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EVALUATION OF AN ECOLOGICAL NUTRIENT GRADIENT  
IN THE CENTRAL PINE SECTION OF MINNESOTA

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Robert Noyce Coats

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## I. INTRODUCTION

This study is part of a larger project on the characteristics of forest ecosystems and the foundations of forest productivity in Minnesota. The basic premise of the project is that ecosystems can be characterized by their positions in a four dimensional space with moisture, nutrients, heat and light as coordinate axes.

The first phase of the project involved a preliminary reconnaissance of Minnesota. Species lists, cover estimates and site descriptions were compiled. The second phase, of which this study is a part, involves the quantitative description of the multi-dimensional ecosystem space by means of field instrumentation, and analysis of soils and plants.

The purpose of this study was to define an ecological nutrient gradient in terms of synecological coordinates and to evaluate the relationship between mineral soil and humus nutrients, foliar nutrients in herbaceous plants and synecological coordinates along the chosen gradient. Three hypotheses were formulated. First, synecological coordinates are positively correlated with humus, soil and foliar nutrients. Second, the species that segregate at the lower end of a nutrient gradient have a lower average foliar nutrient concentration than those

at the upper end. Third, plants of a species growing in an infertile stand have a lower foliar nutrient concentration than plants of the same species growing in a fertile, nutrient-rich stand.

It was hoped that this study would lead to a better understanding of the relationship between synecological coordinates and the nutrient relation of forest ecosystems. This would enable foresters and silviculturists to assess rapidly the fertility or potential productivity of forest sites without having to rely solely on site index or costly chemical analysis of soil and plant material.



## II. BACKGROUND OF THE STUDY

### 1. Synecological Coordinates, Ecological Groups and Environmental Gradients

The synecological coordinate method is part of the general method of ecological groups. According to the definition of Ellenberg (1956), an ecological group is a group of species which are similar in their relationships to major site factors, such as moisture, light, etc.

Ellenberg constructed his ecological groups in the following manner: First, a relative value between one and five was assigned to each species for each factor, based on the predominant occurrence of the species. The community value was then computed for each factor by taking the average of the individual values of the species present. The value for each species was adjusted according to the average of the stands in which it occurs. The community values were then recomputed and the community assigned a relative position along a qualitative environmental gradient.

The method of Bakuzis (1959) is similar to that of Ellenberg. Bakuzis, however, considers all species, including ubiquitous plants, whereas Ellenberg considers only the species

that appear to respond to effective environmental factors.

An example will illustrate the method of synecological coordinates. According to literature sources, Pinus banksiana<sup>1</sup>, Gaultheria procumbens and Maianthemum canadense occur on infertile, acid sites and are therefore assigned a nutrient coordinate value of one. If only these three species occurred on a plot, then the unadjusted synecological nutrient coordinate for that plot would be one. The adjusted value for each species, however, is taken from an extensive regional survey as the average of the plots on which that species occurred. Since Maianthemum canadense in Minnesota also occurs frequently with species characterizing rich sites, the adjusted nutrient coordinate is close to two. Thus the adjusted value for each species is based on its association with all other species. It is important to note that the coordinate values do not necessarily reflect the physiologic requirements of the species; rather they reflect the occurrence of the species under competition.

Rowe (1956) used a method similar to that of Bakuzis to develop a vegetation moisture index. Understory species were classified according to position in the vegetation strata and common occurrence along a moisture gradient. The plants were assigned to ecological moisture groups on a basis of frequency and dominance data and given values (wet to dry) of 1, 2, 4, 8 and 16. The geometric increase was used because of the narrower

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<sup>1</sup>The botanical names follow the Latin nomenclature as given in the eighth edition of Gray's Manual of Botany (Fernald, 1950).

ecological amplitude of species on wet sites. Ubiquists were omitted and the value assigned to each species was not adjusted as was done by Bakuzis.

The method of environmental gradients is closely related to the method of ecological groups. This approach is based on quantitative measurement of effective environmental factors, coupled with vegetation analysis. The term "gradient" usually refers to a ranking of stands or sites on a basis of some environmental factor, although in some cases variation along a transect is implied. Once the factor gradient has been defined, then either individual species or ecological groups can be ordinated along it.

A number of authors have constructed synthetic regimes for expressing environmental factors. Whittaker (1960), for example, defined a moisture gradient in terms of topographic position. He then grouped species into four different moisture classes on a basis of the maximum population levels along the moisture gradient and used the moisture class of each species to compute a weighted average for each stand. This was done separately for each parent material and elevation class. Whittaker's method is thus basically floristic; the moisture classes are not based on quantitative environmental measurements.

Locks (1962) developed synthetic gradients for moisture, nutrients and local climate. Each gradient or "scalar" was constructed from several different effective components. The synthetic nutrient regime, for example, was constructed separately

for organic and inorganic soils. For inorganic soils the parameters used were thickness of the  $A_1$  horizon, depth of the solum and percentage of silt and clay. The nutrient regime for organic soils was based on organic constituents, depth of decomposition and run-off. Each of these factors was found to correspond to a species composition gradient. Having constructed these three synthetic factor gradients, Loucks then plotted the importance values of individual species with moisture, nutrients and local climate as coordinate axes.

A more quantitative approach to environmental gradients was used by Waring and Major (1964) in the Coast Range of northern California. They developed moisture, nutrient, heat and light gradients on a basis of field and laboratory measurements. The environmental moisture gradient was based on the minimum value of available soil moisture during one year. The environmental nutrient gradient was based on exchangeable calcium per  $m^2 \times 30$  cm. This variable was selected for its high correlation with redwood and barley yield in pot tests. Soil samples were also analyzed for nitrogen, phosphorus, potassium and magnesium. These nutrients were also significantly related to fertility, but given the calcium concentration they did not add much information.

Light was measured over a one-day period using anthracene, and the ecological light gradient was based on percent of full sunlight. Heat was not measured directly but was evaluated from temperature records for weather station occurring in vegetation zones that corresponded to the zones covered in the study.

With the environmental gradients thus defined, the segregation of individual competing species along the respective gradients was shown. Species were then placed in ecological groups according to their relative positions along the four factor gradients, and an index number was assigned to each group. On a basis of the index values for each species, the average moisture, nutrient, heat and light values for each stand were computed. This method is similar to that of Bakuzis, except that the ecological groups of Waring and Major are based on field instrumentation, whereas those of Bakuzis are based on plant associations. An environmental nutrient gradient may be defined as a ranking or gradation of stands on a basis of soil fertility. An ecological nutrient gradient is thus defined as a ranking or gradation of plant response to an environmental or soil nutrient gradient. Plant response in this case includes presence or absence, growth and development under competition and uptake of nutrients. Response may be defined in terms of individual species or ecological groups.

In the previous work on environmental gradients investigators have seldom used the chemical composition of trees, shrubs or herbaceous plants to characterize or define a nutrient gradient. It was hoped that this study would shed light on this possibility and provide detailed information on the effective variables that comprise an ecological nutrient gradient.

## 2. Foliar Analysis

### a. Foliar analysis in general. The technique of using

the nutrient concentration of plants as an index of their nutritional status is old and widespread. The advantage of foliar analysis over soil analysis is based on the fact that the growth of plants is related not to the concentration of nutrients in the soil, but rather to the concentration of nutrients in the plant (Lundegårdh, 1951). Thus the plant itself rather than the investigator does the extracting from the soil. This overcomes the problem of choosing a technique that will measure the nutrients actually available for plant growth.

As work on foliar analysis has accumulated, however, the pitfalls and complications have become increasingly clear. First, the concentration of a given nutrient in plant tissue is a function not of the supply of that one element but rather of the total supply of all elements (Coddall and Gregory, 1947). With an increase of one or more nutrients, the growth of the plant may increase, thus decreasing the foliar concentration of one or more nutrients. This is the so-called "dilution effect." This principle applies not only to nutrients, however, but to other growth factors as well. Thus if the growth of a plant is depressed by lack of available water, heat or light, the concentration of nutrients may be increased. Furthermore, if a given element is available in excess relative to other factors, the uptake and foliar concentration may exceed the physiologic requirements of the plant. This is commonly called "luxury consumption."

Aside from direct environmental effects, the internal concentration of nutrients is strongly influenced by the growth

and developmental cycle of the plant. During the period of vegetative growth, the concentration of nutrients generally decreases whereas the absolute weight in a given organ increase. Mitchell (1936), for example, found a decrease over time in the foliar concentration of nitrogen, phosphorus, and potassium in oak, maple and hickory leaves. The concentration of calcium, however, increased during the growth period.

The processes of flowering and fruiting result in a decrease in foliar nutrient concentration. Lundegårdh (1951), for example, found a decrease in foliar potassium during flowering of clover. Goodall and Gregory (1947) reported numerous similar examples.

Most of the research on foliar analysis has been concerned with agricultural problems. Foliar analysis is frequently used for assessing fertilizer requirements of crop plants, either by sampling crops from the field or growing crop plants in soil samples under controlled conditions. Less frequently has foliar analysis been used in ecological situations.

b. Ecological applications of foliar analysis. The ecological applications of foliar analysis fall into two groups. The first of these is concerned with the role of forest litter and subordinate forest plants in mineral cycling. Scott (1955), for example, analyzed the subordinate forest plants and leaves of forest trees for thirteen elements. He found that the litter from subordinate plants contained almost twice the concentration

of potassium as the litter of tree species. He concluded that understory plants play an important role in the mineral cycle of a forest.

Madgwick (1965) analyzed 19 understory species in a Fraxinus woodland and expressed nutrients on both a percent and area basis. He found that the foliage of understory plants contained between 37 and 44 percent of the potassium, calcium, magnesium, phosphorus and nitrogen of the total for all of the foliage in the stand.

P'yavchenko (1960) measured the nutrient content of mosses, shrubs and grass in a bog forest. He found that these understory plants cycled a larger quantity of calcium and nitrogen annually than did the overstory of spruce, fir and pine.

Whereas mineral cycling is concerned primarily with the effect of herbaceous and tree litter on forest soils, some research has been devoted to the effect of soil differences on nutrient uptake by trees and herbs. Bard (1945) investigated the nutrient content of several tree species growing on three different soil types. He found that the calcium content of the leaves was a function more of species than of soil type. Phosphorus, however, was related both to species and soil acidity; trees on the more acid soils had a higher foliar concentration of phosphorus. This was attributed to the greater availability of phosphorus in the acid soils. The potassium content also seemed to be positively correlated with supply in the soil as well as species. Nitrogen content of the foliage, however, was a function only of species.



In 1949, Bard followed up this study with a similar investigation of herbaceous species. The general pattern for the herbaceous species followed that of the trees. Calcium concentration was controlled by species rather than soil type, whereas phosphorus concentration in the foliage was correlated with available soil phosphorus. No clear trends were shown for potassium and nitrogen, although plants with high moisture requirements had the highest potassium concentrations.

The findings of Ovington (1956) tend to contradict those of Bard. Ovington found that plants on calcareous soils were higher in calcium than those on acid soils. He did not, however, consider individual species, but lumped all herbaceous species on each plot. Forest plants had a higher percentage nutrient content than plants growing in the open; Ovington attributed this to enrichment of the forest floor rather than differences in light or moisture.

Ovington found no significant differences in the nutrient content of plants associated with hardwood and conifer forests on the same soil type. In some instances, however, the overstory species seemed to exert a strong influence on the chemical composition of the ground flora. For example, the Urtica growing beneath Alnus was high in nitrogen, potassium, calcium and magnesium.

Although most of the studies on the nutrient content of forest plants have been largely descriptive, some authors have used this technique to study plant distribution or to assess

gradients in nutrient availability and forest productivity.

Pigott and Taylor (1964), for example, studied the distribution of Urtica dioica in competition with other herbs. They found a high foliar concentration of both phosphorus and nitrogen in Urtica foliage. With fertilizer and plot studies they showed that the distribution of Urtica was controlled by the amount of available soil phosphorus; that is, under competition, Urtica segregated out at the upper end of a gradient of soil phosphorus.

Olsen (1961) analyzed soils and foliage along a transect from a Danish beech-wood into a chalk barren. He found an increase in foliar calcium and decreases in phosphorus, nitrogen, potassium and iron. He concluded that competition for the limited nutrient supply was responsible for the exclusion of beech from the glades.

Gagnon et al. (1958), working in Canada, attempted to relate the nutrient content of understory forest plants and humus layers to forest site quality. They analyzed two herbs, four shrubs and four mosses for calcium, potassium, phosphorus, magnesium, iron and manganese. They also analyzed humus samples for the same elements. The results of the foliar analysis were disappointing; no significant differences were found in nutrient content within species among sites. Large differences among species were found. The nutrient content of the humus layer, however, was found to be highly correlated with site index. This was explained in part by the dominance of Oxalis and Cornus on the better sites. These species have a higher cation content

than the shrubs and mosses and thus form a more base-rich humus.

### 3. General Description of the Area

Data for this study were collected during the summer of 1966 in and around Itasca State Park, in the Central Pine Section of Minnesota (Balazis, 1961).

The present upland vegetation of the Itasca region is a mosaic of hardwood and conifer stands, consisting chiefly of northern hardwoods, aspen-paper birch, red pine and jack pine. Both fire and logging have played important roles in the history of these stands.

Itasca Park lies within 40 miles of the prairie-forest border. This suggests that moisture stress on upland sites is probably an important limiting factor in the distribution of vegetation. This is especially likely in view of the history of the climate and vegetation of the region.

The Itasca area was glaciated in late Wisconsin time, the last ice having left the area 12,000 to 13,000 years B. P. (before present). From about 12,000 B. P. to 8,500 B.P. the region was dominated by a Picea-Fopulus boreal forest. This was replaced by oak savanna, which in turn was succeeded by a mesic deciduous forest about 4,000 B.P. This forest was invaded by pine about 1,000 to 2,000 years ago (McAndrews, 1966).

The present climate is characterized by warm humid summers and cold dry winters (see Figure 1). The mean annual precipitation is 26.3 inches and the mean annual temperature is 38.7°F

(Kurmis, 1967).

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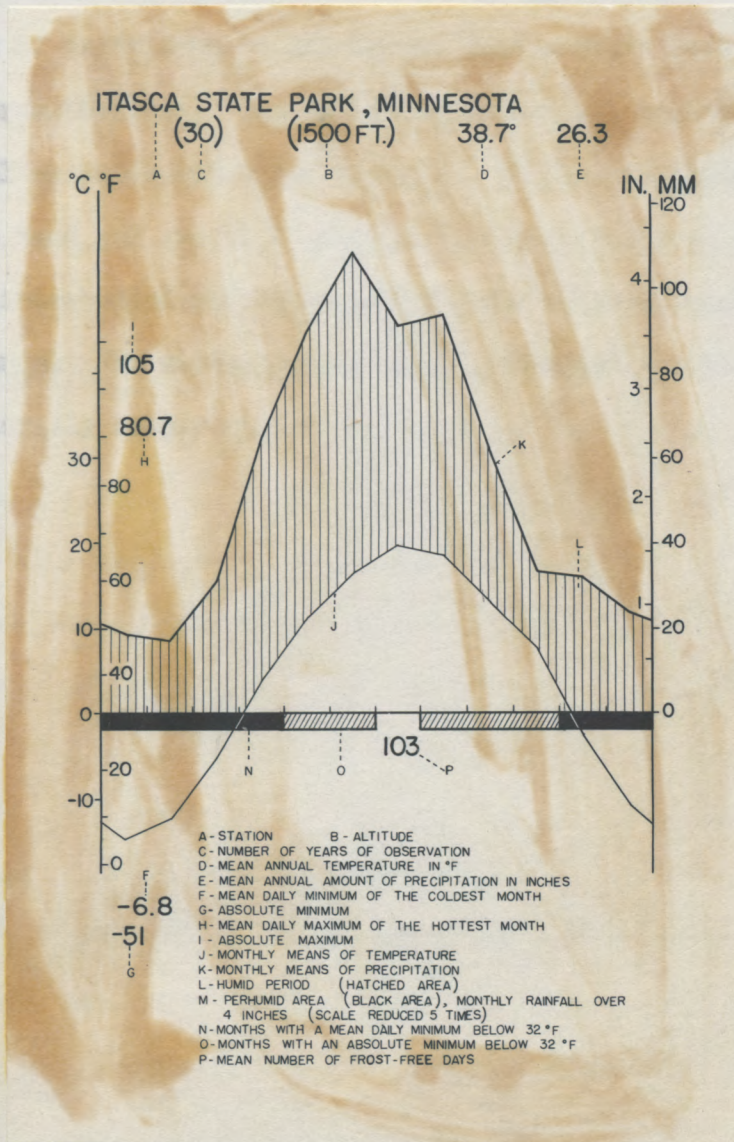


Figure 1. Walter's (1961) climatic diagram for Itasca State Park, Minnesota. The diagram was compiled and constructed by Kurmis (1967).

(Kurnis, 1967).

The role of soil as a controlling factor in the distribution of vegetation in and around Itasca State Park can hardly be over-emphasized. The upland soils are all derived from glacial or water-deposited material, varying in texture from coarse sand and gravel to loam. Different textural classes are often interbedded with one another, and extreme local variation is common. The mineralogy is mixed, although the parent material is generally calcareous.

### III. METHODS

#### 1. Experimental Design

Six stands were selected, two in each of three forest types: jack pine (Pinus banksiana), red pine-balsam fir (Pinus resinosa-Abies balsamea) and northern hardwoods. Stands were selected on a basis of uniformity of soils and vegetation and lack of dominance of a shrub layer. An attempt was made to find stands that represented a broad range of nutrient conditions as approximated by the synecological nutrient coordinates. The defined gradient in this case represented a ranking of stands rather than a continuous gradation. Since the "gradient" included both hardwood and conifer stands, large discontinuities in plant response were expected.

Table 1 shows the location and code name for each plot. The names were arbitrarily taken from local place names or geographic features. The two jack pine stands are located in the Lake George Township; otherwise all of the plots are in Itasca State Park. Figures 2-1 through 2-6 show the general aspects of the stands chosen for this study.

Within each stand a plot four chains by two chains (264 feet by 132 feet) was delineated. Eight subplot centers were then located systematically within each plot and marked with stakes. Each stake became the center of a nest of concentric

TABLE 1

## PLOT LOCATIONS AND FOREST TYPES

Plot Name	Location	Forest Type
Woodlot	S.E. $\frac{1}{4}$ N.W. $\frac{1}{4}$ Sec. 16, T143N, R34W	jack pine
Scott's	S.E. $\frac{1}{4}$ N.W. $\frac{1}{4}$ Sec. 21, T143N, R34W	jack pine
Mary Lake	N.E. $\frac{1}{4}$ S.W. $\frac{1}{4}$ Sec. 19, T143N, R36W	red pine- balsam fir
Wilderness Sanctuary	S.W. $\frac{1}{4}$ S.E. $\frac{1}{4}$ Sec. 16, T143N, R36W	red pine- balsam fir
Campground	S.W. $\frac{1}{4}$ N.W. $\frac{1}{4}$ Sec. 12, T143N, R36W	maple-bass- wood
Bearpaw Point	S.W. $\frac{1}{4}$ N.E. $\frac{1}{4}$ Sec. 11, T143N, R36W	basswood- elm

subplots with areas of 1, 2, 4, 8, 16, 32, 64, and 128 square meters. This method was chosen to maintain consistency with the method used by Bakuzis (1961).

Species presence was noted for each subplot through eight square meters. These data were used for computing the synecological coordinates for each subplot. Species cover percent was estimated for the eight square meter subplots. Synecological coordinates on a dominance basis were computed by weighting each species with its assigned cover estimate; the weighting factors used were 0.1, 0.5, 1.0, 17.5, 37.5, 62.5 and 87.5. Shrubs were counted and classified according to species, age and height on each subplot through eight square meters. Live trees, dead trees and stumps were counted for all subplots. Diameters of trees one inch or greater were measured on all subplots through 32 square meters; diameters of trees four inches or greater were measured for the 64 and 128-square meter rings. The tree data were used for volume computations.

Within each stand, heights and diameters were measured on a sample of trees. These data were used to prepare height-diameter curves for volume estimates. Borings for age determinations were taken from dominant and codominant conifers for site index calculations (Gevorkiantz, 1957).

Two soil pits were dug in each stand. The profiles were described according to the procedures of the Soil Survey Manual (Soil Survey Staff, 1951). Samples from each horizon were collected, air-dried and stored for future analysis. Bulk density





Figure 2-1. Woodlot jack pine stand. Note the uniformity of the stand and lack of a shrub layer.



Figure 2-2. Scott's jack pine stand. The canopy of this stand is more open than that of the Woodlot stand. The well developed shrub layer is chiefly Corylus cornuta.



Figure 2-3. Mary Lake red pine-balsam fir stand. Note the wind-throw in the background and the discontinuous distribution of the herb layer.

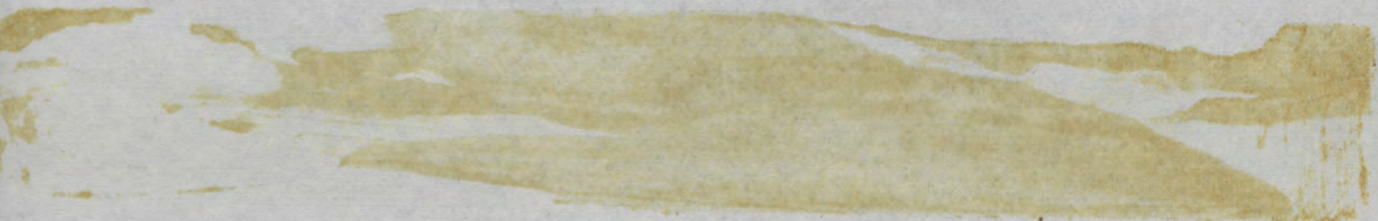




Figure 2-5. Campground maple-basswood stand. Note the single red pine in the background, right.



Figure 2-6. Bearpaw Point basswood-elm stand. One of the soil pits appears at the right.

measurements were taken in each horizon with a core sampler.

It was planned that data from the soil analysis would be converted to an area basis. One quarter cubic foot samples were taken from three profiles so that the gravel fraction could be estimated.

Foliage samples were collected twice. The first sampling was done in the middle of July. Approximately one kilogram (wet weight) of plant foliage was collected for each of four ubiquitous species in each stand. The species collected were Aster macrophyllus, Aralia nudicaulis, Maianthemum canadense and Corylus cornuta. Because of a scarcity of material, no Maianthemum was collected from the Bearpaw Point Stand, and neither Aralia nor Corylus were collected from the Woodlot stand. All samples were collected along the margins of the delineated plots. The plant material was dried within 4 hours of collection at 90° C and stored in paper bags.

The major sampling was done during the third week of August. In most cases, plant material was collected on the eight square meter subplots. For a few species, however, not enough material was available on the subplots, and so additional material was collected elsewhere in the stand. The amount collected for each species ranged from about twenty grams to one kilograms, depending largely on the quantity available. The samples collected in August were dried at 65° within 4 hours of collection.

Table 2 shows the species sampled in each plot, the stage of development at the time of the second sampling and the part taken. It should be noted that the same species was sometimes in

TABLE 2  
FOLIAGE SAMPLING METHOD

Species	Developmental Stage	Part Taken
1. Bearpaw Point		
<i>Aralia nudicaulis</i>	past fruiting	stems broken at base; stems and leaves (lvs) collected
<i>Maianthemum canadense</i>	healthy; most plants senescent	lvs and petioles above 3rd node; lvs adjacent to fruit not taken
<i>Celastrus scandens</i>	small, heavily browsed	lvs only collected
<i>Dirca palustris</i>	yellow spots on some lvs	lvs only were collected
<i>Corylus cornuta</i>	nuts ripening	lvs from top of canopy
<i>Actea rubra</i>	fruit ripening	lvs and petioles above 2nd node
<i>Aster macrophyllus</i>	not yet flowering	basal lvs from sterile plants
<i>Thalictrum dioicum</i>	past fruiting	lvs and petioles above 3rd node
<i>Amphicarpa bracteata</i>	healthy	lvs only
<i>Asarum canadensis</i>	healthy	stems and lvs above base
<i>Uvularia grandiflora</i>	capsules broken	lvs and stem above lowest leaf; capsules not taken
<i>Sanguinaria canadensis</i>	some plants senescent	healthy plants only; stems and lvs above ground level
<i>Smilacina racemosa</i>	fruit ripe	lvs only; fruit not taken
2. Campground		
<i>Uvularia grandiflora</i>	capsules broken; some plants senescent	lvs and stem above lowest leaf; mostly healthy plants



TABLE 2---Continued

Species	Developmental Stage	Part Taken
<i>Hepatica americana</i>	healthy	lvs and petioles above ground level
<i>Maianthemum canadense</i>	healthy, mostly sterile	as above
<i>Aster macrophyllus</i>	not flowering; some lvs chewed by insects	basal lvs only; broken at ground level
<i>Dirca palustris</i>	healthy	lvs only
<i>Clintonia borealis</i>	healthy; berries ripe	lvs above base
<i>Corylus cornuta</i>	healthy, sterile	as above
<i>Ostrya virginiana</i>	healthy	lvs from top of trees 6 ft high
<i>Acer saccharum</i>	healthy	lvs from reproduction 4.5 ft high
3. Wilderness Sanctuary		
<i>Maianthemum canadense</i>	many plants senescent; past fruiting	healthy lvs, mostly from sterile plants
<i>Aster macrophyllus</i>	some plants flowering	basal lvs from sterile plants only
<i>Aralia nudicaulis</i>	healthy; past fruiting	as above
<i>Corylus cornuta</i>	healthy	as above
<i>Parthenocissus inserta</i>	healthy; still growing	mature lvs only
<i>Lonicera canadensis</i>	healthy; past fruiting	lvs only
<i>Clintonia borealis</i>	berries ripe; some lvs senescent	lvs above base; sterile plants only
<i>Cornus canadensis</i>	berries ripe	lvs and stems above ground level; sterile plants only
<i>Hepatica americana</i>	healthy	as above

TABLE 2--Continued

Species	Developmental Stage	Part Taken
4. Mary Lake		
<i>Maianthemum canadense</i>	most plants senescent	mostly healthy plants taken
<i>Aralia nudicaulis</i>	healthy	as above
<i>Aster macrophyllus</i>	damaged by insects, otherwise healthy	as above
<i>Corylus cornuta</i>	brown spots on some lvs	as above
<i>Clintonia borealis</i>	healthy; berries ripe	as above
<i>Cornus canadensis</i>	berries ripe	sterile plants only; lvs and stems
<i>Hepatica americana</i>	healthy	as above
<i>Pteridium aquilinum</i>	healthy	fronds only; no stems
<i>Fragaria virginiana</i>	damaged by insects	leaflets only
5. Scott's		
<i>Maianthemum canadense</i>	berries ripe; many plants senescent	mostly healthy plants; no berries
<i>Aralia nudicaulis</i>	some plants senescent	healthy plants only
<i>Gaultheria procumbens</i>	both fruit and flowers present	lvs from non-fruiting plants only
<i>Arctostaphylos uva-ursi</i>	fruit ripe	lvs only
<i>Hepatica americana</i>	healthy	as above
<i>Aster macrophyllus</i>	healthy	as above
<i>Corylus cornuta</i>	healthy	as above
<i>Pyrola elliptica</i>	healthy	lvs above base of plant
<i>Diervilla lonicera</i>	healthy; fruiting	lvs only

TABLE 2---Continued

Species	Developmental Stage	Part Taken
<i>Epigea repens</i>	healthy	mature lvs
<i>Pteridium aquilinum</i>	healthy	fronds only
<i>Cladonia</i> sp.	healthy	picked clean of litter
<i>Pleurozium schreberii</i>	healthy	picked clean of litter
6. Woodlot		
<i>Lithospermum canescens</i>	fruit mature	whole plants; no fruit
<i>Maianthemum canadense</i>	mostly healthy	as above
<i>Aralia nudicaulis</i>	healthy but small, mostly sterile	as above
<i>Aster macrophyllus</i>	mostly sterile	as above
<i>Frageria virginiana</i>	psat fruiting	as above
<i>Gaultheria procumbens</i>	fruit and flowers present	lvs only, mostly from sterile plants
<i>Arctostaphylos uva-ursi</i>	healthy	as above
<i>Pyrola elliptica</i>	past fruiting	as above
<i>Epigea repens</i>	healthy	sterile plants only
<i>Pleurozium schreberii</i>	healthy	living parts only; litter removed
<i>Cladonia</i> sp.	healthy	as above
<i>Apocynum androsaemifolium</i>	chlorotic; declining	leaflets and petioles

a different stage of development in different stands. This no doubt contributed to the sampling error but it cannot be helped.

It was hoped that all samples could be collected between storms. The collection of samples in the Bearpaw, Wilderness and Woodlot plots, however, was followed by three days of rain. This could have leached nutrients from the foliage, particularly potassium. Also, there could have been an increase in soil nitrification following the storm. There is no way to evaluate this; the effect must be lumped with the variability within types.

Since light is one major variable affecting nutrient uptake, integrative light measurements were taken on each plot using the ozalid paper method of Friend (1961). A more detailed discussion of the technique appears in the appendix.

## 2. Analysis of Soils and Plant Material

The air dry soil samples were screened through a 2 mm sieve and the gravel fraction was discarded. Exchangeable soil and humus cations were extracted with neutral 1 N ammonium acetate according to the method of Black (1965). Calcium, magnesium and potassium were analyzed on a Beckman DU flame spectrophotometer. The interference of calcium at the magnesium wave-length used was corrected for by running calcium standards at the same setting and subtracting the appropriate amount.

Exchangeable hydrogen was analyzed by equilibrating soil and humus samples with 1 N ammonium acetate at a pH of 7.00,

according to the method of Brown (1943). The pH of the humus was measured in the laboratory with a Beckman pH meter.

Available soil phosphorus was extracted with Bray's No. 1 solution (0.03N  $\text{NH}_4\text{F}$  in 0.025N HCL). The concentration was determined spectrophotometrically with the molybdate blue method, according to the procedures used in the University of Minnesota Soil Testing Laboratory (Grava, 1962).

Mechanical analysis was done with the hydrometer method (Black, 1965). The sand fractions from the jack pine stands were separated by screening, but this was later abandoned because of the time required. Correction for the organic fraction in the  $A_1$  horizons were made by burning ten gram samples at  $300^\circ\text{C}$  for two days. It was hoped that this would eliminate the organic matter without a concomitant loss of the water of crystallization.

Nitrogen in both the soil and plant samples was determined by the macro-Kjeldahl method; the soil samples were not ground prior to digestion. Selenium was added as a catalyst.

All other plant nutrients were analyzed on a Jarrell-Ash model 66-000 direct reading emission spectrograph. Material was prepared for analysis by grinding in a wiley mill, dry-ashing at  $525^\circ\text{C}$ , and dissolving in hydrochloric acid containing 0.5 percent lithium.

#### IV. RESULTS AND DISCUSSION

##### 1. Soil Descriptions

Table 3 contains the profile descriptions for each stand; Table 4 contains the mechanical analysis data. These data illustrate the complexity and diversity of the local upland soils.

The Menahga series supporting the two jack pine stands is derived from glacial outwash sands. Since the percentage of silt and clay gives little useful information about this soil, the sand fraction has been subdivided (Table 5). The differing proportions of fine sand in the two Menahga soils appears to be correlated with the jack pine site index.

The A<sub>2</sub> of the Menahga soils is dark, possibly reflecting a residual prairie influence. Horizon development below seven inches is negligible.

The red pine-balsam fir stands are supported by soil derived from stratified sand, silt and gravel. These soils are part of the Marquette series, although extreme local variability makes them difficult to classify. Both stands have a gravelly layer within fourteen inches of the surface, although this is absent from one of the horizons in the Mary Lake stand. These soils are characterized by well developed A<sub>2</sub> horizons and weakly developed sub-horizons.

TABLE 3  
SOIL PROFILE DESCRIPTIONS  
Bearpaw Point, Profile 1

Horizon	Thickness	Color, moist	Structure	Consistence	pH	Miscellaneous
A <sub>11</sub>	4"	10YR 2/1	single grain	soft, friable	6.8	
A <sub>12</sub>	17"	10YR 3/1	single grain	"	6.8	shows signs of degradation
A <sub>13</sub>	3-8"	10YR 3/1	weak crumb	"	6.4	high in organic matter
B <sub>3</sub>	10-15"	10YR 4/3	weak crumb	loose	6.2	bands of organic matter
IIB <sub>3</sub>	4"	10YR 5/3	single grain	"	5.6	very fine sand
IIIC	xxx	xxx	"	"	6.8	stratified fine and coarse sand; lumps of organic matter

Bearpaw Point, Profile 2

A <sub>11</sub>	2"	7.5YR 2/3	weak crumb	soft, friable	6.4	
A <sub>12</sub>	9"	10YR 3/1	" "	"	6.2	
B <sub>3</sub>	6"	10YR 3/2	single grain	loose	6.6	mottling from organic matter

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TABLE 3--Continued

Horizon	Thickness	Color, moist	Structure	Consistence	pH	Miscellaneous
C <sub>1</sub>	15"	10YR 4/4	single grain	loose	6.4	iron bands and mottling from organic matter
Campground, Profile 1						
A <sub>1</sub>	3"	10YR 3/1	crumb	friable	5.8	
A <sub>21</sub>	3"	10YR 4.5/3	platy	firm	5.6	
A <sub>22</sub>	10"	"	"	"	"	
A <sub>2</sub> B <sub>2</sub>	9"	10YR 4/3 rubbed	angular blocky	friable	5.4	A <sub>2</sub> tonguing into B <sub>2</sub> ; possible gleying.
B <sub>2</sub>	11"	5YR 4/8- 10YR 5/4	angular blocky	slightly sticky; plastic	5.4	contains fragments of decomposed rock
C <sub>1</sub>	6"	10YR 4/2	weak ang. blocky	slightly plastic	7.2	much rotten rock; free CaCO <sub>3</sub>
C <sub>2</sub>	x	x	x	x	x	much free lime; rotten rock
Campground, Profile 2						
A <sub>1</sub>	2"	10YR 3/2	crumb	friable	5.0	
A <sub>2</sub>	14"	10YR 5/4	platy	very fr.	5.2	
A <sub>2</sub> B <sub>2</sub>	16"	10YR 4/3- 10YR 5/4	med. sub- ang. blocky;	friable	5.4	smells bad; gleyed upper 4" free of rock fragments



TABLE 3--Continued

Horizon	Thickness	Color, moist	Structure	Consistence	pH	Miscellaneous
B <sub>2</sub>	11"	10YR 4/3 rubbed	med. sub-	slightly sticky, plastic	5.6	many rotten rocks, clear clay flow surfaces
C	x	x	x	x	7.2	free CaCO <sub>3</sub>
Wilderness, Profile 1						
A <sub>12</sub>	2.5"		single grain	loose		
A <sub>2</sub>	3"	10YR 3/3	"	"	4.8	
C <sub>1</sub>	7-9"	10YR 4/3	"	"	5.2	
IIIC	6-8"	10YR 4/3	"	"	5.4	approx. 54 percent gravel; color variable
IIIC	16"	10YR 4/3	"	"	5.8	underlain by stratified sands and gravel
Wilderness, Profile 2						
A <sub>12</sub>	2"		single grain	loose		thin and discontinuous
A <sub>2</sub>	5-11"	10YR 4/2	"	"	4.6	

TABLE 3--Continued

Horizon	Thickness	Color, moist	Structure	Consistence	pH	Miscellaneous
C <sub>1</sub>	7"	10YR 3/4	single grain	loose	5.2	A <sub>2</sub> tonguing into C <sub>1</sub>
IIC	11"	10YR 4/3	"	"	5.2	depth varies from 11 to 20 in.
IIIC	6"	10YR 4/4	"	"	5.7	underlain by stratified sand, gravel and discontinuous till
Mary Lake, Profile 1						
A <sub>1</sub>	1/2"	10YR 3/1	single grain	very friable	6.6	
A <sub>2</sub>	14"	10YR 4/3	"	loose	5.0	
A <sub>2</sub> C	6"	10YR 4/3- 10YR 4/4	"	"	5.6	A <sub>2</sub> tonguing into C, iron stain in C; 37 percent gravel
C	5"	10YR 4/4	"	"	5.6	37 percent gravel
IIC	3"	10YR 5/4	weak sub-angular blocky	very friable	5.6	some clay flow surfaces; iron banding; underlain by stratified sand & gravel

TABLE 3--Continued

Horizon	Thickness	Color, moist	Structure	Consistence	pH	Miscellaneous
Mary Lake, Profile 2						
A <sub>1</sub>	0.5"	10YR 3/2	single grain	loose	5.2	thin and discontinuous
A <sub>21</sub>	5"	10YR 4/3	"	very friable	5.0	
A <sub>22</sub>	5"	10YR 4/3	"	"	5.0	
C <sub>1</sub>	17"	10YR 4/4	"	loose	5.6	variable in depth
IIC	5"	10YR- 2.5Y 4/4	weak sub- ang. blocky	firm	5.8	some clay flow surfaces; underlain by stratified sand and silt with iron bands
Scott's, Profile 1						
A <sub>1</sub>	1"	10YR 3/1	single grain	loose	5.4	thin, discontinuous
A <sub>2</sub>	5"	10YR 3/2	"	"	5.6	
C <sub>1</sub>	6"	10YR 3/3	"	"	5.7	
C <sub>2</sub>	11"	10YR 4/3	"	"	5.7	
C <sub>3</sub>	17"	10YR 4/4	"	"	6.2	

TABLE 3--Continued

Horizon	Thickness	Color, moist	Structure	Consistence	pH	Miscellaneous
Scott's Profile 2						
A <sub>1</sub>	1"	10YR 2/1	single grain	loose	5.0	
A <sub>2</sub>	4"	10YR 2/2	"	"	5.4	
C <sub>1</sub>	11"	10YR 3/3	"	"	5.8	
C <sub>2</sub>	13"	10YR 4/3	"	"	5.8	
C <sub>3</sub>	17"	10YR 4/3	"	"	5.9	
Woodlot, Profile 1						
A <sub>2</sub>	5-7"	10YR 3/2	single grain	loose	5.4	
C <sub>1</sub>	8-10"	10YR 4/3	"	"	5.4	
C <sub>2</sub>	6"	10YR 5/6	"	"	5.6	
C <sub>3</sub>	7"	10YR 5/4	"	"	5.8	
Woodlot, Profile 2						
A <sub>2</sub>	3-7"	10YR 3/4	single grain	loose	5.6	
C <sub>1</sub>	13-17"	10YR 4/4	"	"	5.6	
C <sub>2</sub>	6"	10YR 5/6	"	"	5.6	
C <sub>3</sub>	11"	10YR 5/4	"	"	5.6	

TABLE 4  
 PARTICLE SIZE ANALYSIS OF SOIL SAMPLES

Profile	Horizon	Percent Sand	Percent Silt	Percent Clay
1. Bearpaw Point				
1	A <sub>11</sub>	75.2	21.6	3.2
1	A <sub>12</sub>	81.8	15.2	3.0
1	A <sub>13</sub>	73.4	17.4	9.2
1	B <sub>3</sub>	89.0	8.0	3.0
1	IIIB <sub>3</sub>	62.0	31.0	7.0
1	IIIC	91.0	6.0	3.0
2	A <sub>11</sub>	83.0	13.8	3.2
2	A <sub>12</sub>	85.8	10.1	4.1
2	B <sub>3</sub>	93.0	5.0	2.0
2	C <sub>1</sub>	97.0	1.0	2.0
2	IIC	98.0	1.0	1.0
2. Campground				
1	A <sub>1</sub>	50.8	40.6	8.6
1	A <sub>21</sub>	57.0	36.0	7.0
1	A <sub>22</sub>	62.0	31.0	7.0
1	A <sub>2</sub> B <sub>2</sub>	55.0	28.0	17.0
1	B <sub>2</sub>	51.0	27.0	22.0
1	C <sub>1</sub>	61.0	24.0	15.0
2	A <sub>1</sub>	39.7	48.7	11.6
2	A <sub>2</sub>	45.0	44.0	11.0
2	A <sub>2</sub> B <sub>2</sub>	71.0	16.0	13.0
2	B <sub>2</sub>	57.0	25.0	20.0
2	C	66.0	27.0	7.0

TABLE 4--Continued

Profile	Horizon	Percent Sand	Percent Silt	Percent Clay
3. Wilderness				
1	A <sub>1</sub>	73.3	17.4	4.6
1	A <sub>2</sub>	86.0	10.0	4.0
1	C <sub>1</sub>	86.0	10.0	4.0
1	IIC	90.0	6.0	4.0
2	A <sub>12</sub>	70.0	25.0	5.0
2	A <sub>2</sub>	82.0	14.0	4.0
2	C <sub>1</sub>	86.0	9.0	5.0
2	IIC	95.0	3.0	2.0
4. Mary Lake				
1	A <sub>1</sub>	73.3	23.4	3.3
1	A <sub>2</sub>	83.0	13.0	4.0
1	A <sub>2C</sub>	94.0	4.0	2.0
1	C <sub>1</sub>	96.0	1.0	3.0
1	IIC	82.0	12.0	6.0
1	IIIC	80.0	17.0	3.0
2	A <sub>1</sub>	71.5	23.0	5.5
2	A <sub>21</sub>	84.0	12.0	4.0
2	A <sub>22</sub>	90.0	7.0	3.0
2	C <sub>1</sub>	98.0	1.0	1.0
2	IIC	57.0	32.0	11.0
2	IIIC	97.0	trace	3.0

TABLE 4--Continued

Profile	Horizon	Percent Sand	Percent Silt	Percent Clay
5. Scott's				
1	A <sub>1</sub>	92.8	4.1	3.1
1	A <sub>2</sub>	93.0	5.3	2.2
1	C <sub>1</sub>	95.0	2.8	2.2
1	C <sub>2</sub>	96.0	2.8	1.2
2	A <sub>1</sub>	94.6	1.8	3.6
2	A <sub>2</sub>	94.1	3.5	2.4
2	C <sub>1</sub>	95.6	1.0	3.4
2	C <sub>2</sub>	96.7	0.0	3.3
6. Woodlot				
1	A <sub>2</sub>	85.4	11.2	2.4
1	C <sub>1</sub>	86.6	9.0	4.4
1	C <sub>2</sub>	94.2	4.4	1.4
1	C <sub>3</sub>	98.2	1.8	0.0
2	A <sub>2</sub>	85.5	9.0	4.5
2	C <sub>1</sub>	87.0	6.6	6.4
2	C <sub>2</sub>	100.0	0.0	0.0
2	C <sub>3</sub>	100.0	0.0	0.0

TABLE 5

SAND FRACTIONS FOR MENAUGA SOIL,  
AS PERCENT OF TOTAL SAND

## 1. Woodlot

Profile	Horizon	v. coarse	Coarse	Med.	Fine	v. fine
1	A <sub>2</sub>	8.5	26.8	33.6	27.9	3.4
1	C <sub>1</sub>	12.2	26.6	29.2	28.6	3.1
1	C <sub>2</sub>	15.7	25.6	27.6	38.2	3.0
1	C <sub>3</sub>	7.0	20.0	34.6	35.0	2.2
2	A <sub>2</sub>	9.3	29.9	32.0	26.0	2.8
2	C <sub>1</sub>	11.5	32.0	28.9	25.0	2.7
2	C <sub>2</sub>	10.8	23.2	31.8	30.0	3.4
2	C <sub>3</sub>	27.9	29.0	23.9	18.2	0.9

## 2. Scott's

1	A <sub>11</sub>	1.4	19.9	42.3	34.0	2.3
1	A <sub>12</sub>	1.1	14.2	40.9	40.5	3.2
1	C <sub>1</sub>	1.0	15.0	40.9	40.4	3.2
1	C <sub>2</sub>	0.8	12.9	37.2	45.8	3.1
2	A <sub>11</sub>	1.0	17.7	40.1	38.1	3.1
2	A <sub>12</sub>	1.4	16.3	37.1	40.7	3.7
2	C <sub>1</sub>	0.7	14.4	39.4	42.4	3.0
2	C <sub>2</sub>	0.8	13.6	37.3	44.4	3.8



The soil supporting the Campground maple-basswood stand is in the Beltrami series, belonging to the Gray Wooded great soil group. It is derived largely from calcareous till, but is overlain locally by water-deposited silt. A few large boulders are present in the zone below the silt-cap. The humus is a well developed mull, in contrast with the mor humus of the conifer stands. Both the  $A_2$  and subhorizons are well developed. The soil is poorly drained and shows evidence of gleying in the  $A_2B_2$  horizon and below.

The soil of the Bearpaw point basswood stand may be classified as a Poppelton or ~~Nymore~~ loamy sand. The presence of mesic-rich vegetation on loamy sand presents an interesting anomaly. This is explained, however, by the thickness of the  $A_1$  horizon, which varies from 30 to 12 inches throughout the stand. Probably this plot, which lies within one hundred yards of Lake Itasca, was formerly dominated by lake-shore or bog vegetation. With the recession of the Lake, the organic deposits decomposed and worked downward into the stratified sands. Subsequent encroachment of the forest appears to have furthered the degradation of the organic matter and may eventually produce an  $A_2$  horizon.

The  $A_0$  horizon in this stand is also somewhat unique. Although the humus resembles a mor humus morphologically, the base saturation is almost 100 percent. According to the classification scheme of Duchaufour (1961), this could probably be called a sandy moder.

## 2. Soil Chemical Analysis

Table 6 shows the results of the soil chemical analysis for each horizon. Because only two profiles were examined in each stand, a statistical treatment of the soil nutrient data would be meaningless. Some qualitative conclusions, however, are possible.

a. Nitrogen. There is an increase in percent total nitrogen in the soil profile from the jack pine stand to the hardwood stands, particularly in the surface horizons. An exact ranking is difficult to assign on a percentage basis, however, since the volume of gravel and bulk density are not considered. The highest concentration of total nitrogen was found in the Poppelton soil, due to the high organic matter content. The decrease with depth here was rapid as compared with the Beltrami, due to the sandy parent material.

Pluth and Arneman (1963) found a significant correlation between the plot synecological nutrient coordinate and percent total nitrogen in the O<sub>1</sub>, O<sub>1</sub> and A<sub>1</sub> horizons of soils in the central pine section. Bakuzis (1961) also reported a significant correlation between total soil nitrogen and synecological nutrient coordinate, but found that the moisture coordinate was more highly correlated with percent nitrogen.

b. Phosphorus. The distribution pattern for "available" phosphorus is somewhat anomalous. In most horizons, available phosphorus was low in the surface horizon. It generally increased

TABLE 6  
SOIL CHEMICAL ANALYSIS

Profile	Horizon	Ca <sup>++</sup>	Mg <sup>++</sup>	K <sup>+</sup>	H <sup>+</sup>	Per- cent Base Sat.	Percent N	PPM P
1. Bearpaw Point								
1	A <sub>11</sub>	10.19	1.66	0.35	2.0	86	0.200	65.7
1	A <sub>12</sub>	4.30	0.66	0.21	2.5	68	0.042	67.5
1	A <sub>13</sub>	7.05	0.50	0.31	3.0	72	0.101	131.1
1	IIB <sub>3</sub>	3.50	0.33	0.12	3.0	57	0.024	52.6
1	B <sub>3</sub>	1.50	0.27	0.08	1.5	55	0.019	48.2
2	A <sub>11</sub>	10.90	1.90	0.30	2.0	87	0.247	29.7
2	A <sub>12</sub>	3.87	0.58	0.28	1.5	76	0.056	79.6
2	B <sub>3</sub>	1.44	0.25	0.08	2.0	47	0.019	45.2
2	C	1.75	0.29	0.11	1.5	59	0.009	50.7
2	IIC	2.06	0.16	0.08	1.5	60	0.010	50.7
2. Campground								
1	A <sub>1</sub>	7.57	1.75	0.22	5.0	66	0.167	46.8
1	A <sub>21</sub>	1.62	0.41	0.12	3.0	41	0.029	43.7
1	A <sub>22</sub>	1.20	0.41	0.09	3.0	36	0.014	36.9
1	A <sub>2</sub> B <sub>2</sub>	4.32	1.32	0.14	3.0	66	0.019	15.5
1	B <sub>2</sub>	6.71	3.50	0.22	3.0	78	0.023	10.1
1	C <sub>1</sub>	5.26	3.38	0.15	0.0	100	0.020	10.5
2	A <sub>1</sub>	4.07	1.68	0.51	12.0	34	0.269	50.0
2	A <sub>2</sub>	1.87	0.66	0.13	2.5	52	0.024	26.0
2	A <sub>2</sub> B <sub>2</sub>	4.07	1.57	0.15	2.5	70	0.021	14.0
2	IIA <sub>2</sub> B <sub>2</sub>	4.40	1.66	0.13	2.5	71	0.019	11.5
2	B <sub>2</sub>	5.63	3.80	0.14	3.0	76	0.027	8.2
2	C	4.41	2.56	0.05	0.0	100	0.010	5.0

TABLE 6--Continued

Profile	Horizon	Ca <sup>++</sup>	Mg <sup>++</sup>	K <sup>+</sup>	H <sup>+</sup>	Per- cent Base Sat.	Percent N	PPM P
3. Wilderness								
1	A <sub>12</sub>	3.03	0.92	0.17	5.3	44	0.097	44.3
1	A <sub>2</sub>	0.94	0.33	0.06	2.5	35	0.057	38.4
1	C <sub>1</sub>	1.18	0.41	0.08	1.5	53	0.013	42.8
1	IIC	1.06	0.37	0.06	1.5	50	0.011	30.9
2	A <sub>12</sub>	3.59	1.41	0.26	9.0	37	0.144	37.0
2	A <sub>2</sub>	0.88	0.41	0.06	3.0	31	0.029	35.8
2	C <sub>1</sub>	0.93	0.45	0.08	5.0	23	0.019	50.0
2	IIC	0.74	0.30	0.05	3.0	27	0.008	43.7
4. Mary Lake								
1	A <sub>1</sub>	7.18	1.16	0.30	6.0	59	0.194	14.6
1	A <sub>2</sub>	0.31	0.21	0.13	2.5	21	0.020	57.9
1	AC <sub>1</sub>	0.81	0.33	0.08	1.0	52	0.006	13.9
1	C <sub>1</sub>	1.18	0.24	0.07	2.0	43	<del>xxxx</del>	11.9
1	IIC	2.17	0.27	0.09	3.0	45	0.010	12.0
1	IIIC	2.00	0.41	0.07	2.0	56	0.006	10.9
2	A <sub>1</sub>	11.10	2.67	0.56	4.0	79	0.184	11.1
2	A <sub>21</sub>	0.95	0.31	0.13	4.0	26	0.040	56.4
2	A <sub>22</sub>	0.63	0.21	0.08	3.0	23	<del>xxxx</del>	55.9
2	C <sub>1</sub>	1.06	trace	0.06	1.5	53	0.006	33.3
2	IIC	3.68	0.78	0.16	2.5	65	0.013	27.3
2	IIIC	0.81	trace	0.04	2.0	42	0.0	7.2

TABLE 6--Continued

Profile	Horizon	Ca <sup>++</sup>	Mg <sup>++</sup>	K <sup>+</sup>	H <sup>+</sup>	Per- cent Ease Sat.	Percent N	PPM P
5. Scott's								
1	A <sub>11</sub>	1.18	0.12	0.06	2.2	39	0.054	12.4
1	A <sub>2</sub>	1.99	0.45	0.19	2.0	57	0.027	35.9
1	C <sub>1</sub>	0.75	trace	0.05	1.5	35	0.010	71.5
1	C <sub>2</sub>	0.69	trace	0.03	1.5	32	0.006	62.2
1	C <sub>3</sub>	0.50	trace	0.04	1.0	33	0.001	38.5
2	A <sub>11</sub>	3.06	0.70	0.24	5.5	38	0.109	17.1
2	A <sub>2</sub>	1.18	0.16	0.08	3.3	30	0.031	25.5
2	C <sub>1</sub>	0.81	trace	0.07	5.3	14	0.010	80.0
2	C <sub>2</sub>	0.69	trace	0.06	1.0	43	0.006	63.5
2	C <sub>3</sub>	0.69	0.21	0.06	xxx	xx	0.004	70.9
6. Woodlot								
1	A <sub>2</sub>	2.01	0.58	0.12	2.5	51	0.041	20.2
1	C <sub>1</sub>	1.44	0.58	0.16	1.5	60	0.013	41.2
1	C <sub>2</sub>	0.81	0.16	0.08	1.5	39	0.001	124.9
1	C <sub>3</sub>	0.44	trace	0.04	1.0	33	0.002	127.4
2	A <sub>2</sub>	2.20	0.66	0.11	2.5	55	0.034	23.1
2	C <sub>1</sub>	1.88	0.66	0.09	2.0	57	0.020	38.4
2	C <sub>2</sub>	0.81	0.16	0.04	1.5	40	0.021	105.6
2	C <sub>3</sub>	0.31	trace	0.01	1.0	24	0.0	50.0
2	C <sub>4</sub>	0.37	trace	0.03	xx	xxx	0.001	28.9

to a maximum value in the  $A_2$  or  $C_1$  horizon and then decreased with depth. Thus some of the highest values were from the subhorizons of the Menahga profiles. Only in the Beltrami profiles did phosphorus decrease continuously from the  $A_1$  to the C horizon. There appears to be no relationship between stand synecological coordinate and ppm of "available" soil phosphorus in the stands and profiles sampled.

These somewhat negative results may be due to the variation in the chemical state of phosphorus among horizons and soil types. For example, in the acid subhorizons of the Menahga, most phosphorus is probably bound with iron or aluminum, whereas in the Beltrami subhorizons, it is probably present as calcium phosphates. In the surface horizons and humus layer, however, it is probably tied up in organic compounds. Thus it is unreasonable to expect one extraction method to approximate the available phosphorus for a wide range of soil types, horizons and plant species.

The lack of a direct relationship between soil phosphorus and forest soil fertility or productivity has long been known. Valmari (1921), in fact found an inverse relationship between forest productivity and kilograms per hectare of  $P_2O_5$  extracted with 0.2 N HCl. Viro (1952) measured total clay phosphorus as well as extracting with Ca lactate, Na zeolite, acid  $NH_4F$  and neutral  $NH_4F$ . When a broad range of forest stands were considered, none of these methods gave values which were correlated with fertility of the forest soil. When all measurements above a certain limit were discarded, however, then the lactate-soluble, flouride-soluble and total clay phosphorus values were well correlated

with forest soil fertility. Thus he concluded that the fertility of forest soil does depend on the phosphate content.

The lack of a direct relationship between soil phosphorus and synecological coordinates has been reported for Minnesota. Pluth and Arneman (1963) using the dilute acid flouride extracting method, found no significant correlation between plot synecological nutrient coordinate and available phosphorus in the humus, A<sub>1</sub>, A<sub>2</sub> or C horizons. They did, however, report a negative correlation with available phosphorus in the B<sub>2</sub> horizon. Pakuzis (1961) also found no clear distribution of available phosphorus in moisture-nutrient coordinates. Thus it is possible that phosphorus is not a limiting factor in forest productivity in the Central Pine Section. Further studies on the phosphorus problem should involve different extraction methods and fertilizer experiments.

c. Exchangeable Bases. Given the broad range of soil types selected for this study, it is not surprising that there appears to be a good relationship between exchangeable bases and stand synecological nutrient coordinate. The concentration of calcium, potassium and magnesium within a given profile, however, appears to be closely related to the percentage of silt plus clay. Between profiles, the degree of profile development is an important controlling factor. For example, the A<sub>2</sub> horizons of the Marquette profiles are much lower in calcium than the A<sub>2</sub> horizons of the Menahga profiles, although the surface layer of the Marquette is higher in calcium.

In the subhorizons of the Campground maple-basswood stand and at least two of the profiles under the red pine stands, the concentration of exchangeable bases increases with depth. This is due in part to soil development and in part to parent material. Thus in the Beltrami, the calcium profile is the result of leaching of the  $A_2$ , accumulation in the  $B_2$  horizon and presence of free calcium in the parent material. In the Marquette, particularly at Mary Lake, the increase of calcium with depth reflects the presence of layers of silt and very fine sand with a higher exchange capacity than the coarse sand above.

### 3. Vegetation Analysis

Table 7 shows the total cubic foot volume by species on each 128 square meter subplot. These data show the relative dominance of different species within each stand. They also illustrate the irrelevance of total volume or biomass as a measure of productivity.

Table 8 shows the basal area and the site index by species within each stand. Red pine site index for the Woodlot stand was measured on trees outside of the plot. The site index reported for red pine in the Campground stand is unreliable, since the three trees measured were probably suppressed during part of their development. For the other stands, however, the site index is based on adequately large samples.

The plot synecological coordinates are given in table 9. These values are based on species presence in the eight square



TABLE 7

TOTAL CUBIC FOOT VOLUME ON 128 m<sup>2</sup> SUBPLOTS

Species	Subplot				Number			
	1	2	3	4	5	6	7	8
1. Campground								
<i>Acer saccharum</i>	3.0	12.4	9.5	2.1	30.4	1.5	3.8	2.2
<i>Quercus rubra</i>	40.5	23.8	x	23.8	54.0	42.7	x	x
<i>Populus tremuloides</i>	51.4	x	x	80.0	x	x	45.2	4.4
<i>Tilia americana</i>	4.4	x	25.0	20.5	42.6	8.4	4.4	x
<i>Betula papyrifera</i>	7.3					27.0	14.4	11.2
<i>Pinus strobus</i>	284.					160.		
<i>Pinus resinosa</i>								169.
<i>Acer rubrum</i>			9.2			3.2		
<i>Quercus macrocarpa</i>							7.5	
2. Bearpaw Point								
<i>Tilia americana</i>	135.2	138.6	47.5	46.0	147.7	29.1	127.2	69.9
<i>Betula papyrifera</i>	9.7	22.0			16.3			
<i>Ulmus americana</i>			12.1	13.3	3.0	32.5		18.1

TABLE 7--Continued

Species	Subplot Numbers							
	1	2	3	4	5	6	7	8
3. Wilderness								
Pinus resinosa	333	371	438	297	328	269	258	286
Abies balsamea	2.8	5.3	3.8	10.9	1.8	15.8	2.9	10.5
Betula papyrifera	3.2	5.8		1.6	4.5		1.6	
4. Mary Lake								
Pinus resinosa	67.0		78.0		124.5	106.5	157.5	
Abies balsamea	22.7	28.1	78.9	10.4	12.3	15.8	7.8	32.6
Betula papyrifera	19.2			46.2	18.8	7.0	7.4	4.5
5. Scott's								
Pinus banksiana	97.4	54.0	110.3	40.0	85.8	125.3	83.3	111.2
Betula papyrifera				16.6				
6. Woodlot								
Pinus banksiana	64.9	76.4	72.6	74.8	89.1	60.9	65.8	60.8

TABLE 8  
 BASAL AREA AND SITE INDEX

Stand	Species	Basal Area, sq. ft./acre	Site Index
Woodlot	jack pine	109	49
Scott's	red pine		35*
Scott's	jack pine	113	68
Mary Lake	red pine	48	45
Mary Lake	balsam fir	22	
Mary Lake	paper birch	35	
Mary Lake	white pine	2	
Wilderness	red pine	155	53
Wilderness	balsam fir	15	
Wilderness	white pine	2	
Bearpaw Point	basswood	91	
	American elm	17	
	paper birch	7	
	bur oak	4	
Campground	sugar maple	30	
	quaking aspen	10	
	bigtooth aspen	4	
	red oak	19	
	white pine	28	
	red pine	6	51
	paper birch	10	
	hop hornbeam	2	
	basswood	6	

\* Measured outside of the plot.

TABLE 9  
SYNECOLOGICAL COORDINATES, PRESENCE BASIS

Subplot	Moisture	Nutrients	Heat	Light
1. Bearpaw Point				
1	2.4	3.4	3.2	2.5
2	2.2	3.1	3.1	2.8
3	2.2	2.9	2.9	2.9
4	2.5	3.5	3.2	2.4
5	2.6	3.3	2.8	2.3
6	2.2	3.0	2.9	2.4
7	2.3	3.1	2.9	2.9
8	2.4	3.4	3.1	2.5
2. Campground				
1	2.2	3.1	2.6	2.2
2	2.3	3.0	2.6	2.2
3	2.3	3.0	2.7	2.5
4	2.1	3.5	2.8	2.1
5	2.0	3.2	3.1	2.4
6	2.1	3.4	3.9	2.3
7	2.6	2.8	2.5	2.5
8	2.2	3.2	2.8	2.2

TABLE 9--Continued

Subplot	Moisture	Nutrients	Heat	Light
3. Wilderness				
1	2.5	2.3	2.2	2.9
2	2.4	2.3	2.1	2.5
3	2.5	2.3	2.2	3.2
4	2.9	2.1	1.9	2.8
5	2.2	2.2	1.8	2.8
6	2.6	2.1	1.9	2.4
7	2.6	2.1	2.0	2.7
8	2.5	2.3	2.0	2.5
4. Mary Lake				
1	2.4	2.1	1.9	2.6
2	2.3	2.1	2.0	2.8
3	2.0	2.0	2.1	3.1
4	2.1	2.3	2.1	3.5
5	2.3	2.2	2.0	3.3
6	2.5	2.0	2.0	3.1
7	1.9	2.0	2.1	4.0
8	2.0	2.1	1.9	3.6

TABLE 9--Continued

Subplot	Moisture	Nutrients	Heat	Light
5. Scott's				
1	1.6	2.0	2.5	4.1
2	1.5	1.9	2.2	4.1
3	1.7	1.9	2.2	4.0
4	1.6	2.0	2.2	3.8
5	1.6	1.8	2.1	4.3
6	1.7	1.8	2.2	4.1
7	1.6	1.8	2.2	4.2
8	1.4	1.8	2.2	4.4
6. Woodlot				
1	1.3	1.6	2.3	4.2
2	1.7	1.8	2.2	4.3
3	1.5	1.7	2.2	4.3
4	1.6	1.9	2.3	4.4
5	1.6	1.9	2.2	4.2
6	1.5	1.7	2.2	4.4
7	1.5	1.9	2.4	4.3
8	1.6	1.8	2.3	4.3

meter plots. Coordinate values for nutrients and light were also computed on a dominance basis for the herb layer. These data are presented in Table 10.

Figure 3 shows the position of each plot in the edaphic field with moisture and nutrients as coordinate axes. The two jack pine stands fall somewhat below the moisture-nutrient coordinates for the jack pine stands sampled by Rakuzis (1961). The red pine stands are roughly centered in the edaphic field with respect to moisture, but the nutrient coordinates are somewhat low. The hardwood stands have roughly the same moisture coordinates as the red pine stands, but the nutrient coordinates are in the upper portion of the edaphic field for Minnesota.

#### 4. Light Measurements

The integrative light measurements showed a high correlation between percent of full sunlight and synecological light coordinates. The results are plotted in Figure 4. On a presence basis the correlation coefficient is +0.86, but on a dominance basis it is only +0.66. A more detailed discussion of the results of the light measurements appears in the appendix.

#### 5. Analysis of Humus Samples

The humus samples collected at each subplot were leached with ammonium acetate and analysed for exchangeable calcium, magnesium, potassium and hydrogen. The results show that the relationship between these four cations varies from one end of the nutrient

TABLE 10  
SYNECOLOGICAL COORDINATES, DOMINANCE BASIS

Subplot	Nutrients	Light
1. Bearpaw Point		
1	3.4	2.5
2	3.2	2.6
3	4.0	2.1
4	4.1	2.1
5	3.9	1.5
6	3.0	2.4
7	3.7	2.0
8	2.9	2.5
2. Campground		
1	3.6	2.0
2	3.3	2.1
3	4.4	1.5
4	3.5	2.4
5	2.6	3.2
6	4.8	1.2
7	4.3	1.5
8	4.3	1.5



TABLE 10--Continued

Subplot	Nutrients	Light
3. Wilderness		
1	2.0	3.7
2	2.0	3.1
3	1.9	3.5
4	2.0	3.3
5	1.9	3.4
6	2.0	3.0
7	1.8	2.5
8	2.0	3.0
4. Mary Lake		
1	2.0	3.0
2	2.2	2.0
3	2.2	3.3
4	2.0	3.2
5	2.0	3.9
6	1.7	3.7
7	2.0	3.4
8	2.0	3.2

TABLE 10--Continued

<u>Subplot</u>	<u>Nutrients</u>	<u>Light</u>
5. Scott's		
1	1.9	3.6
2	1.7	4.4
3	1.7	3.4
4	2.0	3.3
5	1.7	4.3
6	1.6	4.0
7	1.2	4.8
8	1.3	4.8
6. Woodlot		
1	1.5	3.7
2	1.7	3.3
3	1.7	3.5
4	1.9	4.4
5	1.7	4.7
6	2.0	4.1
7	1.8	3.8
8	1.8	3.8

# PERCENT BASE SATURATION OF HUMUS IN MOISTURE-NUTRIENT COORDINATES

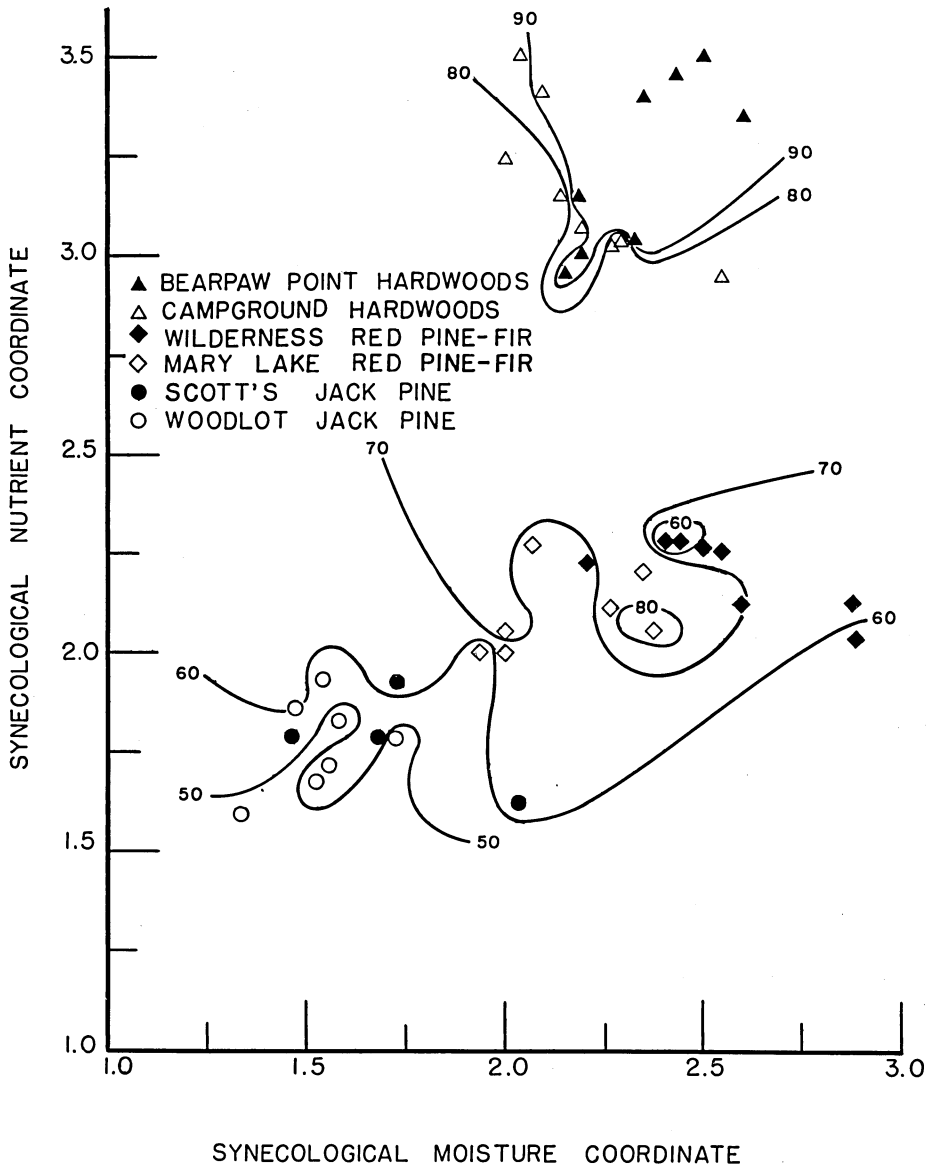


Figure 3. The distribution of subplots in moisture-nutrient coordinate axes and horizontal lines showing the percent base saturation of the humus collected at each subplot. Well developed humus was absent from some subplots; the points were omitted from the diagram.

# RELATIONSHIP BETWEEN LIGHT INTENSITY AND SYNECOLOGICAL LIGHT COORDINATES IN AN UPLAND FOREST GRADIENT

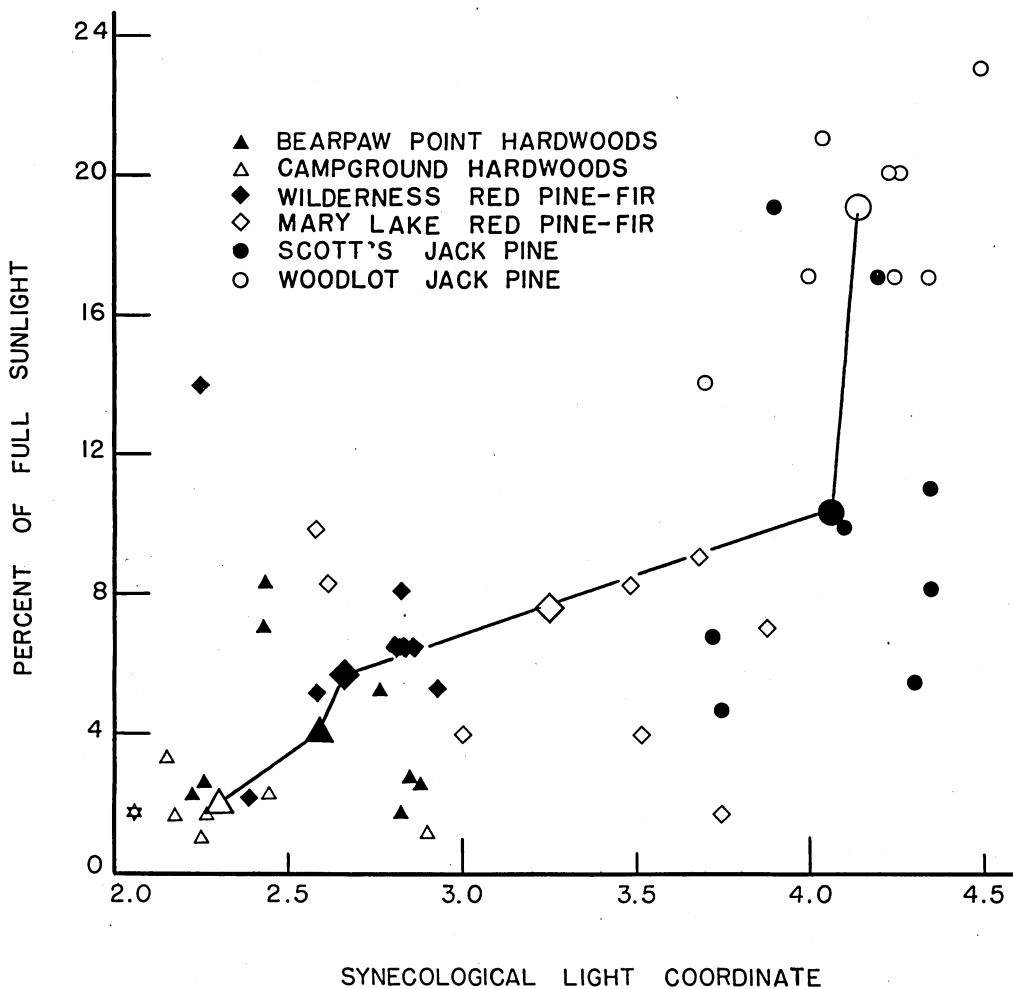


Figure 4. The relationship between integrated light intensity for one day and synecological light coordinates in six forest stands. Light intensity was measured with ozalid paper. The large symbols connected by the solid line represent stand averages.

gradient to the other.

On the assumption that the exchange capacity can be approximated by the sum of the exchangeable cations, the percent base saturation of each sample was calculated (see Table 11). This value shows a high degree of correlation with plot synecological nutrient coordinate; the linear correlation coefficient obtained was +0.83 on a presence basis and +0.73 on a dominance basis. Both values are highly significant.

Figure 3 shows the percent base saturation as contours in the moisture-nutrient coordinate field. The greatest variation occurs in the red pine-balsam fir stands. This may be due to the heterogeneous ground flora or the influence of scattered birch trees. The Campground maple-basswood stand also shows considerable variation; this appears to be related to the presence of isolated red pine and white pine trees in the stand. In contrast, the jack pine stands show relatively little variation. The humus layer in the Bearpaw Point basswood stand is the least variable and is practically saturated with bases, chiefly calcium.

In order to explore the relationships between these exchangeable cations along the nutrient gradient, triangular coordinates were used. This involves grouping the cations three at a time and mathematically adjusting the values so that the sum of the three cations (e.g. calcium, magnesium and potassium) is equal to 100 percent (see Table 12 for sample computations). In this computation, the average of all points is exactly at the center of the triangle. Thus each sample is expressed in relation to the average of a given cation for all samples, and each of the three

TABLE 11

EXCHANGEABLE CATIONS IN HUMUS, MEQ. PER 100 GRAMS

Subplot	H <sup>+</sup>	Ca <sup>++</sup>	K <sup>+</sup>	Mg <sup>++</sup>	Percent Base Sat.
1. Bearpaw Point					
1	1.1	51.4	1.57	14.4	98.4
2	0.6	63.3	1.77	10.6	99.2
3	0.2	51.6	1.43	9.8	99.7
4	1.1	51.6	1.58	12.1	98.3
5	3.0	52.0	1.75	10.9	95.6
6	2.0	61.5	1.12	11.0	97.4
7	0.9	77.9	1.93	13.6	99.0
8	4.4	51.5	2.19	14.2	93.9
2. Campground					
1	2.6	8.7	0.25	2.3	81.3
2	11.8	20.3	0.98	7.7	71.1
3	6.4	13.9	0.46	4.3	74.5
4	2.6	12.4	0.38	2.6	85.6
5	5.4	11.3	0.55	3.5	74.0
6	1.0	11.8	0.37	3.5	94.0
7	7.8	14.9	0.64	5.2	72.6
8	9.8	16.4	1.04	7.1	71.4

TABLE 11--Continued

Subplot	H <sup>+</sup>	Ca <sup>++</sup>	K <sup>+</sup>	Mg <sup>++</sup>	Percent Base Sat.
3. Wilderness					
1	15.0	24.5	0.88	6.2	67.8
2	18.2	15.1	1.14	5.1	53.9
3	20.5	19.8	1.42	6.0	57.0
4	14.8	22.2	1.12	4.9	65.6
5	11.8	20.2	0.81	3.6	67.6
6	14.8	26.4	1.02	7.1	70.0
7	16.0	17.8	1.09	4.4	59.3
8	17.7	20.5	0.83	6.1	60.8
4. Mary Lake					
1	6.0	36.9	1.37	12.6	89.5
2	12.1	36.8	1.04	7.4	78.9
3	15.5	20.7	0.94	4.4	64.1
4	12.1	20.0	1.02	6.1	69.1
5	11.8	26.6	1.20	6.2	74.2
6	<del>XXX</del>	<del>XXX</del>	<del>XXX</del>	<del>XXX</del>	<del>XXX</del>
7	20.5	17.0	1.63	5.1	53.6
8	10.2	25.7	1.02	7.6	77.1

TABLE 11.--Continued

Subplot	H <sup>+</sup>	Ca <sup>++</sup>	K <sup>+</sup>	Mg <sup>++</sup>	Percent Base Sat.
5. Scott's					
3	10.6	11.9	0.61	4.3	61.3
4	14.0	16.9	0.66	4.1	60.8
6	12.1	10.4	0.51	3.8	54.9
8	11.5	11.1	0.54	3.1	56.1
6. Woodlot					
1	8.5	5.1	0.28	1.9	46.2
2	11.0	7.4	0.75	2.5	49.3
3	11.0	10.6	0.45	2.7	55.8
4	9.8	8.06	0.46	2.3	52.4
5	9.2	8.7	0.46	3.0	56.2
7	8.6	9.2	0.66	3.0	60.0
8	8.2	5.5	0.43	2.0	49.1



TABLE 12

SAMPLE CALCULATION FOR CENTERING NUTRIENT  
CONCENTRATIONS IN TRIANGULAR COORDINATES

1. The following values were assumed as original data:

sample number	percent N	percent P	percent K
1	2.28	0.314	1.68
2	1.83	0.237	1.88
3	0.67	0.063	0.27
Column totals:	4.78	0.614	3.83

2. Average values were computed for each of the nutrients.

$$\bar{N} = \frac{4.78}{3} = 1.59$$

$$\bar{P} = \frac{.614}{3} = 0.205$$

$$\bar{K} = \frac{3.83}{3} = 1.28$$

3. The original values were divided by the respective nutrient averages, giving the following values:

sample number	N	P	K	row total
1	1.43	1.53	1.31	4.27
2	1.15	1.16	1.47	3.78
3	0.42	0.31	0.21	0.94

4. Each value was then divided by the total for the row in which it occurred, giving the final adjusted values:

sample number	N	P	K
1	0.335	0.358	0.307
2	0.304	0.307	0.389
3	0.446	0.330	0.223

cations taken is given equal weight. For example, one plot had a K:Mg:Ca ratio of 1:5:15, but the adjusted values for that plot are 45.0 percent for potassium, 31.5 percent for magnesium and 23.5 percent for calcium. The data were adjusted for plotting by an IBM 360 computer.

Table 13 shows the average exchangeable cations for the 42 samples analyzed. Given this population, any sample containing these proportions of cations would fall at the center of the triangular coordinates. For example, a humus sample containing 8.1 meq. of calcium per 100 gm., 2.05 of magnesium and 0.32 of potassium would fall in the center of the Ca-Mg-K coordinates.

The four possible combinations of calcium, magnesium, potassium and hydrogen in forest humus have been plotted in Figure 5. In Figure 5-1, calcium, magnesium and potassium are plotted. The different configurations for the jack pine, red pine-fir and hardwood stands suggests that the ratios of calcium to potassium and magnesium to potassium are higher for the hardwood stands than for the pine stands.

This trend is illustrated in greater detail by the other three combinations. Given the high correlation between synecological nutrient coordinates and percent base saturation, it is natural that the points are well spread out along the  $H^+$  axis in these three diagrams. In the H-Ca-Mg coordinates (Figure 5-2) there is only a slight and inconclusive slant toward calcium with respect to the H-axis. In the H-K-Mg coordinates (Figure 5-3), however, the distribution is slanted toward the potassium side of the H-axis at the low end of the nutrient gradient and toward

TABLE 13

AVERAGE VALUES FOR NUTRIENTS PLOTTED IN  
TRIANGULAR COORDINATES

## 1. Humus Samples

<u>Element</u>	<u>Average for 42 plots, mg. per 100 gm.</u>
calcium	24.40
magnesium	6.14
potassium	0.961
hydrogen	9.33

## 2. Foliar Samples

<u>Nutrient</u>	<u>Average for 94 samples, percent ovendry weight</u>
nitrogen	2.01
phosphorus	0.312
potassium	2.45
calcium	1.23
magnesium	0.340

## EXCHANGEABLE HUMUS CATIONS IN TRIANGULAR COORDINATES

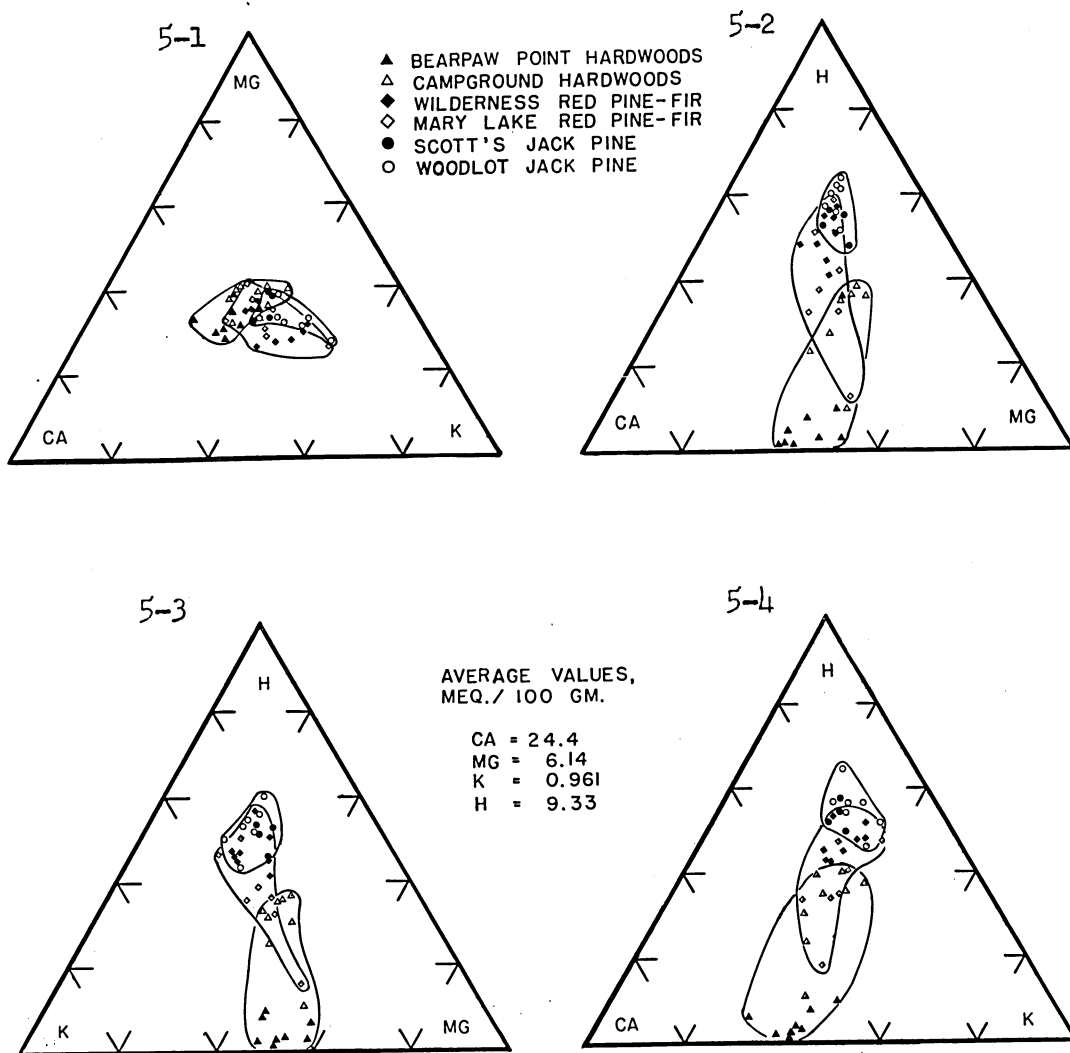


Figure 5. The relationship between exchangeable humus cations from six forest stands. The data were transformed prior to plotting so that the average of all points in each set of coordinates falls at the center of the graph. Lines have been drawn around the points from each forest type. The configurations indicate that the K: Ca plus Mg ratio decreases toward the upper end of the nutrient gradient.

the magnesium side at the upper end. In the H-K-Ca coordinates (Figure 5-4) the pattern is clearest. The Bearpaw Point plots fall almost entirely on the calcium side of the H-axis and the Woodlot plots are entirely on the potassium side.

This relationship is further illustrated in Figure 6, which shows the distribution of synecological nutrient coordinates in K-Ca-Mg coordinates. The high coordinate values are associated with low relative potassium values. Thus it appears that the proportions of calcium and magnesium in the humus exchange complex increase relative to potassium from the lower end of the gradient, dominated by jack pine, to the upper end of the gradient, dominated by northern hardwoods.

In order to evaluate the relationship between the relative and absolute values for exchangeable cations, simple correlations were run between the absolute values, relative values and synecological nutrient coordinates (presence basis). Tables 14 and 15 show the simple correlation coefficients obtained. A comparison of these data further supports the previous conclusions. For example, the correlation coefficient between nutrient coordinates and exchangeable hydrogen is  $-0.69$ , whereas for calcium, magnesium and potassium it is respectively  $+0.54$ ,  $+0.55$  and  $+0.32$  (Table 14). This implies that the sum of the exchangeable bases is more important than any one alone, but calcium and magnesium are both more closely related to the defined gradient than is potassium.

This relationship is shown more strongly by a comparison of the correlation coefficients between nutrient coordinates and

ABSOLUTE AMOUNT OF ELEMENTS, BASE SATURATION PERCENT, AND SYNECOLOGICAL COORDINATES IN CA-MG-K AND N-P-K COORDINATES OF AN UPLAND FOREST GRADIENT IN CENTRAL MINNESOTA PINE SECTION

MILLIEQUIVALENTS OF ELEMENTS PER 100 G OF SOIL HUMUS AND SYNECOLOGICAL COORDINATES

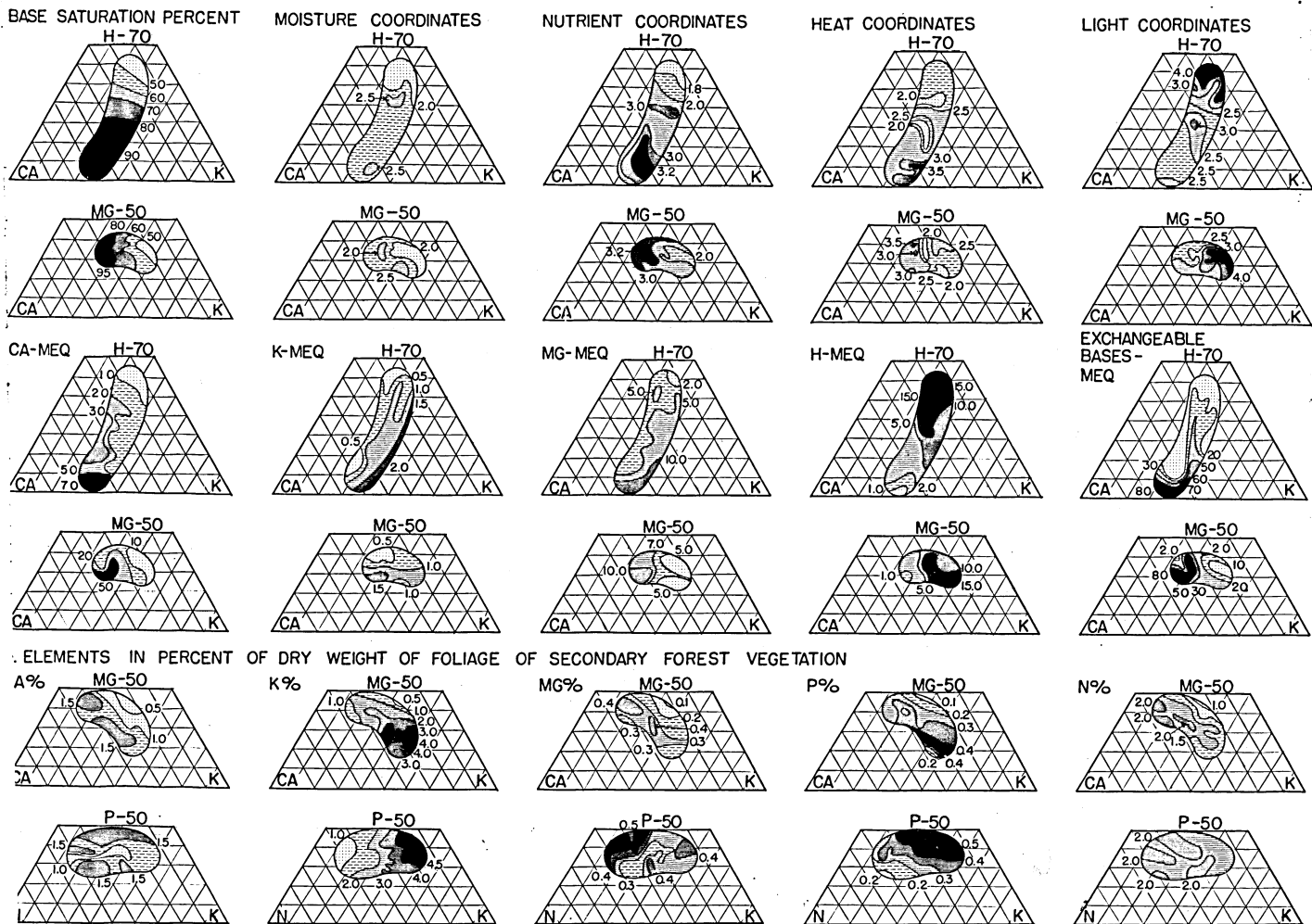


Figure 6. The relationship between relative and absolute humus cation concentrations in Ca-Mg-K and Ca-H-K coordinates and the relationship between humus cations and synecological coordinates. Foliar nutrient concentrations are shown in Ca-Mg-K and N-P-K coordinates.

TABLE 14

## ECOSYSTEM CORRELATION COEFFICIENTS

	Ca	K	Mg	H	pH	Base Sat. Percent	Synecological Coordinates			
							Moisture	Nutrient	Heat	Light
Ca	.7029**	.9074**	.5040**	.8291**	.8113**	.4785**	.5362**	.4449**	-.4092**	
K		.7758**	-.0912	.4380**	.5076**	.5642**	.3281*	.1942	-.3421*	
Mg			-.4043**	.7450**	.7689**	.5488**	.5511**	.4313**	-.4878**	
H				-.8207**	-.7695**	.0081	-.6888**	-.7292**	.3272*	
pH					.9447**	.3181*	.7519**	.6859**	-.5272**	
Base saturation percent						.4963**	.8260**	.6659**	-.6884**	
Moisture coordinate							.4945**	.1548	-.8174**	
Nutrient coordinate								.7333**	-.7853**	
Heat coordinate									-.4092**	
Light coordinate										

\* Significant at the 5 percent level

\*\* Significant at the 1 percent level

TABLE 15

RELATIONSHIP BETWEEN RELATIVE AND ABSOLUTE VALUES  
IN CA-K-MG AND CA-K-H COORDINATES

Relative Values, Ca-K-Mg Coordinates

<u>Absolute Value</u>	<u>Ca</u>	<u>K</u>	<u>Mg</u>
Ca	+0.719**	-0.517**	-0.170
K	+0.245	-0.075	-0.223
Mg		+0.084	+0.551*
Nutrient Coordinate	+0.503**	-0.584**	+0.237

Relative Values, Ca-K-H Coordinates

<u>Absolute Value</u>	<u>Ca</u>	<u>K</u>	<u>H</u>
Ca	+0.845**	+0.608**	-0.847**
K	+0.454**	+0.672**	-0.570**
H	-0.787**	-0.507**	+0.769**
Nutrient Coordinate	+0.763**	+0.685**	-0.810**

\* Significant at the 5 percent level

\*\* Significant at the 1 percent level



relative cation values (Table 15). In the Ca-K-Mg coordinates, for example, the correlation coefficient with synecological nutrient coordinates is +0.50 for calcium, +0.24 for magnesium and -0.58 for potassium.

The correlation coefficient between the relative and absolute value for a given element indicates the degree to which the absolute value is changed by the transformation. With a high positive correlation between three variables, there must be a negative correlation between the relative and absolute value for at least one element. Intuitively, it appears that the element with the highest variance in concentration will be best correlated with the corresponding adjusted value, whereas the least variable element will be the least correlated. In the Ca-K-Mg coordinates, the correlation coefficients between relative and absolute values decrease in the order  $\text{Ca} > \text{Mg} > \text{K}$ ; the variances of these cations decrease in the same order. If at least one element is inversely related to the other two, however, (as with the Ca-K-H coordinates) then there may be a high correlation between the relative and absolute value for all three elements.

Figure 6 shows the absolute values of calcium, potassium, magnesium and hydrogen in Ca-K-Mg and Ca-K-H coordinates. These illustrations show graphically that the effect of the transformation on a single variable is determined by the variance of each element and the degree of correlation between them. If this triangular transformation is used in the future, then the effect of unequal variances and correlation between elements should be solved rigorously.

Duchaufour (1961) suggested that the ecological group on a given site is closely related to the humus type, and that ecological groups should be defined together with humus types. Because of the key position occupied by the humus layer in the nutrient cycle of a forest, a detailed study of the chemical and biological properties of the humus should be part of any future study on the characteristics of an ecological nutrient gradient.

## 6. Foliar Analysis

The elements in the foliage samples that were analyzed by the emission spectrograph are phosphorus, potassium, calcium, strontium, sodium, iron, magnesium, zinc, copper, molybdenum, manganese and boron. The levels of sodium and molybdenum, however, were too low to be measured accurately. Also, the iron values may represent contamination from the Wiley mill or muffle furnace liner. The original data are reported in Tables 16 and 17. The stand numbers in Tables 16 and 17 refer to the stands in the following order: 1) Woodlot jack pine, 2) Scott's jack pine, 3) Mary Lake red pine-balsam fir, 4) Wilderness red pine-balsam fir, 5) Campground hardwoods and 6) Bearpaw Point hardwoods.

A comparison of these data with previously reported concentrations shows a general consistency within species. For example, Gerloff (1964) reported very high potassium concentrations for Clintonia borealis and Aster macrophyllus and low potassium concentrations for Corylus cornuta and Qutrya virginiana (see Table 18). These findings are consistent with the data

TABLE 16-1

## PERCENT FOLIAR CONCENTRATION OF NITROGEN, DRY WEIGHT BASIS

Code No.	Species	Stand Number					
		1	2	3	4	5	6
01	<i>Lithospermum</i> <i>canescens</i>	1.61					
02	<i>Fragaria</i> <i>virginiana</i>	2.23	2.21	2.37			
03	<i>Pteridium</i> <i>aquilinum</i>	1.83	1.92	1.93			
04	<i>Cladonia</i> sp.	0.67	0.74				
05	<i>Epigaea repens</i>	1.47	1.49				
06	<i>Maianthemum</i> <i>canadense</i>	1.73	1.69	1.86	2.01	1.97	1.93
07	<i>Gaultheria</i> <i>procumbens</i>	1.27	1.31				
08	<i>Aster</i> <i>macrophyllus</i>	1.66	1.91	1.83	2.23	1.87	1.93
09	<i>Aralia</i> <i>nudicaulis</i>	1.17	1.88	2.07	2.43	1.69	1.88
10	<i>Arctostaphylos</i> <i>Uva-ursi</i>	1.29	1.41				
11	<i>Pyrola elliptica</i>	1.60	1.66				
12	<i>Pleurozium</i> <i>schreberii</i>	1.17	1.17				
13	<i>Apocynum</i> <i>androsaemifolium</i>	2.31					
14	<i>Corylus cornuta</i>		2.33	2.30	2.56	2.14	2.28
15	<i>Hepatica americana</i>		1.81	1.84	1.94	1.86	
16	<i>Diervilla lonicera</i>		2.03				
17	<i>Thalictrum dioicum</i>			2.46		2.17	2.53
18	<i>Cornus canadensis</i>			1.73	1.69		
19	<i>Clintonia borealis</i>			2.04	2.16	1.91	
20	<i>Parthenocissus inserta</i>				3.51		2.87
21	<i>Lonicera canadensis</i>				1.88		
22	<i>Uvularia grandiflora</i>					2.07	2.38
23	<i>Dirca palustris</i>					2.38	2.38
24	<i>Acer saccharum</i>					1.86	
25	<i>Ostrya virginiana</i>					2.21	
26	<i>Amphicarpa bracteata</i>						4.27
27	<i>Caulophyllum thalictroides</i>						2.41
28	<i>Celastrus scandens</i>						3.21
29	<i>Actaea rubra</i>						2.70
30	<i>Smilacina racemosa</i>						2.24
31	<i>Sanguinaria canadensis</i>						1.31
32	<i>Asarum canadense</i>						1.03

TABLE 16-2

## PERCENT FOLIAR CONCENTRATION OF PHOSPHORUS, DRY WT. BASIS

Code No.	Species	Stand Number					
		1	2	3	4	5	6
01	<i>Lithospermum</i> <i>canescens</i>	0.167					
02	<i>Fragaria</i> <i>virginiana</i>	0.314	0.316	0.376			
03	<i>Pteridium</i> <i>aquilinum</i>	0.237	0.303	0.248			
04	<i>Cladonia</i> sp.	0.063	0.080				
05	<i>Epigaea repens</i>	0.132	0.166				
06	<i>Maianthemum</i> <i>canadense</i>	0.264	0.314	0.332	0.309	0.296	0.410
07	<i>Gaultheria</i> <i>procumbens</i>	0.133	0.137				
08	<i>Aster</i> <i>macrophyllus</i>	0.300	0.312	0.347	0.267	0.329	0.393
09	<i>Aralia</i> <i>nudicaulis</i>	0.269	0.319	0.454	0.380	0.386	0.369
10	<i>Arctostaphylos</i> <i>Uva-ursi</i>	0.155	0.174				
11	<i>Pyrola elliptica</i>	0.192	0.173				
12	<i>Pleurozium</i> <i>schreberii</i>	0.162	0.209				
13	<i>Apocynum</i> <i>androsaemifolium</i>	0.304					
14	<i>Corylus cornuta</i>	0.300	0.384	0.289	0.282	0.309	
15	<i>Hepatica americana</i>	0.241	0.255	0.204	0.239		
16	<i>Diervilla lonicera</i>	0.454					
17	<i>Thalictrum dioicum</i>		0.280		0.257	0.327	
18	<i>Cornus canadensis</i>		0.367	0.234			
19	<i>Clintonia borealis</i>		0.445	0.393	0.500		
20	<i>Parthenocissus inserta</i>			0.318		0.369	
21	<i>Lonicera canadensis</i>			0.235			
22	<i>Uvularia grandiflora</i>				0.291	0.338	
23	<i>Dirca palustris</i>				0.438	0.517	
24	<i>Acer saccharum</i>				0.280		
25	<i>Ostrya virginiana</i>				0.241		
26	<i>Amphicarpa bracteata</i>					0.278	
27	<i>Caulophyllum thalictroides</i>					0.517	
28	<i>Celastrus scandens</i>					0.632	
29	<i>Actaea rubra</i>					0.582	
30	<i>Smilacina racemosa</i>					0.445	
31	<i>Sanguinaria canadensis</i>					0.526	
32	<i>Asarum canadense</i>					0.530	

TABLE 16-3

## PERCENT FOLIAR CONCENTRATION OF POTASSIUM, DRY WT. BASIS

Code No.	Species	Stand Number					
		1	2	3	4	5	6
01	<i>Lithospermum canescens</i>	3.01					
02	<i>Fragaria virginiana</i>	1.68	1.59	1.96			
03	<i>Pteridium aquilinum</i>	1.88	2.27	2.37			
04	<i>Gladonia</i> sp.	0.27	0.31				
05	<i>Epigaea repens</i>	0.66	1.03				
06	<i>Maianthemum canadense</i>	3.27	2.70	3.36	3.19	2.79	1.94
07	<i>Gaultheria procumbens</i>	0.79	0.73				
08	<i>Aster macrophyllus</i>	3.61	3.94	4.59	3.54	4.37	3.96
09	<i>Aralia nudicaulis</i>	1.38	1.76	2.36	2.08	2.79	2.94
10	<i>Arctostaphylos Uva-ursi</i>	0.77	1.01				
11	<i>Pyrola elliptica</i>	1.08	0.86				
12	<i>Pleurozium schreberii</i>	0.87	1.18				
13	<i>Apocynum androsaemifolium</i>	1.85					
14	<i>Corylus cornuta</i>		0.84	1.30	1.14	1.11	1.04
15	<i>Hepatica americana</i>		2.90	3.03	2.31	2.76	
16	<i>Diervilla lonicera</i>		2.86				
17	<i>Thalictrum dioicum</i>			2.97		3.21	3.56
18	<i>Cornus canadensis</i>			1.78	1.33		
19	<i>Clintonia borealis</i>			4.53	4.55	4.83	
20	<i>Parthenocissus inserta</i>				1.83		1.94
21	<i>Lonicera canadensis</i>				1.66		
22	<i>Uvularia grandiflora</i>					2.91	3.08
23	<i>Dirca palustris</i>					2.10	1.88
24	<i>Acer saccharum</i>					0.85	
25	<i>Ostrya virginiana</i>					0.77	
26	<i>Amphicarpa bracteata</i>						2.17
27	<i>Caulophyllum thalictroides</i>						3.21
28	<i>Celastrus scandens</i>						3.46
29	<i>Actaea rubra</i>						4.17
30	<i>Smilacina racemosa</i>						3.52
31	<i>Sanguinaria canadensis</i>						4.72
32	<i>Asarum canadense</i>						4.47

TABLE 16-4

## PERCENT FOLIAR CONCENTRATION OF MAGNESIUM, DRY WT. BASIS

Code No.	Species	Stand Number					
		1	2	3	4	5	6
01	<i>Lithospermum canescens</i>	0.226					
02	<i>Fragaria virginiana</i>	0.458	0.439	0.570			
03	<i>Pteridium aquilinum</i>	0.240	0.226	0.282			
04	<i>Cladonia sp.</i>	0.045	0.053				
05	<i>Epigaea repens</i>	0.228	0.270				
06	<i>Maianthemum canadense</i>	0.227	0.258	0.311	0.297	0.319	0.278
07	<i>Gaultheria procumbens</i>	0.290	0.314				
08	<i>Aster macrophyllus</i>	0.185	0.331	0.365	0.361	0.381	0.303
09	<i>Aralia nudicaulis</i>	0.272	0.283	0.377	0.394	0.352	0.329
10	<i>Arctostaphylos Uva-ursi</i>	0.193	0.210				
11	<i>Pyrola elliptica</i>	0.253	0.237				
12	<i>Pleurozium schreberii</i>	0.140	0.154				
13	<i>Apocynum androsaemifolium</i>	0.339					
14	<i>Corylus cornuta</i>		0.354	0.384	0.400	0.398	0.377
15	<i>Hepatica americana</i>		0.439	0.362	0.372	0.378	
16	<i>Diervilla lonicera</i>		0.398				
17	<i>Thalictrum dioicum</i>			0.430		0.427	0.365
18	<i>Cornus canadensis</i>			0.584	0.563		
19	<i>Clintonia borealis</i>			0.384	0.423	0.441	
20	<i>Parthenocissus inserta</i>				0.443		0.394
21	<i>Lonicera canadensis</i>				0.435		
22	<i>Uvularia grandiflora</i>					0.436	0.407
23	<i>Dirca palustris</i>					0.623	0.533
24	<i>Acer saccharum</i>					0.239	
25	<i>Ostrya virginiana</i>					0.384	
26	<i>Amphicarpa bracteata</i>						0.452
27	<i>Caulophyllum thalictroides</i>						0.235
28	<i>Celastrus scandens</i>						0.391
29	<i>Actaea rubra</i>						0.300
30	<i>Smilacina racemosa</i>						0.322
31	<i>Sanguinaria canadensis</i>						0.234
32	<i>Asarum canadense</i>						0.467

TABLE 16-5

P.P.H. FOLYAR CONCENTRATION OF MANGANESE, DRY WT. BASIS

Code No.	Species	Stand Number					
		1	2	3	4	5	6
01	<i>Lithospermum canescens</i>	100					
02	<i>Fragaria virginiana</i>	264	227	268			
03	<i>Pteridium aquilinum</i>	146	232	241			
04	<i>Cladonia</i> sp.	49	46				
05	<i>Epigaea repens</i>	334	342				
06	<i>Maianthemum canadense</i>	167	138	168	177	116	39
07	<i>Gaultheria procumbens</i>	231	237				
08	<i>Aster macrophyllus</i>	177	170	171	158	124	66
09	<i>Aralia nudicaulis</i>	450	389	373	412	233	117
10	<i>Arctostaphylos Uva-ursi</i>	28	34				
11	<i>Pyrola elliptica</i>	65	50				
12	<i>Pleurozium schreberii</i>	172	146				
13	<i>Apocynum androsaemifolium</i>	104					
14	<i>Corylus cornuta</i>		360		364	262	141
15	<i>Hepatica americana</i>		101	112	143	98	
16	<i>Diervilla lonicera</i>		124				
17	<i>Thalictrum dioicum</i>			98		66	48
18	<i>Cornus canadensis</i>			110	90		
19	<i>Clintonia borealis</i>			258	307		
20	<i>Parthenocissus inserta</i>				246		121
21	<i>Lonicera canadensis</i>				223		
22	<i>Uvularia grandiflora</i>					79	53
23	<i>Dirca palustris</i>					179	130
24	<i>Acer saccharum</i>					215	
25	<i>Ostrya virginiana</i>					272	
26	<i>Amphicarpa bracteata</i>						120
27	<i>Caulophyllum thalictroides</i>						93
28	<i>Celastrus scandens</i>						68
29	<i>Actaea rubra</i>						50
30	<i>Smilacina racemosa</i>						54
31	<i>Sanguinaria canadensis</i>						43
32	<i>Asarum canadense</i>						76

TABLE 16-6

## PERCENT FOLIAR CONCENTRATION OF CALCIUM, DRY WT. BASIS

Code No.	Species	Stand Number					
		1	2	3	4	5	6
01	<i>Rhodospermum canescens</i>	1.38					
02	<i>Fragaria virginiana</i>	1.53	1.26	1.63			
03	<i>Pteridium aquilinum</i>	0.48	0.56	0.60			
04	<i>Cladonia</i> sp.	0.08	0.08				
05	<i>Epigaea repens</i>	0.53	0.64				
06	<i>Maianthemum canadense</i>	0.77	0.91	1.20	1.13	1.15	1.05
07	<i>Gaultheria procumbens</i>	1.11	1.18				
08	<i>Aster macrophyllus</i>	1.39	1.38	1.46	1.35	1.49	1.30
09	<i>Aralia nudicaulis</i>	1.18	1.01	1.56	1.16	1.15	1.33
10	<i>Arctostaphylos Uva-ursi</i>	0.68	0.79				
11	<i>Pyrola elliptica</i>	1.02	1.02				
12	<i>Pleurozium schreberii</i>	0.38	0.37				
13	<i>Apocynum androsaemifolium</i>	1.06					
14	<i>Corylus cornuta</i>		1.35	1.56	1.41	1.56	1.64
15	<i>Hepatica americana</i>		1.27	1.36	1.24	1.31	
16	<i>Diervilla lonicera</i>		1.19				
17	<i>Thalictrum dioicum</i>			1.55		1.59	1.56
18	<i>Cornus canadensis</i>			2.20	2.02		
19	<i>Clintonia borealis</i>			1.43	1.41	1.40	
20	<i>Parthenocissus inserta</i>				1.72		1.76
21	<i>Lonicera canadensis</i>				1.75		
22	<i>Uvularia grandiflora</i>					1.27	1.28
23	<i>Birca palustris</i>					2.22	1.89
24	<i>Acer saccharum</i>					1.27	
25	<i>Ostrya virginiana</i>					1.61	
26	<i>Amphicarpa bracteata</i>						1.61
27	<i>Gaulophyllum thalictroides</i>						1.20
28	<i>Celastrus scandens</i>						1.83
29	<i>Actaea rubra</i>						1.80
30	<i>Smilacina racemosa</i>						1.36
31	<i>Sanguinaria canadensis</i>						1.29
32	<i>Asarum canadense</i>						1.23



TABLE 16-7

P.S.W. FOLIAR CONCENTRATION OF STRONTIUM, DRY WT. BASIS

Code No.	Species	Stand Number					
		1	2	3	4	5	6
01	<i>Lithospermum canescens</i>	47.4					
02	<i>Fragaria virginiana</i>	55.0	37.2	48.9			
03	<i>Pteridium aquilinum</i>	29.9	44.5	41.9			
04	<i>Gladonia</i> sp.	1.8	1.5				
05	<i>Epigaea repens</i>	11.0	8.5				
06	<i>Maianthemum canadense</i>	26.4	21.0	29.4	17.7	27.4	16.3
07	<i>Gaultheria procumbens</i>	15.8	14.2				
08	<i>Aster macrophyllus</i>	49.0	33.5	36.5	27.5	33.9	18.3
09	<i>Aralia nudicaulis</i>	44.2	21.6	43.7	18.7	26.6	22.1
10	<i>Arctostaphylos Uva-ursi</i>	30.9	20.1				
11	<i>Pyrola elliptica</i>	38.0	23.7				
12	<i>Pleurozium schreberii</i>	10.2	6.2				
13	<i>Apocynum androsaemifolium</i>	65.0					
14	<i>Corylus cornuta</i>		34.4	41.6	28.1	40.2	23.2
15	<i>Hepatica americana</i>		39.9	45.3	37.8	43.2	
16	<i>Diervilla lonicera</i>		36.1				
17	<i>Thalictrum dioicum</i>			32.0		32.1	21.1
18	<i>Cornus canadensis</i>			60.0	59.5		
19	<i>Clintonia borealis</i>			44.0	26.9	31.3	
20	<i>Parthenocissus inserta</i>				36.3		33.4
21	<i>Lonicera canadensis</i>				37.6		
22	<i>Uvularia grandiflora</i>					29.5	20.4
23	<i>Dirca palustris</i>					53.0	32.9
24	<i>Acer saccharum</i>					31.0	
25	<i>Ostrya virginiana</i>					41.6	
26	<i>Amphicarpa bracteata</i>						24.7
27	<i>Caulophyllum thalictroides</i>						16.0
28	<i>Celastrus scandens</i>						35.6
29	<i>Actaea rubra</i>						32.2
30	<i>Smilacina racemosa</i>						24.5
31	<i>Sanguinaria canadensis</i>						22.5
32	<i>Asarum canadense</i>						47.7

TABLE 16-3

## P.P.M. FOLIAR CONCENTRATION OF ZINC, DRY WEIGHT BASIS

Code No.	Species	Stand Number					
		1	2	3	4	5	6
01	<i>Lithospermum canescens</i>	39.5					
02	<i>Fragaria virginiana</i>	48.7	40.1	35.6			
03	<i>Pteridium aquilinum</i>	31.8	35.5	30.1			
04	<i>Cladonia</i> sp.	43.2	42.9				
05	<i>Epigaea repens</i>	37.2	39.2				
06	<i>Maianthemum canadense</i>	47.3	37.8	32.1	62.9	41.6	55.3
07	<i>Gaultheria procumbens</i>	30.7	29.1				
08	<i>Aster macrophyllus</i>	84.9	64.5	57.2	72.3	69.5	69.5
09	<i>Aralia nudicaulis</i>	82.5	70.3	60.9	80.0	63.3	68.0
10	<i>Arctostaphylos Uva-ursi</i>	70.7	86.9				
11	<i>Pyrola elliptica</i>	49.6	36.9				
12	<i>Pleurozium schreberii</i>	68.6	71.4				
13	<i>Apocynum androsaemifolium</i>	33.8					
14	<i>Corylus cornuta</i>		30.5	31.5	34.7	33.6	31.5
15	<i>Hepatica americana</i>		43.5	38.2	52.2	51.0	
16	<i>Diervilla lonicera</i>		109.6				
17	<i>Thalictrum dioicum</i>			44.2		50.5	50.8
18	<i>Cornus canadensis</i>			29.6	46.1		
19	<i>Clintonia borealis</i>			21.7	32.3	44.1	
20	<i>Parthenocissus inserta</i>				26.2		33.0
21	<i>Lonicera canadensis</i>				64.8		
22	<i>Uvularia grandiflora</i>					56.6	40.3
23	<i>Birca palustris</i>					63.9	39.6
24	<i>Acer saccharum</i>					41.3	
25	<i>Ostrya virginiana</i>					25.8	
26	<i>Amphicarpa bracteata</i>						43.0
27	<i>Gaulophyllum thalictroides</i>						35.2
28	<i>Celastrus scandens</i>						39.3
29	<i>Actaea rubra</i>						33.6
30	<i>Smilacina racemosa</i>						22.0
31	<i>Sanguinaria canadensis</i>						40.0
32	<i>Acarum canadense</i>						107.8

TABLE 16-3

## P.P.M. FOLIAR CONCENTRATION OF COPPER, DRY WT. BASIS

Code No.	Species	Stand Number					
		1	2	3	4	5	6
01	<i>Lithospermum canescens</i>	8.50					
02	<i>Fragaria virginiana</i>	8.20	5.67	6.86			
03	<i>Pteridium aquilinum</i>	5.56	5.18	5.45			
04	<i>Gladonia</i> sp.	3.60	3.71				
05	<i>Epigaea repens</i>	6.00	7.18				
06	<i>Maianthemum canadense</i>	4.97	4.69	4.75	5.62	4.58	4.86
07	<i>Gaultheria procumbens</i>	5.07	4.75				
08	<i>Aster macrophyllus</i>	8.63	6.86	8.15	7.51	6.81	7.13
09	<i>Aralia nudicaulis</i>	5.73	5.56	7.24	6.48	5.40	5.24
10	<i>Arctostaphylos Uva-ursi</i>	5.56	6.10				
11	<i>Pyrola elliptica</i>	6.64	5.51				
12	<i>Pleurozium schreberii</i>	6.05	6.54				
13	<i>Apocynum androsemifolium</i>	7.34					
14	<i>Corylus cornuta</i>		6.10	7.40	7.61	5.83	5.89
15	<i>Hepatica americana</i>		6.75	7.45	7.29	6.00	
16	<i>Diervilla lonicera</i>		6.00				
17	<i>Thalictrum dioicum</i>			6.64		5.78	6.54
18	<i>Cornus canadensis</i>			4.15	3.87		
19	<i>Clintonia borealis</i>			5.67	5.56	5.07	
20	<i>Parthenocissus inserta</i>				8.09		9.27
21	<i>Lonicera canadensis</i>				5.89		
22	<i>Utricularia grandiflora</i>					6.16	6.48
23	<i>Dirca palustris</i>					6.48	5.56
24	<i>Acer saccharum</i>					4.91	
25	<i>Ostrya virginiana</i>					6.05	
26	<i>Amphicarpa bracteata</i>						6.05
27	<i>Caulophyllum thalictroides</i>						6.10
28	<i>Celastrus scandens</i>						7.29
29	<i>Actaea rubra</i>						7.51
30	<i>Smilacina racemosa</i>						5.18
31	<i>Sanguinaria canadensis</i>						5.40
32	<i>Asarum canadense</i>						5.35

TABLE 16-10

PPM FOLIAR CONCENTRATION OF BORON, DRY WEIGHT BASIS							
Code		Stand Number					
No.	Species	1	2	3	4	5	6
01	<i>Lithospermum</i>						
	<i>canescens</i>	31.3					
02	<i>Fragaria</i>	24.0	21.5	27.6			
	<i>virginiana</i>						
03	<i>Rberidium</i>						
	<i>aquilinum</i>	10.6	15.8	13.8			
04	<i>Gledonia</i> sp.	12.5	3.0				
05	<i>Epigaea repens</i>	18.4	22.7				
06	<i>Helianthemum</i>	12.1	14.7	16.3	16.3	15.7	17.6
	<i>canadense</i>						
07	<i>Gaultheria</i>						
	<i>procumbens</i>	17.5	12.4				
08	<i>Aster</i>						
	<i>macrophyllus</i>	31.1	33.8	45.9	41.8	41.3	42.7
09	<i>Aralia</i>						
	<i>nudicaulis</i>	24.3	20.6	25.2	18.5	19.2	19.4
10	<i>Arctostaphylos</i>						
	<i>uva-ursi</i>	15.5	15.6				
11	<i>Pyrola elliptica</i>	16.0	9.6				
12	<i>Pleurozium</i>						
	<i>schroberii</i>	31.5	5.3				
13	<i>Apocynum</i>						
	<i>androsaemifolium</i>	12.3					
14	<i>Corylus cornuta</i>		33.9	40.7	41.8	27.6	48.0
15	<i>Hepatica americana</i>		21.4	22.0	22.5	20.8	
16	<i>Diervilla lonicera</i>		30.8				
17	<i>Thalictrum dioicum</i>			27.7		20.4	25.9
18	<i>Cornus canadensis</i>			38.0	25.6		
19	<i>Clintonia borealis</i>			13.6	14.2	14.7	
20	<i>Parthenocissus inserta</i>				19.1		22.9
21	<i>Lonicera canadensis</i>				32.3		
22	<i>Uvularia grandiflora</i>					12.4	16.8
23	<i>Dirca palustris</i>					33.4	30.1
24	<i>Acer saccharum</i>					27.0	
25	<i>Ostrya virginiana</i>					16.0	
26	<i>Amphicarpa bracteata</i>						34.4
27	<i>Caulophyllum thalictroides</i>						23.8
28	<i>Celastrus scandens</i>						27.5
29	<i>Actaea rubra</i>						24.0
30	<i>Smilacina racemosa</i>						20.2
31	<i>Sanguinaria canadensis</i>						38.0
32	<i>Asarum canadense</i>						18.2

TABLE 17-1

## FOLIAR CONCENTRATION OF MACROELEMENTS FROM JULY SAMPLING

No.	Species	Stand Number					
		1	2	3	4	5	6
		Percent Nitrogen					
06	<i>Maianthemum canadense</i>	1.84	1.96	1.94	1.81	1.98	
08	<i>Aster macrophyllus</i>	1.86	1.86	2.00	2.01	1.97	1.93
09	<i>Aralia nudicaulis</i>		2.01	1.96	2.31	1.77	1.71
14	<i>Corylus cornuta</i>		2.56	2.54	2.74	2.33	2.58
		Percent Phosphorus					
06	<i>Maianthemum canadense</i>	0.345	0.336	0.332	0.276	0.370	
08	<i>Aster macrophyllus</i>	0.280	0.312	0.266	0.300	0.369	0.358
09	<i>Aralia nudicaulis</i>		0.300	0.340	0.307	0.323	0.307
14	<i>Corylus cornuta</i>		0.285	0.273	0.300	0.309	0.312
		Percent Potassium					
06	<i>Maianthemum canadense</i>	2.41	3.16	3.61	3.06	3.50	
08	<i>Aster macrophyllus</i>	4.01	4.70	3.62	4.16	4.60	4.27
09	<i>Aralia nudicaulis</i>		1.86	2.17	1.86	2.61	2.21
14	<i>Corylus cornuta</i>		0.97	1.02	1.61	1.24	1.33
		Percent Calcium					
06	<i>Maianthemum canadense</i>	0.79	0.78	0.94	0.82	1.13	
08	<i>Aster macrophyllus</i>	1.18	1.20	1.16	1.23	1.36	1.26
09	<i>Aralia nudicaulis</i>		0.85	0.82	0.80	0.83	0.85
14	<i>Corylus cornuta</i>		1.26	1.30	1.31	1.69	1.58
		Percent Magnesium					
06	<i>Maianthemum canadense</i>	0.280	0.285	0.296	0.246	0.328	
08	<i>Aster macrophyllus</i>	0.266	0.329	0.377	0.330	0.399	0.345
09	<i>Aralia nudicaulis</i>		0.305	0.257	0.333	0.310	0.306
14	<i>Corylus cornuta</i>		0.430	0.411	0.395	0.497	0.421

TABLE 17-2

## FOLIAR CONCENTRATION OF MICROELEMENTS FROM JULY SAMPLING

No.	Species	Stand Number					
		1	2	3	4	5	6
		Parts Per Million Manganese					
06	<i>Maianthemum canadense</i>	161	117	134	126	102	
08	<i>Aster macrophyllus</i>	130	115	105	142	105	59
09	<i>Aralia nudicaulis</i>		272	219	260	170	107
14	<i>Corylus cornuta</i>		392	393	357	360	181
		Parts Per Million Strontium					
06	<i>Maianthemum canadense</i>	31.4	17.4	23.3	15.7	25.3	
08	<i>Aster macrophyllus</i>	43.7	30.3	24.3	31.6	30.4	20.5
09	<i>Aralia nudicaulis</i>		15.8	18.6	14.8	17.0	14.4
14	<i>Corylus cornuta</i>		31.5	31.9	28.7	45.1	34.8
		Parts Per Million Zinc					
06	<i>Maianthemum canadense</i>	56.5	37.8	38.3	47.0	47.5	
08	<i>Aster macrophyllus</i>	57.5	55.0	50.2	72.7	72.6	69.4
09	<i>Aralia nudicaulis</i>		48.7	42.6	47.8	41.0	42.4
14	<i>Corylus cornuta</i>		32.5	29.6	38.9	34.9	31.5
		Parts Per Million Copper					
06	<i>Maianthemum canadense</i>	5.29	4.80	5.29	4.69	5.45	
08	<i>Aster macrophyllus</i>	7.08	6.81	7.88	6.21	6.75	6.43
09	<i>Aralia nudicaulis</i>		6.48	5.67	6.16	5.83	5.18
14	<i>Corylus cornuta</i>		6.98	6.91	8.42	7.18	7.99
		Parts Per Million Boron					
06	<i>Maianthemum canadense</i>	12.5	14.6	13.4	13.7	16.2	
08	<i>Aster macrophyllus</i>	43.2	32.1	31.4	34.8	40.6	42.9
09	<i>Aralia nudicaulis</i>		20.2	18.0	17.4	16.7	17.3
14	<i>Corylus cornuta</i>		34.9	32.9	36.0	34.7	45.5

TABLE 18

COMPARISON OF NUTRIENT CONCENTRATIONS IN FOLIAGE  
WITH PREVIOUSLY REPORTED DATA

Species	Source <sup>1</sup>	N	P	K	Ca	Mg
<i>Acer saccharum</i>	S	0.44	0.14	0.52	1.66	
"	G	0.73	0.12	0.39	1.01	0.45
"	C	1.86	0.28	0.85	1.27	0.24
<i>Apocynum androsaemifolium</i>	G	1.45	0.28	2.12	0.81	0.27
"	G	1.35	0.33	2.43	0.78	0.18
"	C	2.31	0.34	1.85	1.06	0.34
<i>Aralia nudicaulis</i>	G	1.66	0.29	2.53	0.83	0.33
"	G	2.06	0.29	1.51	0.94	0.43
"	G.	1.97	0.32	2.07	1.13	0.26
"	S	0.56	0.22	0.79	1.83	
"	C	1.71	0.27	1.38	1.18	0.27
"	C	1.88	0.32	1.76	1.01	0.28
"	C	2.07	0.45	2.36	1.56	0.38
"	C	2.43	0.38	2.08	1.16	0.39
"	C	1.69	0.39	2.79	1.15	0.35
"	C	1.88	0.37	2.94	1.33	0.33
<i>Caulophyllum thalictroides</i>	G	1.83	0.31	2.09	0.82	0.30
"	C	2.41	0.52	3.21	1.20	0.24
<i>Diervilla lonicera</i>	G	1.31	0.23	2.18	0.73	0.26
"	C	2.03	0.45	2.86	1.19	0.40

<sup>1</sup>"G" indicates Gerloff, 1964; "S" indicates Scott, 1955; "C" indicates the present study.

TABLE 18--Continued

Species	Source	N	P	K	Ca	Mg
<i>Cornus canadensis</i>	G	1.52	0.25	1.14	0.85 <sup>1</sup>	0.68
"	C	1.73	0.37	1.78	2.20	0.58
"	C	1.69	0.23	1.33	2.02	0.56
<i>Epigaea repens</i>	C	1.47	0.13	0.66	0.53	0.23
"	C	1.49	0.17	1.03	0.64	0.27
"	G	1.02	0.10	0.52	0.44	0.27
<i>Gaultheria procumbens</i>	G	0.87	0.09	0.46	0.96	0.46
"	C	1.27	0.13	0.79	1.11	0.29
"	C	1.31	0.14	0.73	1.18	0.31
"	S	1.38	0.12	1.23	1.06	
<i>Clintonia borealis</i>	G	1.76	0.31	5.54	1.19	0.28
"	C	2.04	0.45	4.53	1.43	0.38
"	C	2.16	0.39	4.55	1.41	0.42
"	C	1.91	0.50	4.83	1.40	0.44
<i>Smilacina racemosa</i>	G	1.82	0.33	3.57	1.32	0.22
"	C	2.24	0.45	3.52	1.36	0.32
"	S	1.91	0.17	3.73	1.82	
<i>Uvularia grandiflora</i>	G	1.87	0.28	3.09	1.11	0.24
"	C	2.07	0.29	2.91	1.27	0.44
"	C	2.38	0.34	3.08	1.28	0.41

<sup>1</sup>This value appears disproportionately low.



TABLE 18--Continued

Species	Source	N	P	K	Ca	Mg
<i>Sanguinaria</i> <i>canadensis</i>	G	1.49	0.33	5.73	1.77	0.12
"	S	1.96	0.18	2.70	2.45	
"	C	1.81	0.53	4.72	1.29	0.23
<i>Pteridium</i> <i>aquilinum</i>	G	2.00	0.25	1.76	0.26	0.22
"	G	1.56	0.24	2.50	0.42	0.28
"	G	1.78	0.25	2.68	0.26	0.17
"	C	1.83	0.31	1.88	0.48	0.24
"	C	1.92	0.32	2.27	0.56	0.23
"	C	1.93	0.38	2.37	0.60	0.28
<i>Hepatica</i> <i>americana</i>	G	1.71	0.25	3.36	1.15	0.33
"	C	1.81	0.24	2.90	1.27	0.44
"	C	1.84	0.26	3.03	1.36	0.36
"	C	1.94	0.20	2.31	1.24	0.37
"	C	1.86	0.24	2.76	1.31	0.38
<i>Fragaria</i> <i>virginiana</i>	G	1.44	0.33	1.74	1.02	0.50
"	G	1.29	0.35	1.26	1.22	0.32
"	C	2.28	0.31	1.68	1.53	0.46
"	C	2.21	0.32	1.59	1.21	0.44
"	C	2.37	0.38	1.96	1.63	0.57
<i>Aster</i> <i>macrophyllus</i>	G	1.78	0.24	4.74	1.00	0.39
"	G	1.76	0.25	5.78	1.48	0.23
"	G	1.90	0.29	4.82	1.41	0.26

TABLE 18--Continued

Species	Source	N	P	K	Ca	Mg
Aster macrophyllus	C	1.66	0.30	3.61	1.39	0.19
"	C	1.91	0.31	3.94	1.38	0.33
"	C	1.83	0.35	4.59	1.46	0.37
"	C	2.23	0.27	3.54	1.35	0.36
"	C	1.87	0.33	4.37	1.49	0.38
"	C	1.93	0.39	3.96	1.30	0.30
Maianthemum canadense	S	1.61	0.26	2.68	1.46	
"	C	1.73	0.26	3.27	1.05	
"	C	1.69	0.31	2.70	1.15	
"	C	1.86	0.33	3.36	1.13	
"	C	2.01	0.31	3.19	1.20	
"	C	1.97	0.30	2.79	0.91	
"	C	1.93	0.41	1.94	0.77	
Corylus cornuta	G	1.52	0.17	0.65	1.08	0.23
"	C	2.33	0.30	0.84	1.35	0.35
"	C	2.30	0.38	1.30	1.56	0.39
"	C	2.56	0.29	1.14	1.41	0.40
"	C	2.14	0.28	1.11	1.56	0.40
"	C	2.28	0.31	1.04	1.64	0.38
Ostrya virginiana	G	1.49	0.16	0.64	1.06	0.54
"	G	1.44	0.17	0.43	1.03	0.58
"	C	2.21	0.24	0.77	1.61	0.38

collected for this study. In general, however, the values reported by Gerloff are slightly lower than the values reported here. This is particularly true for nitrogen, phosphorus and calcium. The values reported by Scott (1955), however, are frequently higher. These differences may be due to time of sampling, method of sampling or geographic variables.

There is no simple correlation of nutrient concentration in a given species with synecological nutrient coordinates. There appears, however, to be an optimum range for nitrogen accumulation occurring near the center of the edaphic field (see Figure 7). For the ubiquitous species, the nitrogen concentration is low in the jack pine stands, increases in the red pine-fir stands and again drops in the hardwood stands. This suggests that the importance of nitrogen as a factor in the growth of understory plants varies between stands. It is not possible, however, to infer exactly how the role of nitrogen varies. For example, it is possible that available nitrogen was a limiting factor in the jack pine stands, but was in ample supply in the red pine stands, particularly the Wilderness stand. The soil analysis data show a relative increase of nitrogen in the surface horizon. If soil moisture did not increase concomitantly, the foliar nitrogen content could have increased. Moving to the hardwood stands, an increase of available moisture could again make nitrogen the limiting factor, thus resulting in a decrease in foliar nitrogen concentration. Obviously many other factor combinations could have resulted in the observed pattern.

# FOLIAR NITROGEN IN FOREST PLANTS ALONG AN ECOLOGICAL NUTRIENT GRADIENT

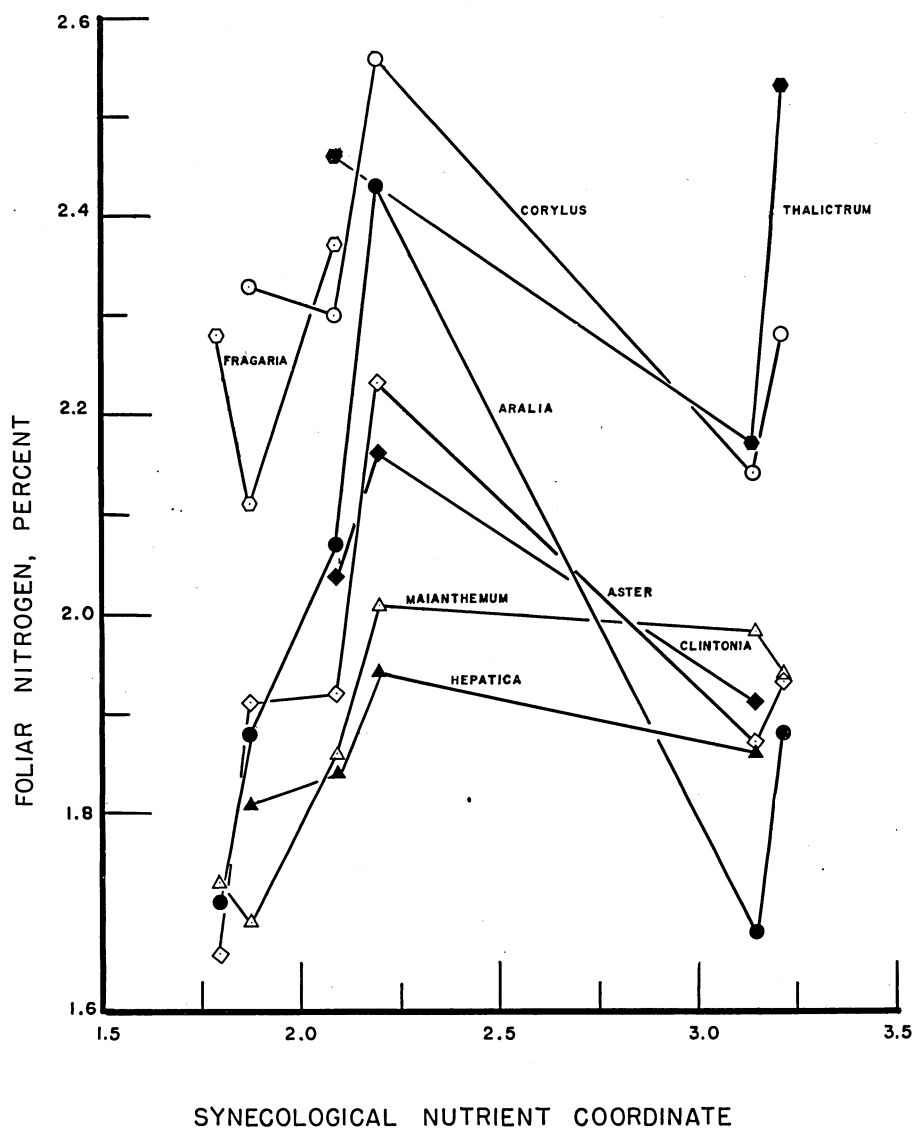


Figure 7. Foliar nitrogen concentration in eight forest plants from six stands. The foliar nitrogen concentration was highest in the red pine-fir stands and lowest in the jack pine and northern hardwood stands. The stands are plotted along the abscissa (from left to right) in the following order: 1) Woodlot jack pine, 2) Scott's jack pine, 3) Mary Lake red pine-fir, 4) Wilderness red pine-fir, 5) Campground maple-basswood and 6) Bearpaw Point basswood-elm.

No consistent site effect was observed for potassium, phosphorus, calcium or magnesium concentrations in the plants sampled.

It appeared on inspection that the species sampled at the upper end of the nutrient gradient had a higher average nitrogen, phosphorus, and potassium content than those at the lower end of the gradient. In order to test this hypothesis, Duncan's new multiple range test (Duncan, 1955) was used. When this test was run on all samples for the nitrogen data, no significant differences between stands were detected. The sampling, however, was designed to include the four ubiquitous species wherever possible. Since the nutrient content of these species does not vary consistently between stands, their inclusion in the multiple range test would mask significant differences in the nutrient content of species that segregate toward the ends of a nutrient gradient. Accordingly, these four species were dropped and the test was re-run for nitrogen, phosphorus and potassium.

The results indicate that the species confined to nutrient-rich stands in general have a higher foliage nutrient concentration than species confined to poor stands. The difference in this case is greatest between the pure basswood stand on Bearpaw Point and the two jack pine stands.

Figure 8 shows graphically the significant differences between stands. A line under two stands indicates lack of significance. Since an unequal number of species for each stand were used in the multiple range test, two least significant

Figure 8

DUNCAN'S NEW MULTIPLE RANGE TEST  
ON FOLIAR NUTRIENT CONCENTRATIONS

1. Potassium

Bearpaw Mary L. Campground Wilderness Scott's Woodlot

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2. Phosphorus

Bearpaw Mary L. Campground Wilderness Scott's Woodlot

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3. Nitrogen

Bearpaw Wilderness Campground Mary L. Scott's Woodlot

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A solid line under two plots indicates no significant difference between the mean foliage nutrient concentrations for the species sampled on those plots. A dotted line indicates a doubtfully significant difference.

ranges were computed for each comparison. A dotted line in Figure 8 indicates that the observed difference fell between the two computed test statistics and the difference is therefore of doubtful significance. The differences between stands are most pronounced for phosphorus and potassium. The differences in average nitrogen concentrations, although significant, are not as great.

In order to explore the interrelationships between different nutrients and draw conclusions as to their relative importance, the foliar analysis data were plotted in triangular coordinates. The use of triangular coordinates makes it possible to plot simultaneously the relationship between three different nutrients in a given sample. The plotting method is similar to that used by the "foliar diagnosis school." Thomas (1939; 1943) used triangular coordinates to show the effect of fertilizer and weather on the N-P-K coordinates and K-Mg-Ca coordinates. Plants were grown under different conditions and sampled at regular intervals. Rather than sampling whole plants, however, Thomas sampled homologous leaves from different plants on the same plot.

Holland (1966) compared the triangular coordinate method with analysis of variance, ratios of pairs of elements and Principal Component Analysis. Each method was judged by its ability to give consistent results in fertilizer trials from year to year. As a technique for interpreting results, Holland found that triangular coordinates were superior to both the analysis of variance and comparisons of ratios. Since both a

statistical analysis and use of more than three elements at a time is impossible with this method, Holland concluded that the method of Principal Component Analysis was preferable to the triangular coordinate method that was used in this study.

In order to plot widely varying values in triangular coordinates, the data from this study were adjusted mathematically. The technique used has been discussed with respect to the humus samples.

When different combination of the nutrients nitrogen, phosphorus, potassium, calcium and magnesium are centered and plotted in triangular coordinates, some interesting relationships are suggested. Although no site effects within species are discernible, individual species occupy different positions within the triangular fields. This is particularly true for combination of N-P-K, K-Ca-Mg, P-K-Ca, and N-K-Ca (Figures 9-1 through 9-4). In each of these cases, most of the spread is along the potassium axis. When potassium is dropped from consideration, as with N-Ca-P, then the species tend to overlap near the center of the triangle (Figure 9-5). The finding that species differences are much stronger than site effects within species is consistent with the results of Scott (1955), Gagnon (1958) and others.

If all species are plotted in triangular nutrient coordinates with potassium as one of the axes, then the points are well spread along the potassium axis, as in Figures 10-1 and 10-2. If potassium is excluded, however, (as in Figure 10-3), then the result is a scatter diagram. Furthermore, species from



### FOLIAR NUTRIENT CONTENT OF FOUR FOREST PLANTS IN TRIANGULAR COORDINATES

- ASTER MACROPHYLLUS
- ◇ MAIANthemum CANADENSE
- △ ARALIA NUDICAULIS
- CORYLUS CORNUTA

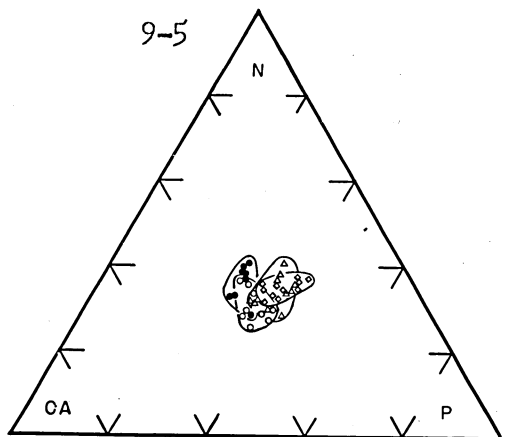
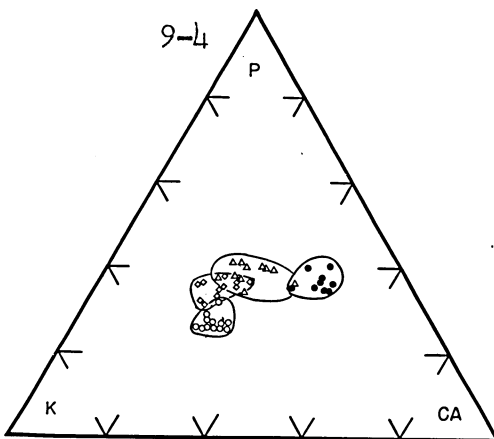
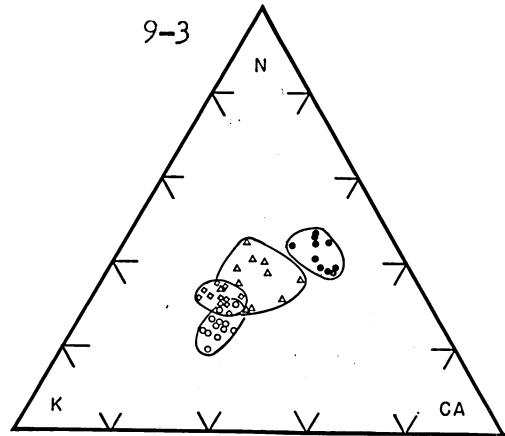
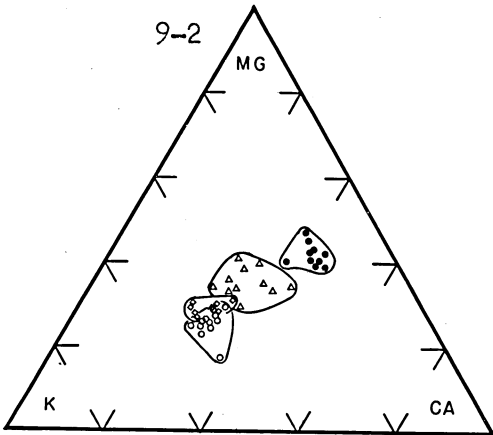
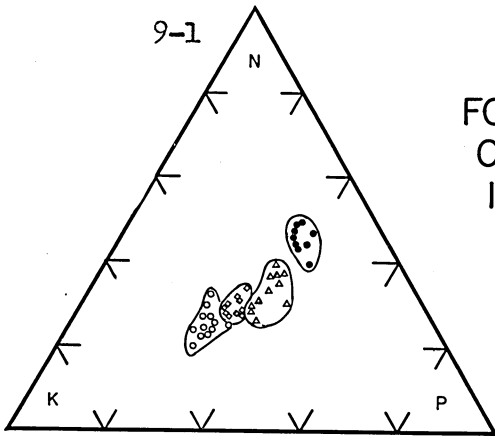
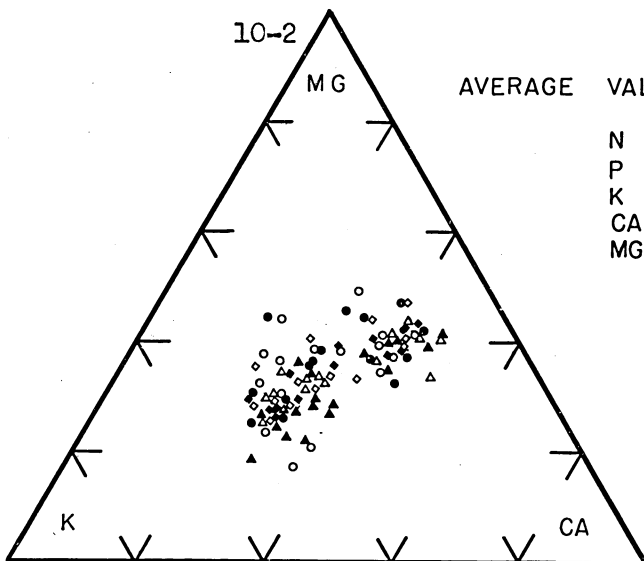
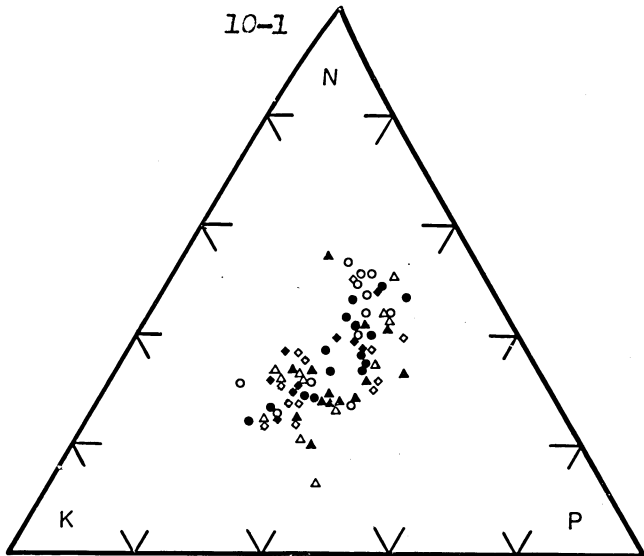


Figure 9. Relative foliar nutrient content of four ubiquitous forest plants in five sets of triangular coordinates. The data plotted here were centered with respect to the 94 foliage samples that were analyzed. Differences between species in triangular nutrient coordinates are most pronounced if potassium is one of the axes.

# FOLIAR NUTRIENT CONTENT OF FOREST PLANTS FROM SIX STANDS IN TRIANGULAR COORDINATES

- ▲ BEARPAW POINT HARDWOODS
- △ CAMPGROUND HARDWOODS
- ◆ WILDERNESS RED PINE-FIR
- ◇ MARY LAKE RED PINE-FIR
- SCOTT'S JACK PINE
- WOODLOT JACK PINE



AVERAGE VALUES, PERCENT

N = 2.01  
 P = 0.312  
 K = 2.45  
 CA = 1.23  
 MG = 0.340

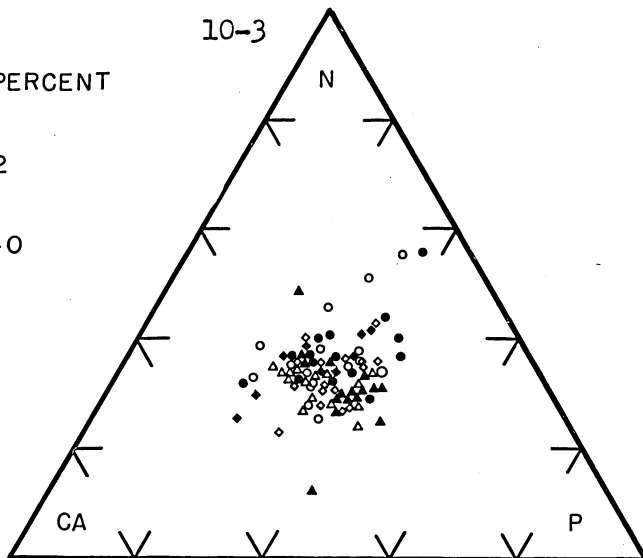


Figure 10. Relative foliar nutrient content of 32 species from six stands centered in three sets of triangular nutrient coordinates. The spread is greatest along the potassium axis.

the hardwood stands appear to be slightly clustered toward the upper end of the potassium axis, whereas species from the jack pine stands are well distributed toward the lower end.

In the N-P-K coordinates, it appears that most of the species fall into two groups along the potassium axis. These groups were arbitrarily divided at 30 percent potassium. The species in each group are listed in Table 19. The high potassium group contains only those species which occur in the middle and upper end of the nutrient gradient, that is, nutrient demanding species, such as Sanguinaria canadensis, and ubiquitous, such as Maianthemum canadense and Aster macrophyllus. The only exceptions are Lithospermum canescens, which is confined to acid xeric sites and Aralia nudicaulis, which falls into both groups. The "low potassium" group, however, contains only species that are confined to the low end of the nutrient gradient, and trees, shrubs and vines. The only exceptions are Cornus canadensis, which is confined to the middle range of the nutrient gradient, and Amhicarpa bracteata. The low relative potassium value for this leguminous species, however, reflects the high internal nitrogen concentration.

If the species are grouped according to relative phosphorus content, then the same trend appears: species from "rich" stands have a higher relative phosphorus content than species from poor stands. The grouping, however, is not as clear as with potassium. Thus species restricted to rich stands have a higher foliar concentration of nutrients; this difference decreases in the order  $K > P > N$ .

TABLE 19

CLASSIFICATION OF SPECIES BY POTASSIUM CONTENT  
RELATIVE TO PHOSPHORUS AND NITROGEN

K-Coordinate above 30 percent

Lithospermum canescens  
 Pteridium aquilinum  
 Maianthemum canadense  
 Aster macrophyllus  
 Aralia nudicaulis  
 Hepatica americana  
 Thalictrum dioicum  
 Clintonia borealis  
 Uvularia grandiflora  
 Caulophyllum thalictroides  
 Actea rubra  
 Smilacina racemosa  
 Sanguinaria canadensis  
 Asarum canadensis

K-Coordinate below 30 percent

Fragaria virginiana  
 Cladonia sp.  
 Epigea repens  
 Gaultheria procumbens  
 Aralia nudicaulis  
 Arctostaphylos uva-ursi  
 Pyrola elliptica  
 Pleurozoum schreberii  
 Apocynum androsaemifolium  
 Cornus canadensis  
 Amphicarpa bracteata  
Shrubs, trees and vines  
 Corylus cornuta  
 Dirca palustris  
 Parthenocissus inserta  
 Ostrya virginiana  
 Acer saccharum  
 Lonicera canadensis

It is unclear, however, whether these differences reflect the physiological ability of the different species to concentrate nutrients, or whether they are the result of deficiencies and luxury uptake. Possibly all three factors are involved. The four ubiquitous species generally had high nutrient concentration, regardless of where they were growing. Thus these plants appear to exceed the low-nutrient species in their ability to extract and concentrate nutrients in their foliage. The same may be true for plants that are restricted to rich stands. This does not imply, however, that the supply of nutrients in the jack pine stands was optimal for the plants growing there. More likely the low minimum nutrient requirements of these species allow them to segregate at the low end of the nutrient gradient under competition.

On a basis of commonly reported optimum and minimum foliar nutrient values, however, the nutrient concentration in some plants certainly exceeded the optimum concentration for growth (Goodall and Gregory, 1947). Several species from Bearpaw Point, for example, had phosphorus concentrations in excess of 0.5 percent and potassium concentrations greater than 4.0 percent. These values are greater than any reported by Goodall and Gregory as optimum for agricultural plants and forest trees. The nitrogen values, however, do not greatly exceed the commonly reported values. Possibly nitrogen is the limiting nutrient in the hardwood stands. This is supported by the N-P-K coordinates in Figure 10-1. Species from the hardwood stands are displaced

further from the nitrogen vertex than species from the jack pine stands.

In the humus samples the ratio of calcium plus magnesium to potassium increased toward the upper end of the nutrient gradient. This pattern was not observed for the foliar samples. Furthermore, foliar potassium showed the greatest site effect of any nutrients, whereas the humus potassium was only weakly correlated with synecological nutrient coordinates. Thus the chemical structure of the ecosystems sampled is strongly affected by the physiology of the subordinate forest plants.

As with the humus data, a simple correlation analysis was run between the absolute and relative foliar nutrient concentrations (Table 20). Of the five macronutrients correlated, calcium vs. magnesium had the highest correlation coefficient (+0.703). Potassium was better correlated with phosphorus than with any other element. This is consistent with the finding that nutrient concentration differences between sites are strongest for potassium and second strongest for phosphorus.

In the N-P-K coordinates, the correlation between relative and absolute values decreased in the order  $K > P > N$ . In the K-Ca-Mg coordinates, the order was  $K > Ca > Mg$ . This is probably an effect of differing variances between elements.

Although the four ubiquitous species were sampled twice during the summer, consistent changes in nutrient concentration are hard to detect. This is probably due to sampling error. Three generalizations, however, are possible. First, the calcium

TABLE 20

## CORRELATION COEFFICIENTS BETWEEN FOLIAR NUTRIENT VALUES

Absolute, percent	Absolute, percent				
	N	P	K	Ca	Mg
N		+0.397**	+0.003	+0.338**	+0.552**
P			+0.557**	+0.403**	+0.424**
K				+0.204*	+0.090
Ca					+0.703**

Absolute, percent	Relative values, N-P-K Coordinates		
	N	P	K
N	+0.226*	+0.092	-0.243*
P	-0.605**	+0.509**	+0.284**
K	-0.874**	-0.294**	+0.908**

Absolute, percent	Relative Values, K-Ca-Mg Coordinates		
	K	Ca	Mg
K	+0.813**	-0.482**	-0.859**
Ca	-0.281**	+0.560**	-0.098
Mg			+0.225*

concentration increased almost uniformly, both in absolute concentration and relative to other nutrients. This is consistent with the findings of Mitchell (1936), Thomas (1967) and others. Second, an increase in concentration was more frequent in the red pine stands than in the hardwood and jack pine stands. Third, the concentration of all nutrients in Aralia nudicaulis increased, whereas Maianthemum canadense, Aster macrophyllus and Corvulus cornuta were erratic.

## 7. Factor Interaction

Although this study was primarily concerned with the evaluation of a nutrient gradient, it must be stressed that nutrients do not function alone in ecosystems but rather in interaction with other factors. The relationships and interactions between the moisture, nutrient, heat and light regimes are suggested by the matrix of simple correlation coefficients between the four coordinates and the humus exchangeable cations (Table 14).

The moisture coordinates show a high degree of correlation with the nutrient coordinates and exchangeable bases. Exchangeable potassium, however, is more highly correlated with moisture than any of the other exchangeable bases. This is especially interesting in view of Bard's (1949) observation that plants with the highest moisture requirements also had high potassium requirements. The foliar potassium data from this study suggest a correlation between synecological moisture coordinates and potassium uptake.



The mechanical analysis data from the mineral soil samples suggest conclusions about the moisture regime. On a basis of silt plus clay content, the moisture holding capacity in the profiles sampled decreases in the order: Campground > Bearpaw > Wilderness = Mary Lake > Scott's > Woodlot. The interpretation is complicated, however, by the presence of thin bands of silt and very fine sand in the Marquette profiles and by the high organic matter content of the Poppleton profile. In view of the local variability in the soils that were sampled, a much larger number of samples is needed to adequately assess the moisture regime.

The synecological light coordinates are negatively correlated with nutrients. The negative correlation is highest for the synecological nutrient coordinates and next highest for percent base saturation. Exchangeable potassium was not as highly correlated with light as were calcium and magnesium. This is somewhat disappointing in view of Bergmann's (1958) observation that potassium requirements are higher at low light intensities. It appears, however, that foliar potassium is more strongly negatively correlated with either actual light measurements or light coordinates than is either calcium or magnesium. Thus the high potassium content of plants in the rich stands may be related to both low light intensities and high moisture requirements.

The synecological heat coordinates are positively correlated with the nutrient regime. The correlation is best for the integrative measurements of nutrients, that is, nutrient-coordinates,

pH, exchangeable hydrogen and percent base saturation. Calcium and magnesium are also positively correlated with heat coordinates but potassium is not. This appears to be related to the increase in the K : Ca:Mg ratio in the nutrient poor (jack pine) stands which have low synecological heat coordinates. It must be stressed, however, that the "heat regime" in this case is based only on the synecological heat coordinates. The coordinates do not necessarily reflect significant differences in effective temperature between stands, since the heat coordinates were developed on a basis of regional distribution of the individual species.

More detailed conclusions on the interactions and relationships between the effective environmental factors are beyond the scope of this study. A complete and integrated picture must await proper field instrumentation.

## V. SUMMARY AND CONCLUSIONS

The purpose of this study was to relate the nutrient content of soils and subordinate forest vegetation to an ecological nutrient gradient in the Central Pine Section of Minnesota. The nutrient gradient was defined in terms of synecological coordinates and was described by means of soil and site description, soil chemical analysis and foliar analysis.

Six stands were selected for the study, with two stands in each of three vegetation types: jack pine, red pine-balsam fir and northern hardwoods. Eight subplots were located within each stand.

Humus samples were collected at each subplot and analyzed for exchangeable cations and pH. Percent base saturation was more highly correlated with synecological nutrient coordinates than any of the other humus parameters ( $r = +0.83$ ). The degree of correlation with nutrient coordinates decreased in the order: percent base saturation > pH > exchangeable H > exchangeable Mg = exchangeable Ca > exchangeable K.

The relatively poor correlation between potassium and synecological nutrient coordinates is due to the decrease in the K : Ca plus Mg ratio in the nutrient-rich hardwood stands. This relationship has been illustrated by means of triangular coordinates.

The good correlation between nutrient status of the humus layer and synecological coordinates obtained in this study suggests that the humus layer occupies a key position in the forest ecosystem. It appears to be an important factor in controlling plant distribution and productivity, and its characteristics are determined by the vegetation itself. Future work on ecological nutrient gradients should be focused sharply on the chemical and biological characteristics of the humus layer.

Thirty-two different species from the six stands were analyzed for thirteen different elements. Samples were collected during the third week of July and third week of August. Four ubiquitous species only were sampled during July. The results for the foliar analysis may be summarized as follows:

1. No statistically significant differences between sites were found in the nutrient concentration of a given species, except possibly for nitrogen. In this case the plants growing in red pine-fir stand have a higher foliar concentration of nitrogen than plants of the same species at either end of the nutrient gradient. Probably the relative importance of nitrogen as a limiting factor in the growth of herbaceous plants differs between stands.

2. Between ecological groups on different sites, there are significant differences in the foliar concentration of nitrogen, phosphorus and potassium. This may be due to differing physiologic requirement, luxury uptake or deficiency. The difference decreases in the order  $K > P > N$ .

3. Although the K : Ca plus Mg ratio in the humus samples decreased in the hardwood stands, a similar trend was not observed in the foliage samples.

4. Consistent changes in foliar nutrient concentration over time were not observed, except for calcium, which consistently increased.

Triangular coordinates provide a useful tool for interpreting data and illustrating relationships. The relative values generated by the transformation, however, appear to be strongly influenced by the variance of each element and the degree of correlation between the three elements. Until this relationship is evaluated rigorously, any inferences based on triangular models should be supported by unadjusted or absolute values.

Many attempts to find significant differences in foliar nutrient concentration within species between sites have, like this one, either failed or yielded ambiguous results. Rather than abandoning all attempts to find significant differences, it might be useful to combine data on nutrient concentration with information on species productivity or biomass per unit area. The sampling should be designed so that nutrient uptake for a given species can be expressed on a per leaf, per plant or unit area basis. The latter is perhaps the most difficult, since the biomass per unit area for a given species is highly variable within most stands. In any case the number of replications within stands should be greatly increased; at least one sample should be collected on each subplot within each stand.

Given the multiplicity of variables that affect nutrient uptake, however, it would be better to concentrate attention on nutrient uptake by plants grown under controlled conditions, at least until more is known about the physiological requirements of the species being considered.

An explanation for the wide differences in nutrient concentration between ecological groups could be sought by growing these groups of species under controlled conditions in the associated surface soil horizon. Fertilizer trials, associated with productivity and foliar analysis, data might suggest which nutrients are limiting to which ecological groups. An alternative method might be to grow each ecological group on different surface soil horizons. For example, understory plants associated with jack pine might be grown on soil from jack pine, red pine and hardwood stands. The results of such a study might indicate to what degree the species distribution is determined by physiologic requirements rather than ecological requirements.

This study has failed to answer many fundamental questions concerning ecological nutrient gradients in Minnesota. As an exploratory study, however, it has raised new questions and suggested worthwhile areas for future inquiry, and it has revealed some of the advantages and pitfalls of applying foliar analysis of subordinate forest plants to ecological situations. A more detailed evaluation of ecological nutrient gradients must await the results of studies of the humus layer, fertilizer trials and greenhouse experiments.

APPENDIX

## INTEGRATIVE LIGHT MEASUREMENTS

In order to provide quantitative data for future studies, light measurements were taken on all subplots. Although these measurements are somewhat incidental to the main purpose of the study, they are relevant in that light is an important variable in the growth and mineral uptake of plants.

Oxalid paper booklets were made according to the method described by Friend (1961). Four masked booklets were taped on each of 48 four-foot laths. On August 18, the laths with the oxalid papers attached were set out and the booklets were unmasked. This was done by centering a lath on top of each stake and nailing it in a horizontal position about 24 inches off the ground. All laths were oriented in a north-south direction to avoid bias. Control papers were set out at three location in the open.

The sun rose at 6:30 A.M.; the last papers were set out before there were many sunflecks on the forest floor. The day was mostly sunny, with no more than two hours of cloudiness. Following collection, the papers were developed over ammonium hydroxide.

According to Friend (1963), the number of papers exposed is a logarithmic function of the total foot-candle minutes. Thus the sample papers could be calibrated by comparison with standards exposed for a known length of time to a known light intensity.

Some of the papers could not be collected until late at night and were therefore discolored by moisture. Within the



stands there was no appreciable dew; apparently exposure to 100 percent relative humidity is enough to discolor the papers. For this reason, no attempt was made to subdivide the interval between the last two papers. This results in a considerable loss of accuracy, particularly at the higher light intensities. This loss is tolerable due to the broad range of light intensities involved in the study.

The light measurements with ozalid paper yielded useful information on the environmental light gradient in the stands studied. Figure 5 shows graphically the relationship between the synecological light coordinate for each plot and the percentage of full sunlight received at the herb level in one day.

If only the hardwood stands and jack pine stands are considered, there appears to be a fairly neat curvilinear relationship between percentage of full sun and synecological light coordinate. When the red pine stands are considered alone, however, the relationship is poor. There are several reasons for this. For one, these stands have a discontinuous second story of balsam fir. Some plots fell beneath a dense fir canopy and others fell in relatively open spots. Second, plots in dense shade frequently have very few species. In these cases the synecological coordinate probably does not mean much. Plot 6 at Mary Lake, for example, had only eight species and hence may be discarded. Third, blow-downs are frequent in the fir and hence the herb flora on some plots may be out of equilibrium with light intensity. In the Wilderness stand, plot 3 occurred near the

north edge of a 30 year-old clear-cut strip and received much more light than other plots in the stand. Hence this plot may also be discarded.

Figure 5 also shows the average percent of full sunlight against synecological light coordinate. In compiling the averages, plot 6 at Mary Lake and plot 3 in the Wilderness stand were omitted. Averages are designated by the larger symbols in Figure 5.

A linear correlation analysis was run using percent of full sunlight and synecological light coordinate computed on both a presence and dominance basis. The correlation coefficients obtained were  $+0.86$  on a presence basis but only  $+0.66$  on a dominance basis. The two plots mentioned previously were also omitted from the calculation.

Future work on light gradients should involve measurement of light both above and below the shrub layer. Measurements should be taken on sunny and cloudy days and during both spring and summer. Synecological light coordinate should be computed separately for the tree, shrub and herb layers.

Given the low light intensity in the two hardwood stands, it seems likely that growth of the ground flora, particularly the ubiquitous, was limited by light intensity. If this were the case, then the foliar concentration of nutrients in the plants sampled should be increased. Without more replication and data on dry-matter weights and leaf size, however, detailed conclusion on the importance of light in mineral uptake are impossible.

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