

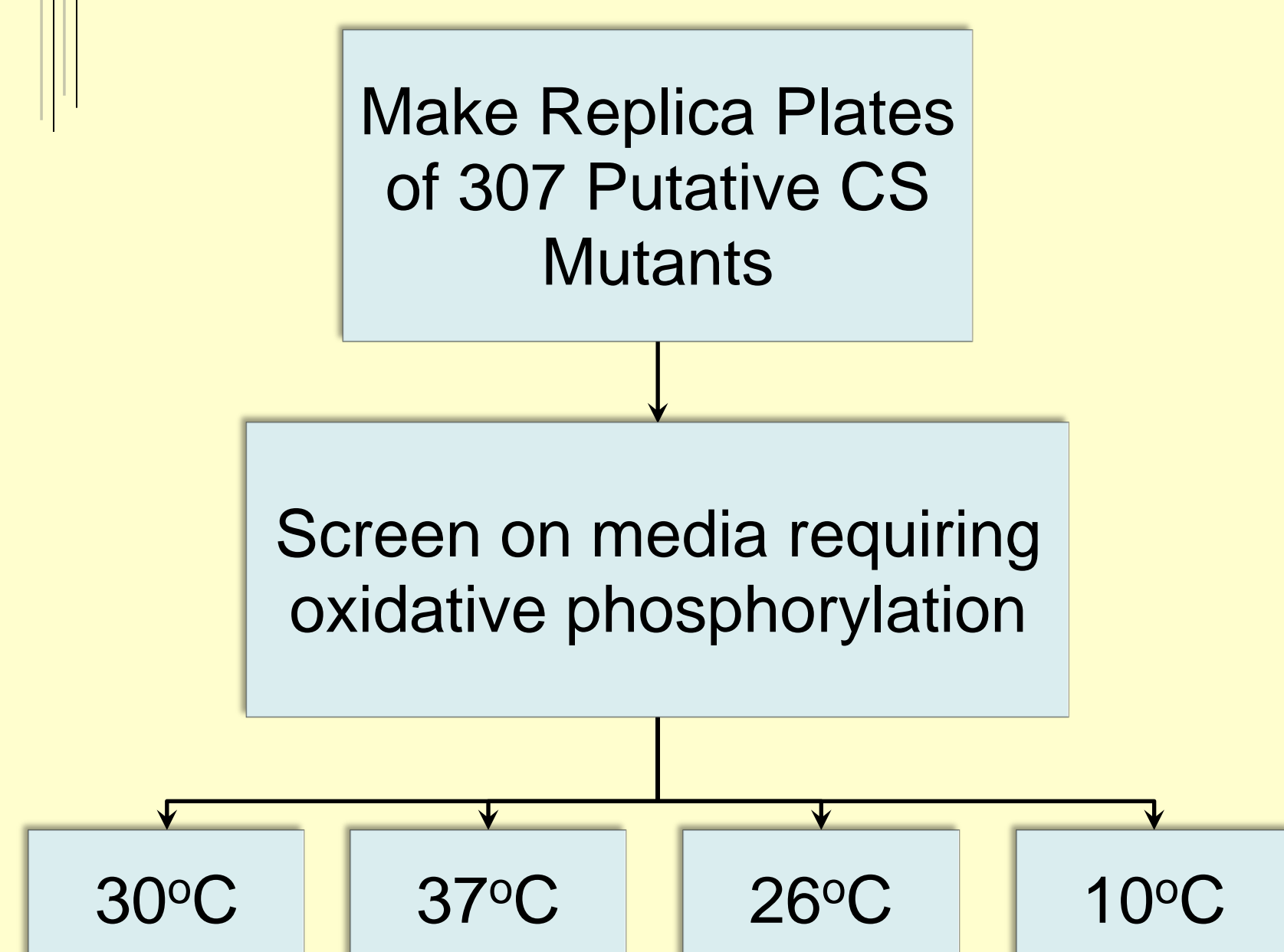
# Cold Adaptation & Mitochondrial Function in *S. cerevisiae*

Phoebe Leeaw - Genetics, Cell Biology & Development

Mentor: Dr. Robin Wright – CBS Department of Genetics, Cell Biology and Development

Is mitochondrial function important for survival in cold temperatures?

## Experimental Design



### Replica Plating: Velvet Stamp Technique

- Replica plates were produced by velvet stamping.
- Stamped by order of assigned temperature: 10°C, 26°C, 30°C, and 37°C.
- It is important to start stamping by lowest temperature first to ensure plates assigned to least favorable conditions contain maximum number of cells.

Temp.	Control (YPD)	Experiment (YPEG)
10°C	1	2
26°C	1	2
30°C	0	2
37°C	0	2

### Data Acquisition

- Each temperature treatment was photographed for two successive days once sufficient growth was observed.
- Day 1: Plates growing at 37°C plates were photographed
  - Day 2: Both YPD and YPEG plates growing at 26°C and 30°C were photographed
  - Day 13: Both YPD and YPEG plates grown at 10°C were photographed

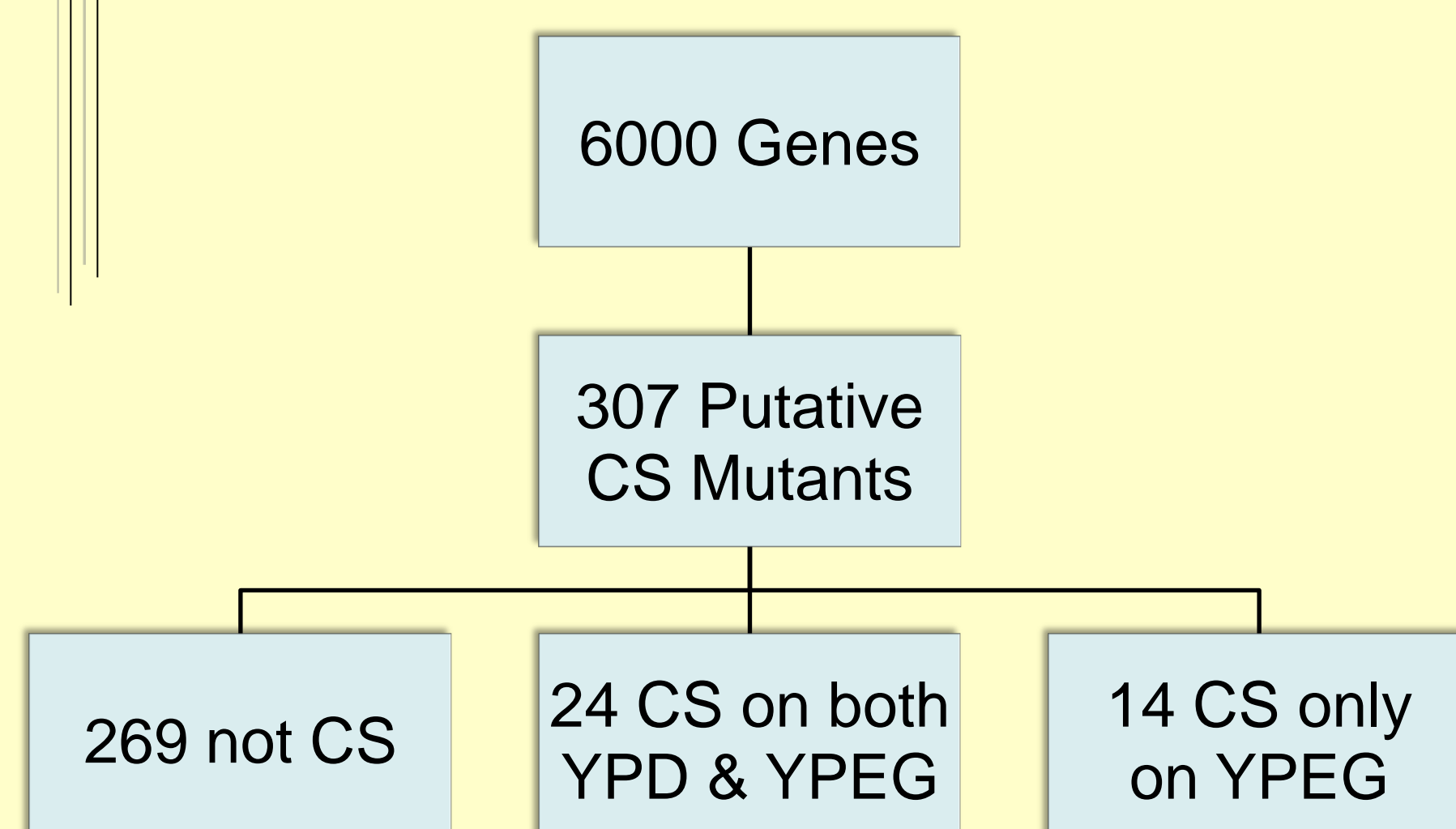
### Data Analysis

- The presence of growth on YPEG denoted that the deletion did not impede cellular respiration or other factors of cold adaptation.
- The absence of growth on YPEG denoted the deleted gene coded for a vital component in the mitochondria that also impacted cold sensitivity.

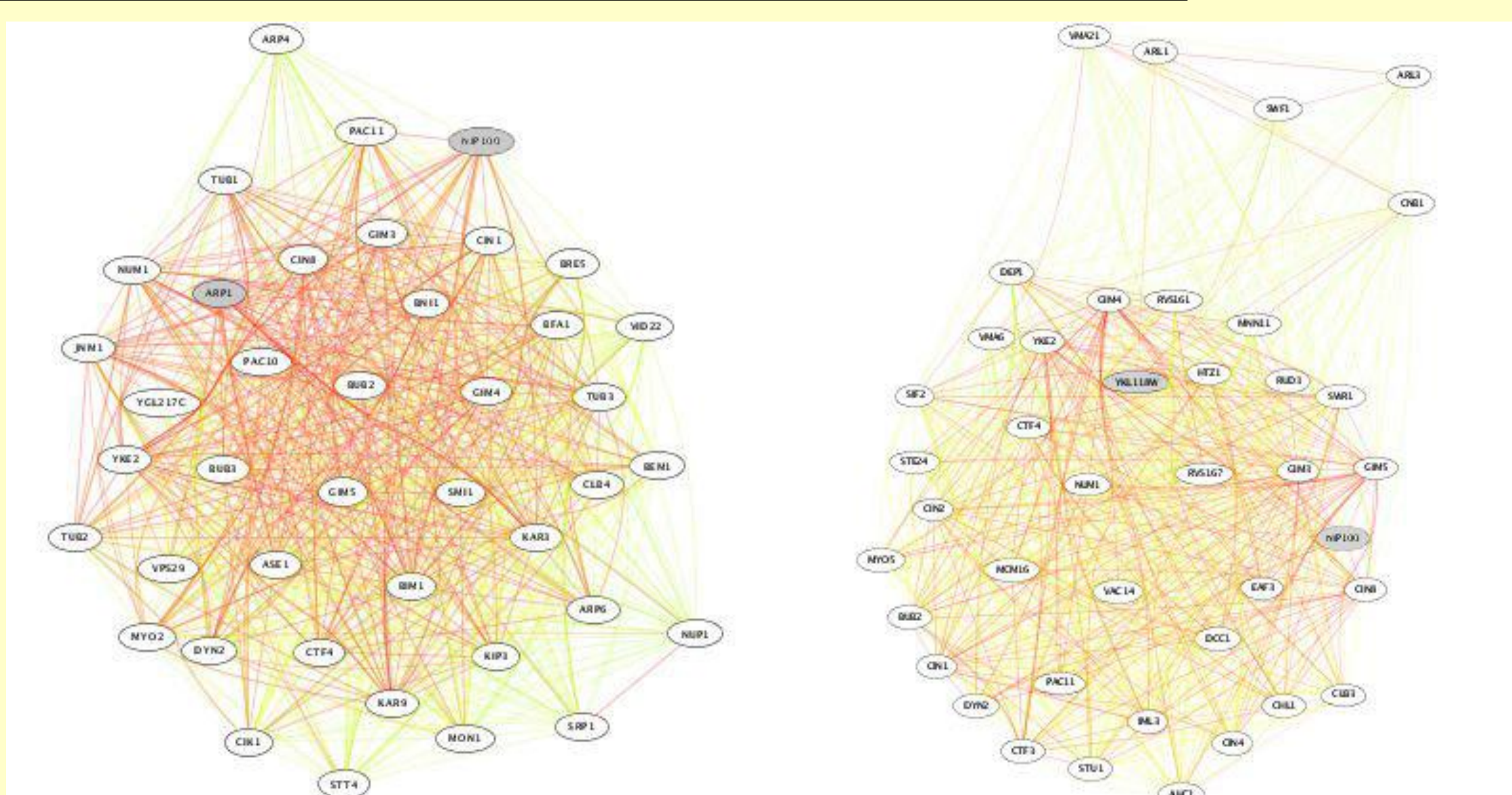
### Expected Results

- All mutants should have reduced or no growth at 10°C on YPD
- Mitochondrial mutants should have more drastic phenotype on YPEG than on YPD at 10°C

## Results



Identified 14 conditional mutants. These mutants utilize mitochondria in cold temperatures only



### Gene Association Chart

ARP1 and NIP100 strongly interact with each other.  
NIP100 and YKL118W have unknown relationship.

Gene	Control (YPD)	Experiment (YPEG)	Known Function
<b>Nuclear Function</b>			
SAC3			Pore-associated protein, forms complex with Thp1p, which is involved in transcription & mRNA export from nucleus
NUP188			Nuclear pore complex (NPC) subunit, involved in structural organization and permeability of envelope
MAC1			Transcription factor, induces genes required for reduction & utilization of Fe and Cu <sup>1</sup>
SNU66			Nuclear RNA splicing
IES6			Protein that associates with INO80 chromatin remodeling complex under low-salt conditions
ARP1			Arp1-actin short filament related to dynactin complex via linking cytoplasmic dynein to its cargo through p150(glued) <sup>2</sup> ; nuclear import
NIP100			mitotic spindle positioning protein in dynactin complex involved in partitioning mitotic spindle between mother & daughter cells <sup>1</sup> ; nuclear import
<b>Vacuolar Function</b>			
PEP3			Protein involved in vacuolar protein sorting <sup>1</sup>
VPH2			Vacuolar ATPase assembly protein that acts in ER <sup>1</sup>
<b>Cytosolic Function</b>			
SER1			3-phosphoserine aminotransferase, catalyzes formation of phosphoserine from 3-phosphohydroxypyruvate, needed for Ser & Gly biosynthesis
RMD8			Cytosolic protein required for sporulation
<b>Mitochondrial Function</b>			
YME1			Degrades improperly folded proteins in intermembrane space <sup>1</sup>
<b>Unknown Function</b>			
BUD28			Unlikely to encode protein
YKL118W			Overlaps with both NIP100 and VPH2 <sup>1</sup>

## Conclusion

Of the fourteen genes found to be cold sensitive on YPEG conditions, only one is directly related to mitochondrial function. The results suggest:

- the reduction of growth in YME1 is due to the dysfunctional mitochondria
- overall, reduced cell growth in the mutants was a result of errors in cell transportation or protein processing and maintenance
- genes other than YME1 may code for proteins that contribute to processes indirectly related to mitochondrial processes

**Acknowledgements:** Kelaine Zimmerman and Alicia Felthausen

### References:

- <sup>1</sup>Dimmer KS, Fritz S, Fuchs F, Messerschmitt M, Weinbach N, Neupert W, Westermann B. Genetic basis of mitochondrial function and morphology in *Saccharomyces cerevisiae*. *Mol Biol Cell*. 2002;13:847–53. doi: 10.1091/mbc.01-12-0588.
- <sup>2</sup>Hirokawa N. Kinesin and dynein superfamily proteins and the mechanism of organelle transport. *Science*. 1998;279:519–526. doi: 10.1126/science.279.5350.519.