner et al. (2007)          |

**Table 3.1 Continued (footer)**

<sup>a</sup>W=water, A=air.

<sup>b</sup>J=minimum temperature for physiological process, 2=none given, 3=commonly used in plant studies, 4=cites McCauley & Kilgour, 5=fit a growth model (e.g. bioenergetics model), 6=used for European perch (Le Cren 1958), 7=cites Purchase et al. (2005), 8=used for Atlantic salmon (*Salmo salar*) egg survival (Ketola 2000).

<sup>c</sup>T<sub>0</sub> not reported; assumed to be 0°C.

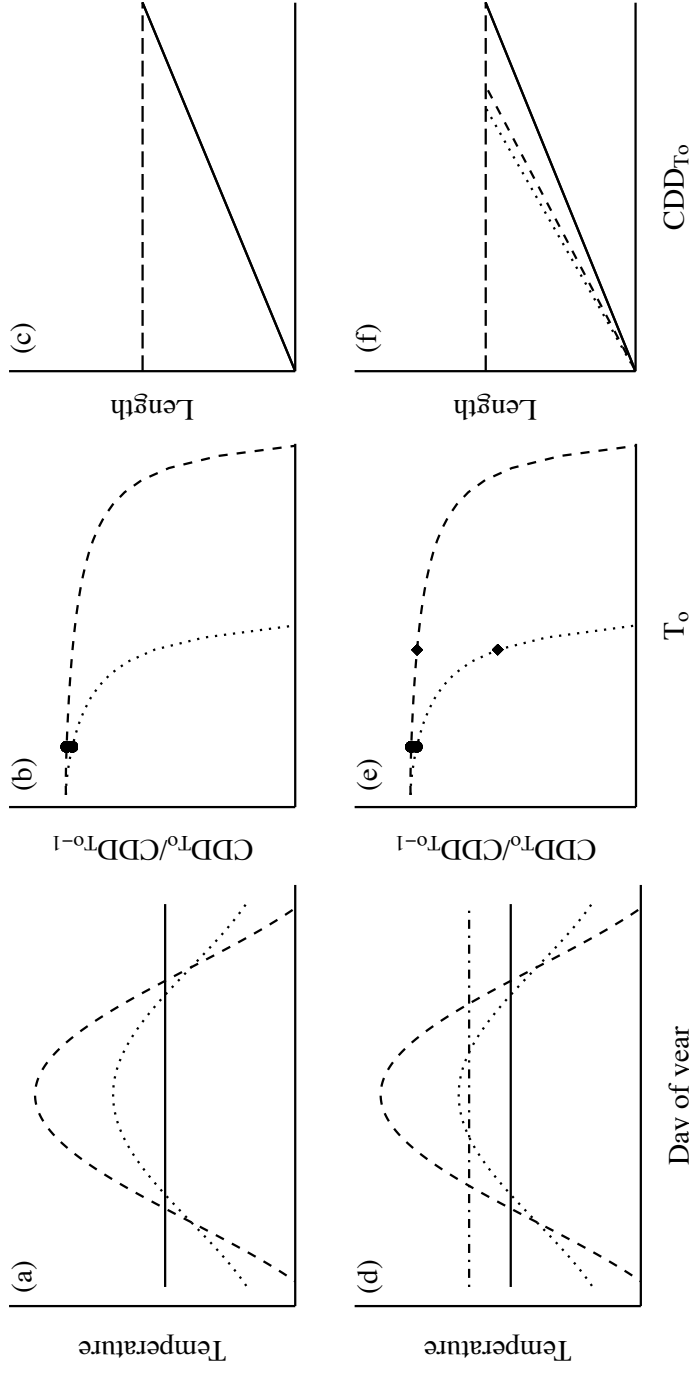
<sup>d</sup>ADD<sub>5</sub> estimated using equation 5 in Chezik et al. (*in review*).

<sup>e</sup>Not given in original literature. Estimated using the 1971-2000 climate normal from the most northern and southern location. Normals were collected from the National Climate Data and Information Archive distributed by Environment Canada and the NOAA National Climatic Data Center.

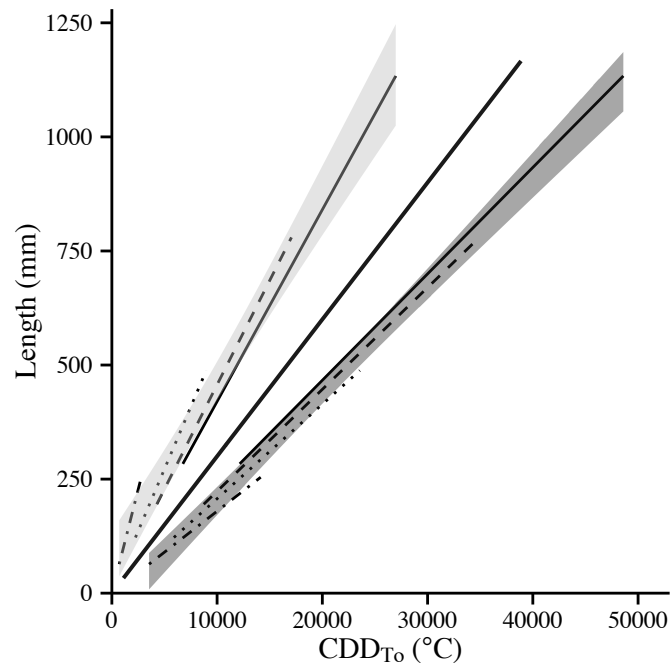
<sup>f</sup>ADD<sub>5</sub> estimated using equation 5 in Chezik et al. (*in review*) beyond the T<sub>0</sub> limit for conversion to T<sub>5</sub>.

**Table 3.2** Length|CDD<sub>T<sub>0</sub></sub> coefficient estimates for three north Alberta lakes (N. Alberta, df=17) and West Blue lake (WBL, df=28) with their associated  $r^2$  values and Chow Test  $F$ -statistic ( $F=2, \alpha=0.05$ ) comparing model estimates. All coefficients were significant at an alpha of 0.05.

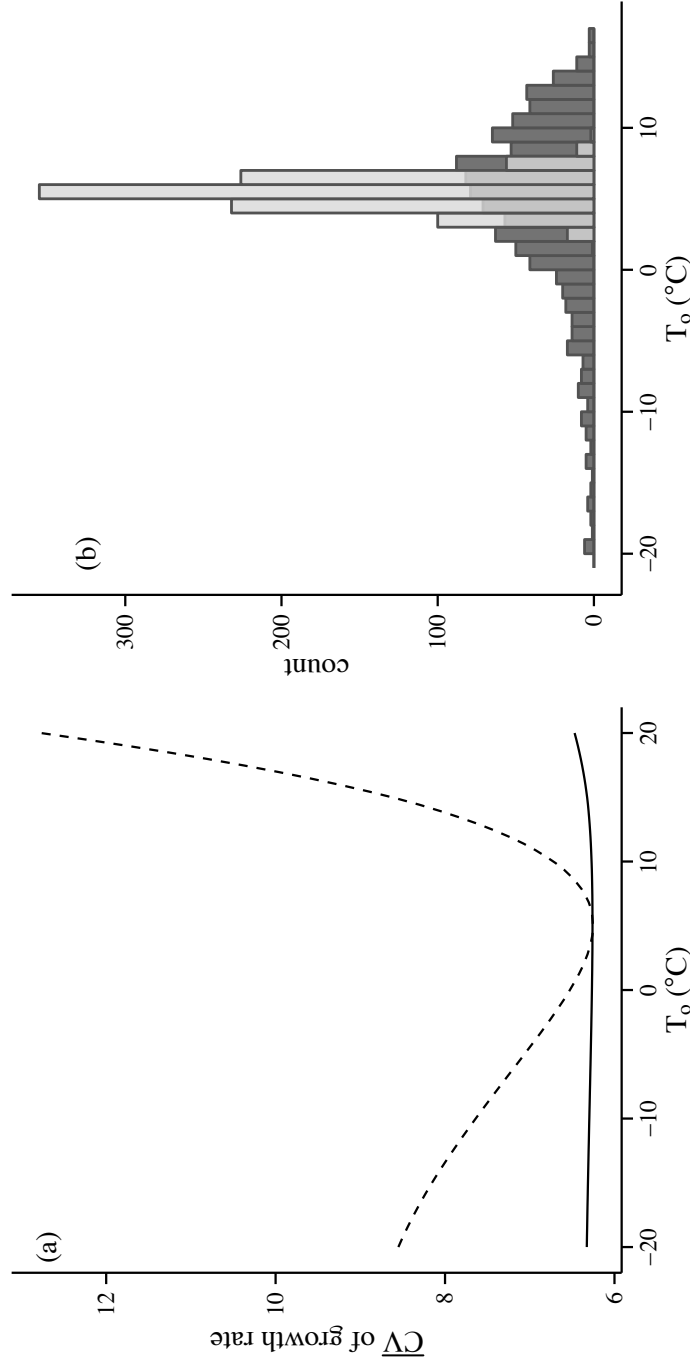
| $T_0$ | $F$ -statistic | $a$ N. Alberta<br>(mm) | $b$ N. Alberta<br>(mm/DD <sub>T<sub>0</sub></sub> ) | $r^2$ N. Alberta | $a$ WBL<br>(mm) | $b$ WBL<br>(mm/DD <sub>T<sub>0</sub></sub> ) | $r^2$ WBL |
|-------|----------------|------------------------|---|------------------|-----------------|--|-----------|
| 5     | 2.624          | 7.478                  | 0.034   | 0.962            | 2.871           | 0.041  | 0.951     |
| 6     | 2.367          | 7.999                  | 0.037   | 0.961            | 3.592           | 0.044  | 0.952     |
| 7     | 2.087          | 8.493                  | 0.041   | 0.960            | 4.430           | 0.047  | 0.954     |
| 8     | 1.782          | 8.905                  | 0.045   | 0.960            | 5.416           | 0.051  | 0.955     |
| 9     | 1.567          | 9.274                  | 0.050   | 0.960            | 6.591           | 0.056  | 0.957     |
| 10    | 1.749          | 9.643                  | 0.055   | 0.960            | 8.018           | 0.062  | 0.959     |
| 11    | 2.932          | 9.979                  | 0.063   | 0.959            | 9.750           | 0.070  | 0.962     |
| 12    | 5.156          | 10.332                 | 0.072   | 0.956            | 11.553          | 0.079  | 0.965     |
| 13.5  | 7.349          | 11.199                 | 0.090   | 0.949            | 13.648          | 0.099  | 0.970     |



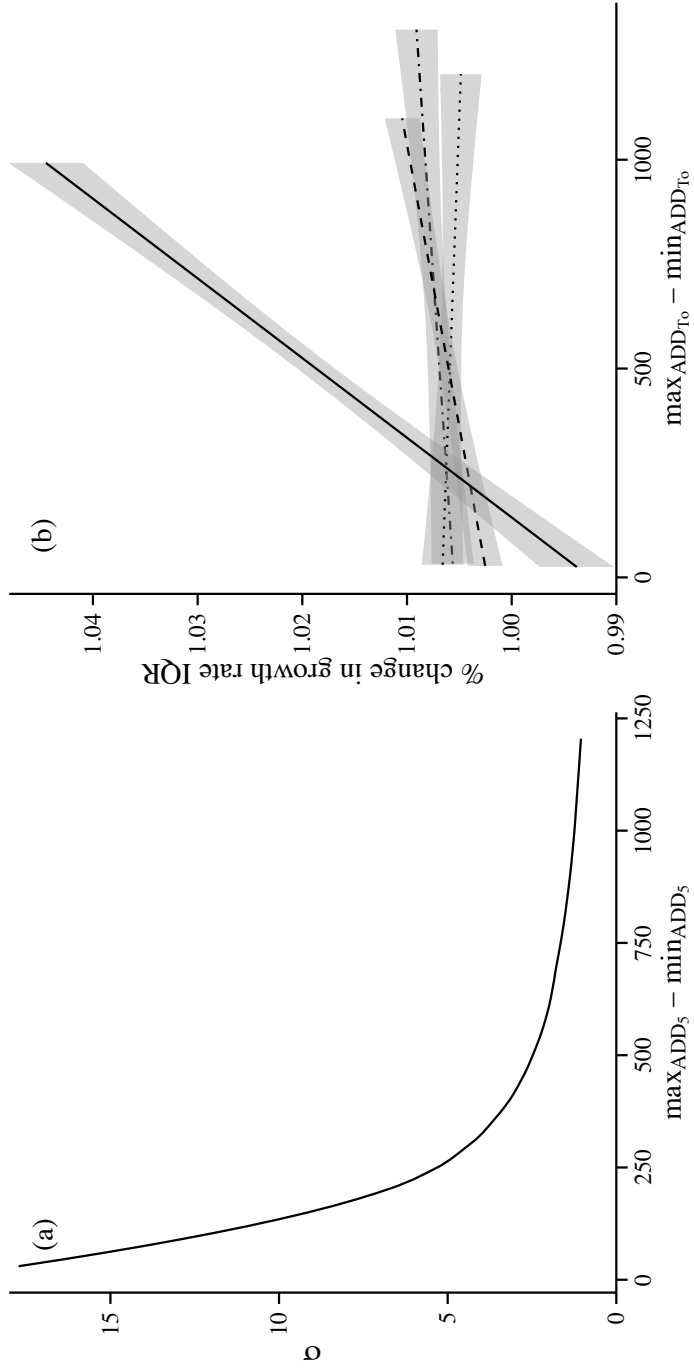
**Figure 3.1** Depiction of a hypothetical thought experiment showing the effect of  $T_0$  on apparent growth rates in two waterbodies from different climates. Panel (a) is the temperature curves for the two waterbodies: one in a cooler climate (•••) and one in a warmer climate (---). The horizontal line in (a) refers to the 'true' base temperature ( $T_0$ ) below which fish in these waterbodies do not grow. Panel (b) shows the proportion of cumulative degree-days (CDD; °C-days) retained at a given  $T_0$  relative to the cumulative degree-days at the previous  $T_0$  for both the cool (•••) and warm (---) waterbodies (Chezik et al. *in review*). The (•) shows CDD at 'true'  $T_0$ . Panel (c) is the apparent growth trajectory of immature fish in these lakes up to a hypothetical length at maturity (long --) as described by cumulative degree-days at 'true'  $T_0$ . Panel (d) depicts the use of an 'incorrect'  $T_0$  (•-•) that is higher than the 'true'  $T_0$ . Panel (e) shows the proportion of CDD retained at the incorrect  $T_0$  (♦) for each waterbody and panel (f) shows the resulting apparent growth rates when using the 'incorrect'  $T_0$ . When CDD are calculated using 'true'  $T_0$ , immature growth rate is the same for both waterbodies (-). When CDD are calculated using an 'incorrect'  $T_0$ , it introduces an error in CDD that is larger in the cool waterbody than it is in the warmer waterbody; as a result, immature growth rate in the cool waterbody (•••) appears to be higher than immature growth rate in the warmer waterbody (short --).



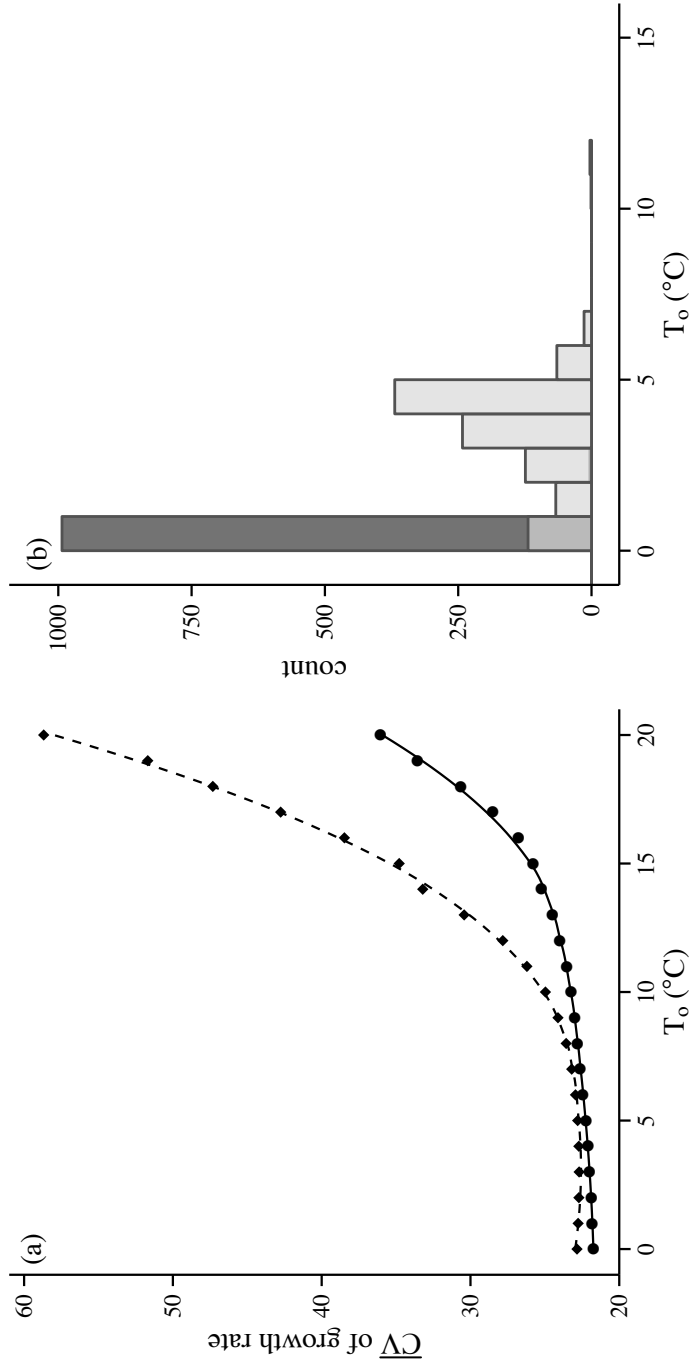
**Figure 3.2** Relationship between length and cumulative degree-days at  $T_0$  values of  $-5$  (dark grey),  $5$  (solid line), and  $15^\circ\text{C}$  (light grey) for four hypothetical waterbodies with mean annual temperatures of  $-7$ ( $\bullet\text{-}\bullet$ ),  $5$ ( $\bullet\bullet\bullet$ ),  $17$ ( $\text{-}\text{-}$ ), and  $29^\circ\text{C}$ ( $\text{-}$ ). At  $T_5$  ('true'  $T_0$ ), all 'populations' share the same slope (i.e., apparent growth rate). At  $T_{-5}$  and  $T_{15}$ , each 'population' has a unique slope. Populations appear to grow more quickly when assumed  $T_0$  is  $>$  'true'  $T_0$ , and more slowly when assumed  $T_0 <$  'true'  $T_0$ . These biases are smallest in the warmest waterbody and largest in the coldest waterbody.



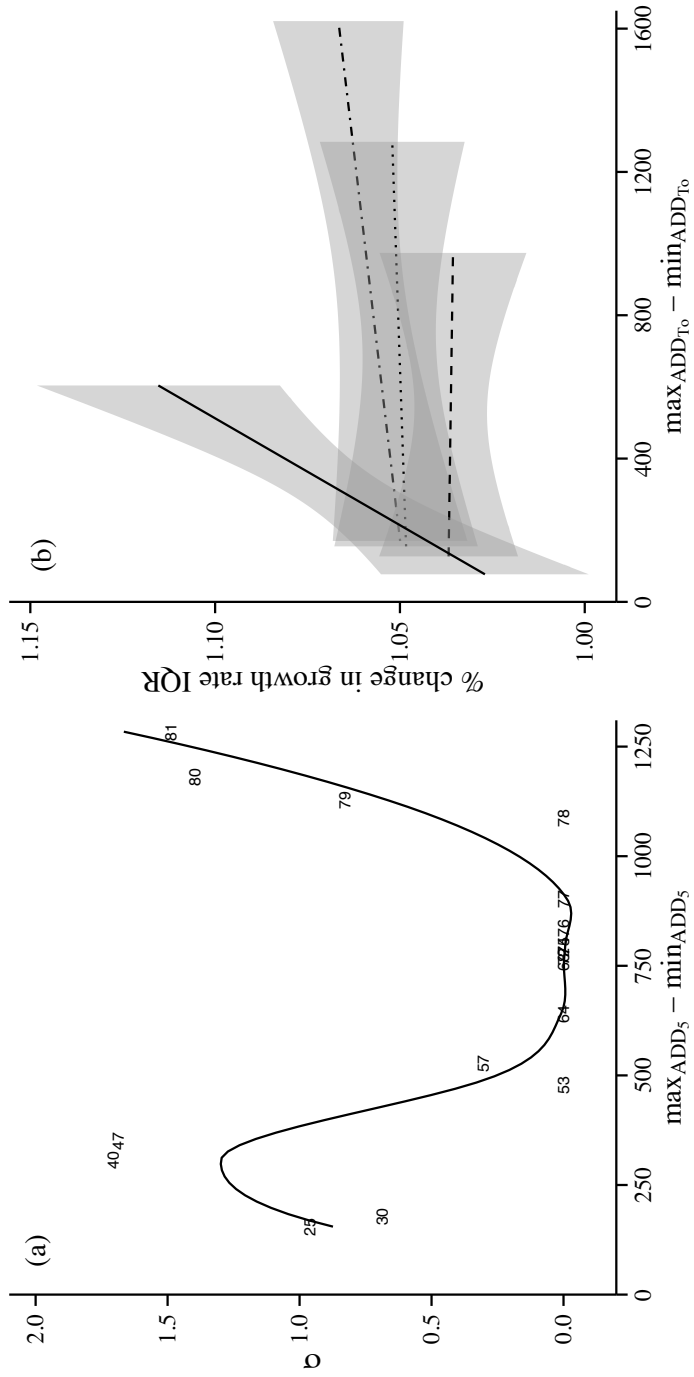
**Figure 3.3** Results of 1000  $T_0$  identification simulations over a small thermal range (7.3-7.9°C,  $\Delta\text{ADD}_5=180$ , [--]) and a large thermal range (0-15°C,  $\Delta\text{ADD}_5=1150$ , [-]). Panel (a) shows the mean among-waterbody coefficient of variation in apparent growth rate at each  $T_0$  over both thermal ranges. Panel (b) shows the distribution of  $T_0$  at which the CV of growth was minimized for both thermal ranges (dark grey, small thermal range and light grey bars, large thermal range, respectively).



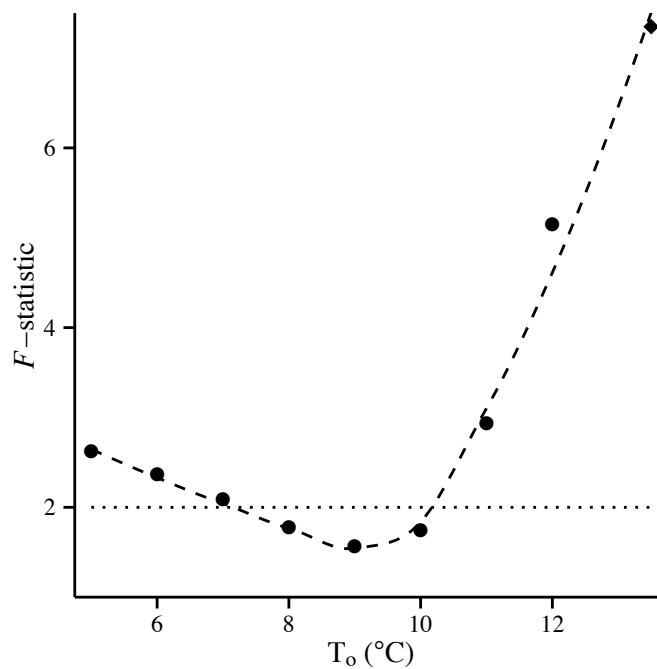
**Figure 3.4** Simulation results along a continuum of thermal ranges. Panel (a) shows how the standard deviation ( $\sigma$ ) of 1000  $T_0$  estimates change with increasing thermal range. Thermal range is described by ADD values at a  $T_0$  of  $5^\circ\text{C}$  but results were the same for all  $T_0$ . Panel (b) shows how the growth rate interquartile range (IQR) changes with increasing thermal range when using  $T_0$  values of  $0$  ( $\bullet\text{--}\bullet$ ),  $5$  ( $\bullet\bullet\bullet$ ),  $10$  ( $\text{---}$ ) and  $15^\circ\text{C}$  ( $\text{-}$ ). Shaded areas are 95% confidence intervals.



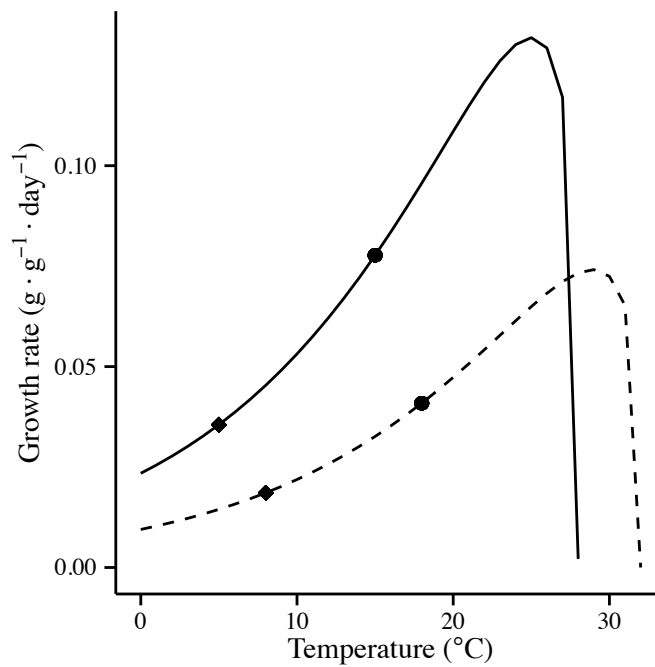
**Figure 3.5** Results for 1000 bootstrapped  $T_0$  estimates among walleye (*Sander vitreus*) populations spanning a large ( $\Delta ADD_5 = 1280$ ,  $n = 25$ ) and a small ( $\Delta ADD_5 = 155$ ,  $n = 81$ ) thermal range. Panel (a) shows the mean coefficient of variation of observed walleye growth rates at each  $T_0$  over a small (—, •) and large (---, ♦) thermal range. Panel (b) shows the distribution of  $T_0$  values given a small (dark grey) and large (light grey) thermal range.



**Figure 3.6**  $T_0$  estimation and apparent growth rates as a function of thermal difference among walleye populations in Minnesota and Ontario. Panel (a) shows how the standard deviation ( $\sigma$ ) of 1000 bootstrapped  $T_0$  estimates change with thermal difference among populations. Relationship depicted using LOESS smoother ( $\alpha = 0.7$ ) Point values indicate the number of populations within a thermal group. Panel (b) shows how the growth rate interquartile range (IQR) changes with thermal difference among populations when using  $T_0$  values of 0 ( $\bullet\text{--}\bullet$ ), 5 ( $\bullet\bullet\bullet$ ), 10 ( $\text{--}\text{--}$ ) and  $15^\circ\text{C}$  ( $\text{--}$ ). Shaded areas are 95% confidence intervals.



**Figure 3.7** The effect of  $T_o$  on significance (Chow Test) when comparing growth model coefficient estimates of yellow perch in northern Alberta, Canada, (3 populations) to West Blue Lake in southern Manitoba, Canada. Fish and temperature data for West Blue Lake were obtained from Wong (1972) and data from Northern Alberta were digitized from Power and van den Heuvel (1999). Missing temperature data from Sucker Lake were predicted using data from Kimowin Lake. The trend line (--) is via a LOESS smoother ( $\alpha = 0.85$ ) and the horizontal line indicates the significance threshold (•••). This re-analysis of Power and van den Heuvel (1999) shows that a significant difference in growth is detected at  $T_o > 10^\circ\text{C}$  and  $< 8^\circ\text{C}$  and these differences are more likely detected at high  $T_o$ . The original  $T_o$  used in Power and van den Heuvel (1999) was  $13.5^\circ\text{C}$  (♦).



**Figure 3.8** Application of the “10°C rule” for estimating  $T_0$  (Charnov and Gillooly 2003) to growth rates of juvenile walleye (-) and yellow perch (--) as predicted by the Wisconsin Bioenergetics model (Hanson et al. 1997). Dots are mean development temperatures (15 and 18°C for walleye and yellow perch, respectively) and diamonds are estimated  $T_0$  values (5 and 8°C, respectively).

## Chapter 4

### General discussion and future directions

Degree-days ( $DD_{T_o}$ ; °C·days) quantify the thermal experience of an organism and can be used to accurately and precisely describe growth and development in ectotherms.

Understanding the underlying mathematical and statistical properties of degree-days is paramount in appropriately using this metric to further elucidate biological processes such as fish growth. Accounting for these underlying properties allows for the discernment of results that reveal biological phenomena from those that are simply mathematical or statistical artifacts.

Research in the fields of agronomy and entomology has identified and accounted for some of the mathematical properties of degree-days. For instance, the ‘historical’ method for calculating a single degree-day (i.e.,  $DD = \left[ \frac{T_{Max} + T_{Min}}{2} \right] - T_o$ ) assumes the shape of a 24 hour temperature curve to be approximated by a rectangle and that an annual temperature curve is the aggregate of these rectangular degree-days (Legg 1998a); however, both daily and annual degree-days are better approximated by a sine curve (Arnold 1959). That said, using a sine curve to approximate a temperature curve can introduce a bias that varies geographically (Pruess 1983) due to extremes in humidity, temperature and variability in season length (Allen 1976). Although corrections for this bias have since been developed (e.g., Allen 1976), the relationship between degree-days and the fundamental mathematical underpinnings of the temperature curve (i.e., sine curve) itself has received limited attention.

Degree-day calculations incorporate a lower temperature threshold ( $T_o$ ) because ectotherm growth is negligible at low temperatures. These values are likely species-specific and may even be population-specific in cases where local adaptation has occurred. Research in entomology and agronomy has developed procedures for estimating lower temperature thresholds (e.g., Arnold 1959, Thomson and Moncrieff

1982, Lysyk 1989 and Legg 1998a) but  $T_0$  estimates for species and populations among and within methods remain somewhat variable. For instance, Yang et al. (1995) noted that Arnold (1959) reported a  $T_0$  of 6°C for corn in 1954 and a  $T_0$  of 4.3°C in 1955. Reported  $T_0$  are also quite variable in fish science; however, unlike entomology,  $T_0$  values are typically unjustified or loosely defined as the lower temperature limit of some physiological or abiotic process (e.g., Hamel et al. 1997, Venturelli et al. 2010).

To date, very little has been done to quantitatively define threshold temperatures in fish science, nor have studies been conducted to understand the effect of a given  $T_0$  on our ability to accurately describe growth. The objective of this study was to determine the effect of  $T_0$  on our ability to use degree-days to account for growth within and among populations. In this study, I used hypothetical and empirical temperature and fish data to show that, although degree-days from different  $T_0$  values account for variation in immature growth within a population nearly equally well, accurate  $T_0$  identification becomes important when comparing growth among populations that are distributed across a wide thermal range. These results stem from the underlying mathematical and statistical properties of degree-days. Specifically, interaction between  $T_0$  and the sine curve.

In Chapter 2 I used hypothetical and empirical data to explore the importance of  $T_0$  when describing growth within a population. I first demonstrated that degree-days at different  $T_0$  values are highly correlated when  $T_0$  values are low, but become increasingly dissimilar as  $T_0$  nears the maximum annual temperature. I then gathered length-at-age data for eight species, collected by the natural resource agencies of Ontario and Minnesota, and demonstrated that this correlative property of degree-days at low  $T_0$  is why degree-days at low  $T_0$  are all able to account for much of the variation in immature growth within a single population. Simply, because degree-days are very similar among low  $T_0$ , they account for growth almost equally well. However, at high  $T_0$ , degree-days become increasingly dissimilar and their ability to account for growth declines similarly. Importantly, I demonstrated that this correlative property is directly a result of the shape of the temperature curve (which is essentially a sine curve), and is why effective  $T_0$ .

values have little to do with growth limitations at temperatures approaching 0°C.

In Chapter 3 I demonstrated that, although  $T_0$  matters little when accounting for growth within a single population,  $T_0$  becomes increasingly important when comparing growth among populations that have very different temperature curves. Initially, I demonstrated the affect of  $T_0$  on apparent growth by building model populations that experienced similar temperature dependent growth but had very different thermal experiences. By then corroborating the results of these simulations with empirical analyses using immature walleye (*Sander vitreus*) length-at-age data spread throughout Minnesota and Ontario, I was able to show that i) growth rates become increasingly biased as our chosen  $T_0$  becomes increasingly dissimilar from the ‘true’ value and ii) this bias increases with increasing thermal differences among comparative populations. This affect of  $T_0$  on growth rates is a direct result of the proportional inequalities that exist among populations with disparate temperature curves. These results have significant implications for studies comparing growth over large spatial scales because an inappropriate  $T_0$  value can lead to artificial apparent differences among populations over a latitudinal gradient (i.e., the appearance of counter-gradient growth).

Although this study has clarified some of the mathematical constraints of the degree-day, more research will be required in order to maximize its utility in fish science. Initially, further work is needed to determine how degree-days derived from air and water temperatures are correlated and how this relationship changes with  $T_0$  and latitude. As latitude increases, the relationship between air- and water-derived degree-days may become non-linear. This could result in an artificial latitudinal affect on apparent growth that is similar to using an inappropriate  $T_0$ . Similarly, although immature fish growth and degree-days are linearly related when viewed on an annual time-step, it is unclear if this remains true when incorporating mid-season growth data. Fish growth is well understood to be linear over a midrange of temperatures, but it becomes non-linear at the extremes (e.g., Edsall 1993). Because of these non-linear regions, it is unclear whether growth continues to aggregate in a linear manner with degree-days over the entire course of a season. Lower and upper temperature thresholds may be required to ensure a linear model

that is not confounded by the curtailing of growth at the thermal limits of a species.

Additional effort is also needed to further standardize degree-days in fish science. Although this study takes the first steps by advising the use of standard  $T_o$  values (i.e., 0, 5, 10, and 15°C), standard methods for identifying  $T_o$  should also be developed. Method standards should include statistical power requirements (e.g., size of  $n$  and extent of thermal range) for estimating  $T_o$  at various developmental stages. Furthermore, standardized methods should take into account and validate the findings of prior research conducted in terrestrial systems. For instance, Legg (1998b) showed that the least variability method for lower threshold estimates is biased towards low  $T_o$ . An example of a least variability method is the iterative search for the  $T_o$  that minimized the coefficient of variation in growth rates among waterbodies (Chapter 3) and  $r^2$  method (Chapter 2). Findings such as these should be validated in aquatic systems. Moreover, although this study offers a method for converting degree-days among  $T_o$  (Chapter 2), these equations are unable to convert among all  $T_o$  and are limited by thermal range. Given that temperature curves are well approximated by a sine curve (Arnold 1959), it should be possible to use a sine curve to convert over a wider latitudinal gradient. This would facilitate standardization and thus future comparative studies.

Apart from the mathematical, statistical and methodological research that is required, we also need to better understand the relationships between the degree-day approach and the biological systems that it measures. For instance, understanding how well air temperature accounts for growth in cold-water species may reveal the significance of direct (i.e., metabolism) and indirect (i.e., primary production) effects of temperature on growth. Furthermore, very little is known about species-specific  $T_o$  values in aquatic systems. This includes determining whether general species characteristics, such as thermal preference (Hokanson 1977, Magnuson et al. 1979), can be used to group species by standardized  $T_o$  values, and if the “10°C rule” (Charnov and Gillooly 2003) provides a reasonable estimate of  $T_o$ . Similarly, it is unclear if  $T_o$  changes with developmental stage within a species. Charnov and Gillooly (2003) suggest that because  $T_o$  falls 10°C below the mean developmental temperature, theoretically the values are

stage dependent. Yet it is unclear how variable these values are and whether a standard  $T_o$  would be equally informative.

By understanding the specificity and implication of  $T_o$ , it becomes possible to quantify the number of degree-days to different developmental stages, allowing for interesting conspecific and heterospecific comparisons of growth and development over spatial and temporal scales. For instance Neuheimer and Grønkjær (2012) determined that the number of degree-days to maturation has declined in North Atlantic cod (*Gadus morhua*) over the past three decades. Using degree-days to account for annual temperature variability, the study concluded that the decline in age at maturation was due to intense fishing pressure, and is an example of fisheries-induced evolution. Pursuing studies such as these would greatly enhance growth models and our understanding of climate on fish biology.

Given that this study focused on temperate climates and freshwater habitats, it is unclear how transferable my findings are to arctic, tropical or marine systems. In environments that are thermally stable and uniform (e.g., tropical), calendar time and degree-days tend to explain growth and development approximately equally well (Neuheimer and Taggart 2007). This equality arises because in thermally stable and uniform environment, temperature variability is low such that degree-days aggregate parallel to calendar time. Similarly, as the thermal environment over a species range becomes increasingly uniform (i.e., tropics and arctic), the affect of  $T_o$  on growth rates among populations is likely to become unobservable and  $T_o$  identification less important., as differences in apparent growth rates brought about by  $T_o$  are a result of differences in the temperature profiles among populations. Species distributed longitudinally are likely to see greater thermal uniformity than those distributed latitudinally. Unlike freshwater fish, which are confined to discrete inland waterbodies, marine populations are able to behaviorally thermo-regulate over great distances. Determining how similar the thermal experiences of marine populations are will determine the importance of such factors as  $T_o$  when comparing growth. This issue is even more complicated for anadromous species, which may experience extreme thermal differences during some life stages but not others.

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