

Purification, crystallization and structure determination of an azurin variant

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Melanie Ann Ladd

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Dedication

This thesis is dedicated to my Father in heaven who made this beautiful earth. Thank You for blessing this time in my life and giving me the opportunity to grow closer to You by learning more about Your creation.

Abstract

X-ray diffraction is a method that allows the three-dimensional structure of a molecule to be determined. To use this technique to study a protein model, high-quality crystals were grown. A biosynthetic approach was taken to model the mammalian protein peptidylglycine α -hydroxylating monooxygenase (PHM), which is a copper-binding protein that hydroxylates the α -carbon of a glycine residue in the production of peptide hormones. In order to understand the mechanism of this reaction, a model of the two copper sites involved in hydroxylation was created using the bacterial protein azurin as a scaffold (Az-PHM). To compare the structural similarity of the model to the native PHM system, Az-PHM crystals were grown for x-ray diffraction using various buffers, salts, polyethylene glycol (PEG) and excess copper. Dozens of the resulting crystals were diffracted, which had lower resolutions (~ 2.5 Å) and higher mosaicities ($0.8 - 1.2^\circ$ on average). Crystal dehydration and cryoprotection techniques were applied and consistently yielded higher resolution and lower mosaicity crystals. The crystal with the highest resolution and low mosaicity was grown in Tris buffer, lithium nitrate, PEG-2000 and copper chloride. Diffraction images for this crystal were collected on a Rigaku RAPID II X-ray Diffractometer using a copper radiation source with capillary optics and an R-Axis image plate detector. Data were indexed to yield a $P2_12_12_1$ space group, which was then followed by integration, scaling and averaging using CrystalClear 2.1 software. Phases were determined using the Molecular Replacement method in the software CCP4. Finally, structural refinement of the model and electron density map in Coot yielded a 1.3 Å structure with an R_{factor} of 17.57% and an R_{free} of 20.70%.

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CHAPTER 1

Chapter 1.1 Introduction

Biological inorganic chemistry is an exciting new field of chemistry that is devoted to exploring the role of metals in biological systems. Many metals such as vanadium, manganese, iron, cobalt, nickel, copper, zinc, molybdenum and tungsten have been found to play essential roles in the structure and function of metalloenzymes (1). After iron and zinc, copper is the third most abundant trace element in humans (2) and has many diverse functions in metalloenzymes as it can be involved in electron transfer, oxygen transport and even catalysis (1). With so many functional roles, surely the coordination around the copper plays a large part in fine-tuning its electronic structure and therefore, its function.

Copper sites in metalloproteins can be divided into six classes based on spectroscopic properties which depend on a copper's active site geometry, the types of ligands coordinated to the copper ion, the electronic structure of the copper atom and its resulting function. The first class is traditionally known as Type 1 (T1) and is characterized by proteins that bind a copper(II) ion that give an unusually strong signal in the UV-visible region near 600 nm (3). The signal is the direct result of the copper's strong covalent-like interaction with sulfur in a cysteine (4). The cysteine sulfur is one of three donors to bind in a trigonal fashion; the other ligands include two nitrogen atoms (Figure 1.1a). In addition, sometimes one or two axial ligands will also coordinate the copper center (sulfur from a methionine amino acid and/or oxygen from a backbone carbonyl group). Proteins containing a T1 site are often referred to as "blue copper proteins" and function in electron transfer (1). Type 2 (T2) is the name that defines the second class of copper sites which is expected to show a weaker blue color with absorption between ~575 and 800 nm when a copper(II) ion is bound to the active site. This weak absorption signal is due to the different geometry and ligand types normally found in T2 copper proteins (5). Typically, the ligands surrounding a T2 copper active site include four donors ("x" number of nitrogen atoms and "4-x" oxygen atoms from various amino acids) in a square planar coordination (Figure 1.1b) and on occasion, an axial donor (also usually a nitrogen or oxygen atom). Copper active sites classified as T2

are often involved in catalysis (1). Other classes of copper active sites are more rare and include Type 3 (T3), Copper A (Cu_A), Copper Z (Cu_Z) and copper chaperones.

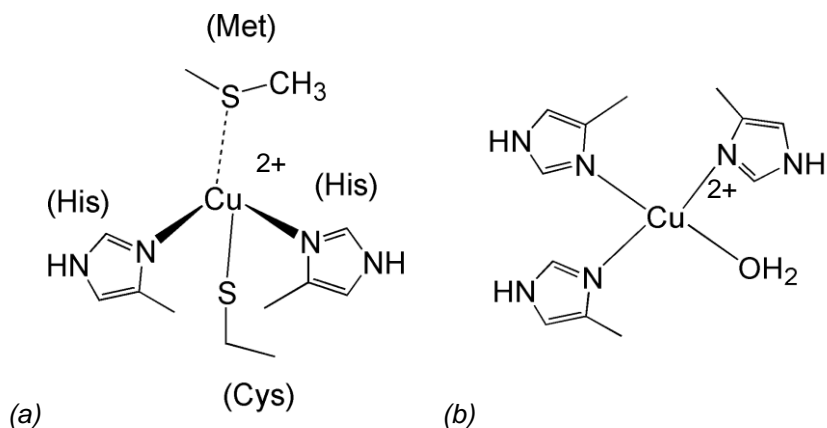


Figure 1.1. Typical coordination of (a) T1 and (b) T2 copper sites in biological molecules.

Understanding why the structure of different copper active sites gives rise to their function is at the heart of this research. Metalloprotein design is one way in which this relationship can be studied. In this study, metalloprotein design involves modeling a naturally-occurring metal site using a biosynthetic approach. Active site residues from the target metal site of interest are mutated into a native protein scaffold that is stable, easy to purify and well-characterized. By using an existing protein as a scaffold, the aqueous environment and common pH of the target system can be preserved. Also, because the number of possible active site residues is limited and their composition is the same as those ligands in the native system, certain aspects of metal coordination are maintained. In modeling the metal site of a target protein in this way, the end goal is to create a model that is a structural and functional mimic of the native system. Generally, native biological enzymes are so efficient and selective that few models can compare. In the future, the ability to mimic the structure and function of any enzyme by way of metalloprotein design will allow researchers to create protein molecules for any desired function (6).

Chapter 1.2 PHM and Azurin

The target protein of interest is peptidylglycine α -hydroxylating monooxygenase (PHM) which is a large metalloenzyme found in mammals. PHM is ~300 residues in

length (7) and is part of a larger protein complex called peptidylglycine α -amidating monooxygenase (PAM) which was first located in the pituitary gland (8). PHM contains two copper-binding sites that function in peptide hormone synthesis, catalyzing the oxidation of C-terminal glycine-extended peptides into α -hydroxylated products (9). These two copper sites are located across a solvent gap of ~ 11 Å (7) and each have different roles in that one of the coppers functions in electron transfer/electron storage (Cu_H) and the other participates in catalysis (Cu_M). The Cu_H site is thought to get its electrons from a physiological reductant, such as ascorbate, while the Cu_M site is thought to be where molecular oxygen and substrate bind prior to the hydroxylation reaction. One of the unknown questions about the roles of the copper ions in the mechanism is the nature of the reactive oxygen species generated at the Cu_M site (9).

Though the functions of these copper ions are characteristic of T1 and T2 copper active sites, structurally-speaking, both of these copper sites resemble the structure of T2 sites (Figure 1.2). Cu_H is bound by three histidines, 107, 108 & 172, and Cu_M is bound by two histidines, 242 & 244, and methionine 314 (the numbering is from human PHM) (9).

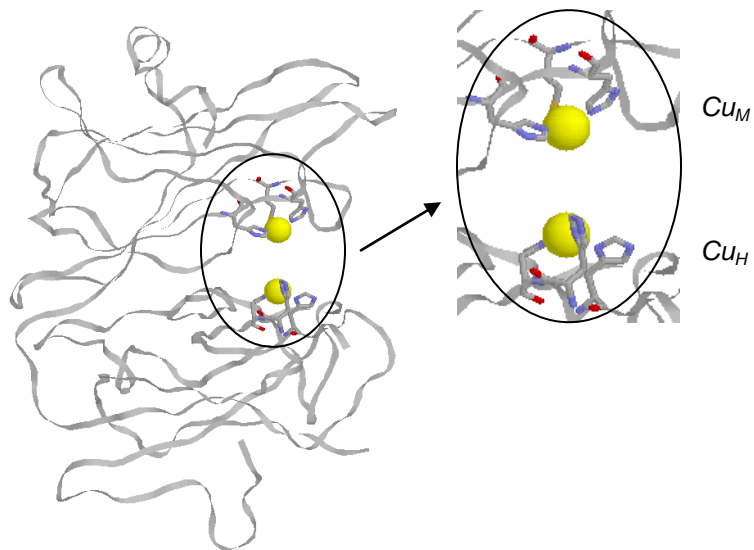


Figure 1.2. Copper active sites of PHM (PDB ID: 1PHM). The copper on the top, Cu_M , is bound by two histidine residues and a methionine and functions in electron transfer. The copper on the bottom, Cu_H , is bound by three histidines and is the site for catalysis.

Interestingly, PHM's copper(II) ions communicate when an electron is passed from Cu_H to Cu_M. This is puzzling given that the two copper ions are located across a solvent gap and are not coupled to one another. The shortest distance for the electron to travel is directly between the two copper centers in the solvent-filled gap that separates the copper ions (9). In order to better understand how the mechanism for electron transport occurs, to clarify the nature of the reactive oxygen species and to explore copper metalloprotein design parameters, a biosynthetic model was created using a protein called Azurin as the scaffold.

Azurin is small protein with a molecular weight of 14 kDa and a length of 128 residues (10). It is found in the soil bacteria *Pseudomonas aeruginosa* and is thought to play a role in the shuttling of electrons in the denitrification pathway (11, 12). It is a well-characterized blue copper protein, and historically, it's stable to mutations (13, 14), making it an ideal candidate to use for design. Azurin was also chosen because it possesses a native Type 1 copper binding site (His46, Cys112, His117 & Met121), shown in Figure 1.3. Using azurin as a scaffold, a Type 2 site was designed where three residues were mutated, namely Gln8Met, Gln14His and Asn16His, to create an Az-PHM variant (15). The overall protein fold of azurin contains mostly beta-sheet secondary structures with a looped section containing some alpha helical segments. Most of the amino acids supporting the T1 copper site are found on this loop section (Cys112, His117, Met 121) with the fourth ligand found on an internal strand of beta-sheet (His46) (16). The section of the protein where the T2 copper binding site was designed is a section that has three parallel strands of beta-sheet structure, with the amino acid residues Gln8Met, Gln14His, and Asn16His being found on two of these adjacent strands.

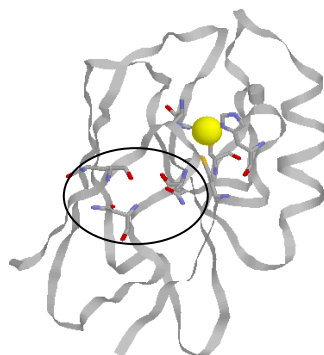


Figure 1.3. Azurin crystal structure (PDB ID: 4AZU) with a copper bound in the T1 copper site. The three residues circled were the amino acids that would be mutated to bind copper in the designed T2 site.

In order to study the function of the Az-PHM model, structural analysis must be carried out to identify whether or not the design of the T2 site allows a copper(II) ion to bind and to determine how closely the model recreates the native copper binding site structure. Protein structure can be determined qualitatively in several ways, however x-ray crystallography and NMR spectroscopy are the only two ways in which a high-resolution, three dimensional structure can be determined (17). In light of this, research began by purifying the protein in order to grow crystals of the mutant. Obtaining pure Az-PHM is vital because growing diffraction-quality crystals depends on the purity of the protein sample (18).

Chapter 1.3 Az-PHM Purification

Az-PHM was purified from BL21* *E. coli* cells previously infected with a pET9a expression vector containing a gene for the azurin preprotein from the bacterium *Pseudomonas aeruginosa* (15). Cell colonies were cultured for 5-8 hours in LB media containing Kanamycin in a shaking incubator at 37°C and then again overnight in a 2x yeast-tryptone (2xYT) medium containing 16 g bactotryptone, 10 g yeast extract, 5 g sodium chloride and 50 mg kanamycin per liter of media for 12-14 hours at 30°C. After cells reached an optical density of >1.5 at 600 nm, the cultures were induced with isopropyl β -D-1-thiogalactopyranoside (IPTG) to a concentration of ~0.07 g/L of culture and then incubated for another 3 hours. The induction with IPTG caused the azurin gene to be aggressively translated and transcribed from the pET9a plasmid.

Cells were pelleted via centrifugation and then lysed through osmotic shock. The first step in shocking the cells was suspension of the cell pellet at 37°C in a sucrose solution consisting of 20% (w/v) sucrose, 1 mM EDTA and 39 mM Tris. After subsequent centrifugation, the resulting pellets were resuspended in purified H₂O at 4°C for ~10 minutes causing the cells to swell, breaking the cell wall. Because the azurin preprotein gene includes code for a 19 amino acid leader sequence that sends the finished protein to the periplasm, Az-PHM is released into solution after cell wall lysis leaving behind intact plasma membranes. A third round of centrifugation follows osmotic shock to separate Az-PHM from the cell lysate.

In order to further purify Az-PHM from other proteins in the periplasmic supernatant, sodium acetate at pH 4.1 was added to a final concentration of ~50 mM to precipitate the unwanted protein. After a final round of centrifugation, Az-PHM's isoelectric point of ~5.1 was utilized in cation-exchange chromatography. SP-Sepharose resin, which consists of negatively-charged sulfonate groups, was added to the supernatant. Because a pH of 4.1 was maintained during this step, the sulfonate groups on the resin bind the positively-charged Az-PHM. The resin was washed in the SP-column before increasing the pH to 6.35, a pH above its isoelectric point, to elute Az-PHM. Next, the protein was added to an anion-exchange column containing Q-Sepharose resin equilibrated to a pH of 6.35 with 50 mM ammonium acetate. At this pH, other proteins in the solution bind to the positively-charged amino groups of the resin, allowing Az-PHM to flow through in a fairly pure form. Lastly, a size-exclusion column containing Sephadex resin separated any proteins in solution by their size. Before running the column, the Az-PHM in solution was titrated with copper.

Throughout the purification, the presence and concentration of Az-PHM was determined using the unique spectroscopic properties of azurin's T1 copper site. When copper is titrated into a sample of apo azurin, each azurin active site binds a copper(II) ion tightly and electrons from the copper are covalently shared with the sulfur of Cys112. This bonding characteristic gives rise to a very intense S→Cu(II) charge transfer band at ~625 nm (3). To calculate the concentration of Az-PHM, a UV-visible spectrum of the apo form of the protein is obtained and overall protein concentration is determined using

Beer's Law, absorbance at ~ 280 nm and a molar absorptivity of $8440 \text{ M}^{-1}\text{cm}^{-1}$. One equivalent of copper is titrated into the measured sample and a spectrum of holo Az-PHM was obtained. Knowing that azurin's molar absorptivity is $5000 \text{ M}^{-1}\text{cm}^{-1}$ at ~ 625 nm, Az-PHM's concentration was determined by measuring the absorbance of the signature peak near 625 nm (15).

This spectroscopic characteristic was also used to assess the purity of the fractions from the size-exclusion column. A pure sample of Az-PHM should ideally produce a spectrum where the ratio of absorbance at 280 nm to the absorbance at 625 nm is equal to 1.8. This ratio was calculated for each fraction that eluted from the size-exclusion column. Typically, fractions ~ 40 to 43 gave ratios close to 1.8 and were noticeably smaller than the ratios of fractions ~ 20 -39. This trend is visible in Figure 1.4 where fractional ratios for the first two growths of Az-PHM are shown.

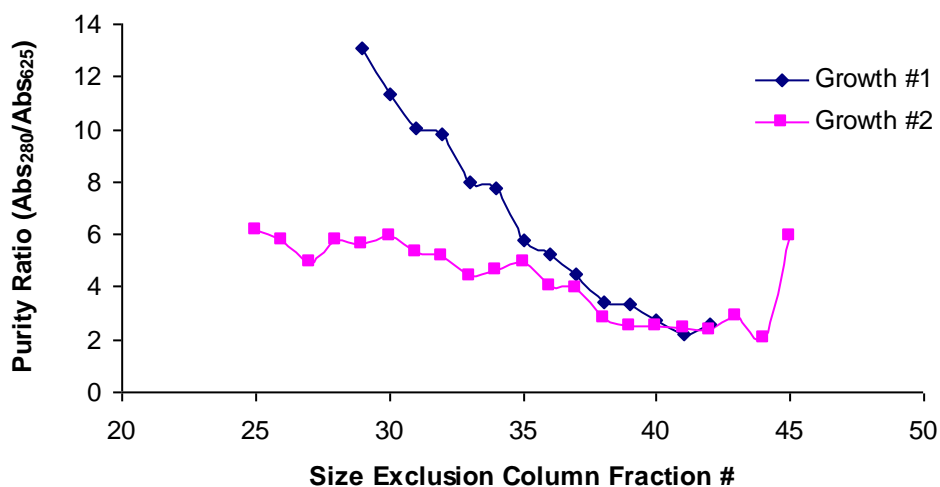


Figure 1.4. Graph showing the purity ratios of fractions containing Az-PHM from the size-exclusion column. A ratio of 1.8 indicates a pure Az-PHM sample.

Protein purity was also examined using sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE). Small samples of Az-PHM taken before and after the size exclusion column were run down an SDS gel and analyzed for contamination by other proteins. Figure 1.5 shows a gel containing samples from two purifications. Lane 1 contains a protein ladder. Lanes 2 and 3 contain samples before running the gel-column.

Lanes 4 and 5 contain samples from fractions between ~25-39 while protein from fractions ~40-43 is found in lanes 6 and 7.

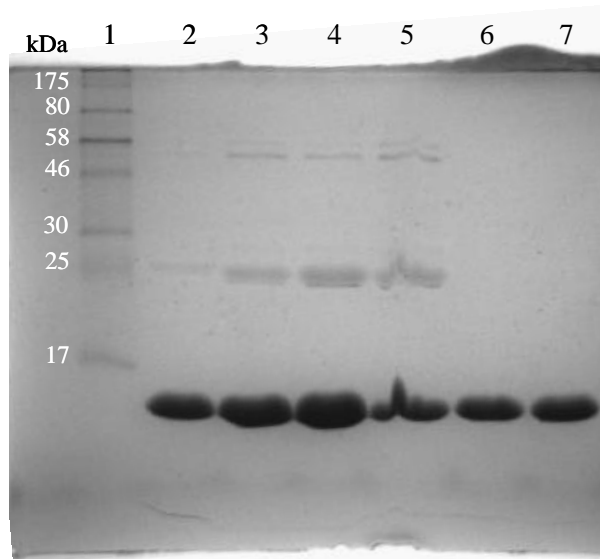


Figure 1.5. Results of SDS-PAGE using Az-PHM samples taken before and after the size-exclusion column. Lane 1 contains a protein ladder. Lanes 2 and 3 contain samples before running the gel-column for growths 1 and 2, respectively. Lanes 4 and 5 contain samples from fractions between ~25-39 while protein from fractions ~40-43 is found in lanes 6 and 7 from growths 1 and 2, respectively.

In analyzing the spectroscopic ratios and the gel from SDS-PAGE, it is apparent that the size exclusion column was successful in purifying Az-PHM from other contaminants. In analyzing the spectroscopic data, the purest protein was typically found in fractions 40-43. Though the ratios for these fractions were not 1.8, they were very close to this ideal value and suggested a nearly pure sample. Their purity was confirmed with SDS-PAGE when the resulting gel revealed single large bands which appear to be under 17 kDa in lanes 6 and 7. The extra bands present in lanes 2-5 also explain the high spectroscopic ratios calculated for fractions ~25-39. These bands near 25 kDa and 50 kDa are clearly not azurin and therefore, serve to increase the spectroscopic ratios. The ultrapure Az-PHM samples from fractions ~40-43 are ideal for crystallography and greatly increase the chances that crystals will form (18).

CHAPTER 2

Chapter 2.1 Introduction

In order to gain structural information from the Az-PHM model system, ultrapure protein samples were used to grow crystals for x-ray diffraction. Most crystallographers would agree that growing diffraction-quality protein crystals is the most time-intensive and least understood part of the crystallography process. Diffraction-quality crystals are well-ordered gelatinous solids in which protein molecules stack together in a repeating pattern. Part of the difficulty in obtaining ordered crystals is due to the irregular shape of most macromolecules. Because protein molecules lack flat edges, they don't stack well, compelling water molecules to form channels between the layers of stacked protein molecules. Water molecules facilitate interactions between protein molecules, which help the crystal nucleate and grow, however, hydrogen bonding interactions are weak which means that protein crystals are fragile and fall apart easily. Another factor that complicates stacking is the inherent chemical properties of protein molecules. Small changes in the pH of the solution cause differences in charge over the surface of the protein molecule, making it difficult to predict conditions that will cause crystallization (17).

Generally, crystallization occurs when pure, concentrated protein is placed in an environment that causes it to become supersaturated and precipitate out of solution. Supersaturation can be achieved through two different methods that are often used simultaneously, namely water loss and the use of a precipitating agent. Water loss, typically via evaporation, causes the protein in solution to become more concentrated by forcing it into a smaller volume of solution. A precipitant does a similar thing by immobilizing water, causing the protein to aggregate together in a smaller volume of solution. Most often the precipitating agent is a salt or polyethylene glycol that is either diffused in or added directly to the buffered protein solution to compete for solvation and salt out the protein. It can, however, be beneficial to use both salt and polyethylene glycol (PEG). Depending on its concentration and the makeup of the protein, salt can either salt out or salt in. Salting in describes the process of allowing the protein to more easily dissolve by raising the ionic strength of the solution. Salt can also facilitate

protein-protein interactions by shielding repulsive charges on the surface of the protein molecules (18).

After finding the exact salt and PEG concentrations coupled with the right pH and temperature that will yield crystals, two techniques called dehydration and cryoprotection can be implemented depending on the quality of the crystals and the need to protect them from ice crystallization during pinning and cryofreezing. Dehydration is a process that involves removing water molecules from a crystal that has already formed. Water is removed when crystals are equilibrated to a solution containing less water than the solution the crystals grew in. By forcing water out of the crystal in this manner, the crystal becomes more ordered. Cryoprotection, on the other hand, decreases ice formation around the crystal by providing a viscous solution to pin crystals from. Crystals are successively equilibrated against solutions of increasing glycerol. The glycerol prevents water from forming crystalline ice which would diffract with the protein crystal and cause unwanted diffraction rings (“ice rings”) to appear in the diffraction pattern. The need for dehydration and cryoprotection is initially determined after screening crystals.

It is necessary to screen crystals to determine if they are well-ordered (diffraction-quality) since it is difficult to visually distinguish a good crystal from a bad crystal. Screening a protein crystal typically involves mounting it on an x-ray diffractometer and exposing it to x-ray radiation to obtain two sample diffraction images. The reflection data from the images are processed, and the resulting statistics are used to assess the quality of the crystal. These statistics include spot intensity, mosaicity, resolution, the prevalence of ice rings and the accuracy of the unit cell prediction. If the crystal is well-ordered then reflections will be intense and have a large intensity relative to background. The crystal’s mosaicity, a measure of its disorder, will be low and resolution will be high, meaning reflections will be located far from the center of the image. In addition to having no ice rings, the high quality of these three statistics helps ensure that the unit cell prediction is accurate.

Chapter 2.2 Method for Crystal Growth & Crystal Box Set-up

Pure protein was used to set up crystal boxes that utilized the Hanging Drop Vapor Diffusion method. Crystallization of the protein using this method relies on the diffusion of water from the protein drop into a reservoir solution below, as depicted in Figure 2.1.

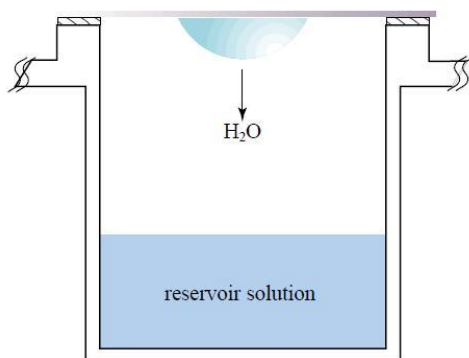


Figure 2.1. Depiction of the hanging drop vapor diffusion method, adapted from reference 19. Protein is located in the drop and becomes concentrated as water leaves the drop.

The hanging drop vapor diffusion method calls for a hanging drop that sits in a sealed environment over a reservoir solution that is typically twice as concentrated with precipitation reagents. Because the drop is in vapor equilibrium with the reservoir solution, water leaves the drop in order to reach equilibrium. As water leaves, the protein located in the drop becomes more concentrated. With the right balance of salts and precipitating agents to facilitate protein-protein interactions, the protein will crystallize out of solution.

To find conditions under which Az-PHM would crystallize, primary literature was consulted (20-31). Numerous conditions were identified where azurin and variants of azurin had previously been crystallized using vapor diffusion (Table 2.1). When crystallizing Azurin on two separate occasions, Faham *et al.* used Tris buffer, lithium nitrate, polyethylene glycol (PEG)-4000 and copper chloride (22). These reagents were initially chosen to crystallize Az-PHM since a crystal structure was desired that showed a copper(II) in the weakly-binding T2 site designed on the surface of the protein.

Table 2.1. Conditions used to crystallize azurin and its variants. All conditions were found in primary literature. Special instructions regarding exact crystallization methods are not included.

Reference	Reservoir Reagents	Drop Reagents	Protein Concentration
20	0.1 M Acetate pH 5.5 3.2 M (NH ₄) ₂ SO ₄ 0.5 M LiNO ₃	0.1 M Acetate pH 5.5 2.0 M (NH ₄) ₂ SO ₄ 0.3 M LiNO ₃	10 mg/mL
21	3.4 M Na/KPO ₄ pH 6.0	1.7 M Na/KPO ₄ pH 6.0	14 mg/mL
22	0.1 M TrisHCl pH 8.0 0.1 M LiNO ₃ 25% PEG-4000 20 mM CuCl ₂	0.05 M TrisHCl pH 8.0 0.05 M LiNO ₃ 12.5% PEG-4000 10 mM CuCl ₂	10 mg/mL
23	0.1 M Tris pH 8.0 0.1 M LiNO ₃ 30% (w/v) PEG-4000 20 mM CuCl ₂	0.05 M Tris pH 8.0 0.05 M LiNO ₃ 15% (w/v) PEG-4000 10 mM CuCl ₂	15 mg/mL
24	0.1 M Imidazole pH 6.0-8.0 0.1 M LiNO ₃ 25-38% PEG-4000/8000 6.25 mM CuCl ₂	0.05 M Imidazole pH 6.0-8.0 0.05 M LiNO ₃ 12.5-18% PEG-4000/8000 3.125 mM CuCl ₂	10-15 mg/mL
25	0.1 M Imidazole pH 8.0 0.1 M LiNO ₃ 20% PEG-8000	0.05 M Imidazole pH 8.0 0.05 M LiNO ₃ 10% PEG-8000	13 mg/mL
26	0.1 M Imidazole pH 7.0 0.1 M LiNO ₃ 20% PEG-4000	0.05 M Imidazole pH 7.0 0.05 M LiNO ₃ 10% PEG-4000	15 mg/mL
27	0.1 M Imidazole pH 7.0 0.1 M LiNO ₃ 20% PEG-4000	0.05 M Imidazole pH 7.0 0.05 M LiNO ₃ 10% PEG-4000	15 mg/mL
28	0.1 M Citric acid pH 3.2 0.1 M LiNO ₃ 20-24% PEG-4000	0.05 M Citric acid pH 3.2 0.05 M LiNO ₃ 10-12% PEG-4000	15 mg/mL
29	0.1 M Imidazole pH 7.0 0.1 M LiNO ₃ 20% PEG-4000	0.05 M Imidazole pH 7.0 0.05 M LiNO ₃ 10% PEG-4000	15 mg/mL
30	0.22 M acetate pH 6.0 0.2 M CaCl ₂ 25% PEG-4000		7.5-10 mg/mL
31	0.08 M Na acetate pH 6.0 0.24 M LiNO ₃ 0.24 M CaCl ₂ 25% PEG-8000 (or 4000)	0.04 M Na acetate pH 6.0 0.12 M LiNO ₃ 0.12 M CaCl ₂ 12.5% PEG-8000 (or 4000)	1 mM

Besides the reagents chosen initially, azurin was also commonly crystallized in buffers such as imidazole and acetate, other salts such as calcium chloride, copper such as copper sulfate and other precipitating agents such as PEG at molecular weights 600, 1000, 2000 and 8000 (Table 2.1). Combinations of all these reagents at various concentrations were explored using 24-well crystal boxes. The 24-well plates have four

rows (labeled A, B, C and D) with six reservoirs in each row (labeled 1, 2, 3, 4, 5 and 6). An example of the conditions used in one of the crystal box trials, called “Box #70”, is presented in Table 2.2. In setting up the box, the buffer, salt, precipitant, copper and water were mixed in each of the 24 reservoirs and final concentrations of these reagents in each reservoir (denoted A1, A2, A3... through D5, D6) are indicated in parentheses in Table 2.2. Then after mixing reservoir solutions, 2 μL was taken from well A1 and mixed with 2 μL of purified Az-PHM on a coverslip. The purified Az-PHM used in crystal box set-up had concentrations between 25-35 mg/mL (or 1.8-2.5 mM). Lastly, the coverslip was sealed over the respective reservoir and the process was repeated for the remaining 23 wells.

Table 2.2. Example of crystal conditions (Box #70). All concentrations in parentheses are final concentrations in the reservoir solution. The drop above each reservoir contained 2 μ L of water, 2 μ L of 29.5 mg/mL Az-PHM in 50 mM imidazole pH 8.07 and 2 μ L of reservoir (for drops over row A, 2 μ L from A3; for drops over row B, 2 μ L from A4; for drops over row C, 2 μ L from A5; for drops over row D, 2 μ L from A6).

	2.0 M Imidazole pH 8.02	5.0 M LiNO₃	50% PEG-8000	0.1 M CuCl₂	Millipore H₂O
A1	25 μ L (0.05 M)	0 μ L	260 μ L (13%)	50 μ L (0.005 M)	665 μ L
A2	25 μ L (0.05 M)	0 μ L	220 μ L (11%)	50 μ L (0.005 M)	705 μ L
A3	25 μ L (0.05 M)	0 μ L	180 μ L (9%)	50 μ L (0.005 M)	745 μ L
A4	25 μ L (0.05 M)	0 μ L	140 μ L (7%)	50 μ L (0.005 M)	785 μ L
A5	25 μ L (0.05 M)	0 μ L	100 μ L (5%)	50 μ L (0.005 M)	825 μ L
A6	25 μ L (0.05 M)	0 μ L	60 μ L (3%)	50 μ L (0.005 M)	865 μ L

	2.0 M Imidazole pH 8.02	5.0 M LiNO₃	50% PEG-8000	0.1 M CuCl₂	Millipore H₂O
B1	25 μ L (0.05 M)	0 μ L	260 μ L (13%)	50 μ L (0.005 M)	665 μ L
B2	25 μ L (0.05 M)	0 μ L	220 μ L (11%)	50 μ L (0.005 M)	705 μ L
B3	25 μ L (0.05 M)	0 μ L	180 μ L (9%)	50 μ L (0.005 M)	745 μ L
B4	25 μ L (0.05 M)	0 μ L	140 μ L (7%)	50 μ L (0.005 M)	785 μ L
B5	25 μ L (0.05 M)	0 μ L	100 μ L (5%)	50 μ L (0.005 M)	825 μ L
B6	25 μ L (0.05 M)	0 μ L	60 μ L (3%)	50 μ L (0.005 M)	865 μ L

	2.0 M Imidazole pH 8.02	5.0 M LiNO₃	50% PEG-8000	0.1 M CuCl₂	Millipore H₂O
C1	25 μ L (0.05 M)	0 μ L	260 μ L (13%)	50 μ L (0.005 M)	665 μ L
C2	25 μ L (0.05 M)	0 μ L	220 μ L (11%)	50 μ L (0.005 M)	705 μ L
C3	25 μ L (0.05 M)	0 μ L	180 μ L (9%)	50 μ L (0.005 M)	745 μ L
C4	25 μ L (0.05 M)	0 μ L	140 μ L (7%)	50 μ L (0.005 M)	785 μ L
C5	25 μ L (0.05 M)	0 μ L	100 μ L (5%)	50 μ L (0.005 M)	825 μ L
C6	25 μ L (0.05 M)	0 μ L	60 μ L (3%)	50 μ L (0.005 M)	865 μ L

	2.0 M Imidazole pH 8.02	5.0 M LiNO₃	50% PEG-8000	0.1 M CuCl₂	Millipore H₂O
D1	25 μ L (0.05 M)	0 μ L	260 μ L (13%)	50 μ L (0.005 M)	665 μ L
D2	25 μ L (0.05 M)	0 μ L	220 μ L (11%)	50 μ L (0.005 M)	705 μ L
D3	25 μ L (0.05 M)	0 μ L	180 μ L (9%)	50 μ L (0.005 M)	745 μ L
D4	25 μ L (0.05 M)	0 μ L	140 μ L (7%)	50 μ L (0.005 M)	785 μ L
D5	25 μ L (0.05 M)	0 μ L	100 μ L (5%)	50 μ L (0.005 M)	825 μ L
D6	25 μ L (0.05 M)	0 μ L	60 μ L (3%)	50 μ L (0.005 M)	865 μ L

Chapter 2.3 Dehydration and Cryoprotection of Crystals

Crystal boxes were stored at room temperature or at 4°C in a walk-in cold room. The boxes were checked daily over the span of days and weeks. Crystals often appeared within 24 hours, with typical growth times requiring 3-7 days. After crystals appeared to stop growing, some crystals were selectively removed from the drops to be dehydrated and cryoprotected following methods adapted from Heras *et al.* (32). To perform the dehydration, crystals were transferred into new reservoir solutions containing the exact conditions they were grown under with two exceptions: 1) the amount of PEG was increased by one-third and 2) glycerol was added to give a final 10% (v/v) reservoir solution. For example, if crystals from well A4 of Box #70 needed to be dehydrated, a dehydration reservoir would be set up with conditions described in Table 2.3. Typically dehydration was carried out at 4°C in which case the original box and the dehydration reservoir would first equilibrate to 4°C for 2 hours. The transfer of crystals was also completed at 4°C by quickly moving them from their original drop to a new coverslip with a 5 µL drop of solution from the dehydration reservoir. The drop was then sealed over the dehydration reservoir to equilibrate for approximately 12 hours at 4°C.

Table 2.3. Dehydration conditions for crystals from Box #70, well A4.

	2.0 M Imidazole pH 8.02	Glycerol	50% PEG-8000	0.1 M CuCl₂	Millipore H₂O
A4	25 µL (0.05 M)	100 µL	186 µL (9.3%)	50 µL (0.005 M)	639 µL

After dehydration, the crystals were cryoprotected in a similar way. For each well of crystals being dehydrated, three reservoirs of cryoprotection solution were set up. Cryoprotection conditions are identical to dehydration conditions with one exception: the amount of total glycerol in each of the reservoirs increased successively from 15 to 20 to 25%. For example, if the dehydrated crystals from well A4 of Box #70 needed cryoprotection, three reservoirs of solution would be set up using the reagents shown in Table 2.4. After setting up the wells, they equilibrated at 4°C for 2 hours.

Table 2.4. Cryoprotection conditions for crystals from Box #70, well A4.

	2.0 M Imidazole pH 8.02	Glycerol	50% PEG-8000	0.1 M CuCl ₂	Millipore H ₂ O
15%	25 μ L (0.05 M)	150 μ L	186 μ L (9.3%)	50 μ L (0.005 M)	589 μ L
20%	25 μ L (0.05 M)	200 μ L	186 μ L (9.3%)	50 μ L (0.005 M)	539 μ L
25%	25 μ L (0.05 M)	250 μ L	186 μ L (9.3%)	50 μ L (0.005 M)	489 μ L

Using the same technique described for dehydration, crystals were first transferred from the dehydration drop into the cryoprotection drop containing 15% glycerol. After equilibrating for only 10 minutes, the crystals were next transferred to the drop containing 20% glycerol. The process was then repeated for the third and final 25% glycerol drop. After a final 10 minutes of equilibration, the crystals were immediately pinned and frozen in liquid nitrogen. Pinning the Az-PHM protein crystals involved looping the crystals with a nylon cryoloop and quickly plunging them into a dewar brimming with liquid nitrogen. The instantaneous freezing process was important because freezing crystals in a dewar that was not filled could result in a more gradual cooling of the crystal as it moves through cool, foggy air. Slower freezing is known to result in more crystalline ice forming on the outside of the crystal, prevent nice glass formation of the cryoprotection solvent and lastly, create non-uniformity within the crystal. After pinning and cryofreezing, crystals were stored in a liquid nitrogen dewar until they were screened for quality.

Chapter 2.4 Method for Screening and Predicting a Unit Cell

The quality of the Az-PHM protein crystals was assessed by obtaining their diffraction images. Crystals were carefully mounted in a Rigaku R-Axis Rapid II Diffractometer and maintained at a temperature of -150°C with a nitrogen generator and helium compressor system. The crystals were exposed to copper K α radiation ($\lambda = 1.541870$) at a voltage of 50 kV and current of 40 mA. These x-rays were transmitted by a flat graphite monochromator to a 0.3 mm collimator with special capillary optics. Two diffraction images were collected 90° apart, with a 0.5° oscillation width and 5-minute exposure times. The resulting images were processed with Rigaku's CrystalClear 2.1 software in the macromolecular processing mode. Using the built-in d*TREK processing

suite, the software determined the crystal's unit cell and used it to predict the location of the reflections on the two images. The steps for processing the reflection data using CrystalClear 2.1 are outlined in the rest of this section.

Before the crystal was exposed to radiation, certain software parameters were set regarding the crystal and the type of x-ray radiation being used. In the Setup window (Figure 2.2) are six tabs: Main, Crystal1, Crystal2, Detector, X-Ray Source and Notes. The parameters that were typically used are shown in the Figures. Under the Main tab (Figure 2.2), values were typically entered for *Temperature*. In the Crystal1 tab (Figure 2.3), *Size (X (mm), Y (mm), Z (mm))*, *Color*, *Mount*, and *Morphology* were recorded. In the Crystal2 tab (Figure 2.4), *Space group* (always Unknown) and the *Molecular formula* of Az-PHM ($C_{610}H_{959}N_{165}O_{191}S_{10}Cu_3$) were entered. (Three copper atoms were entered because Az-PHM was titrated with one copper and two or three additional copper atoms were anticipated to bind to the protein molecule since the crystal was grown in solution containing free copper.) The Detector tab (Figure 2.5) contained information that the instrument automatically records. The X-Ray Source tab (Figure 2.6) required *Optics Type*, *collimation type*, *Voltage (kV)*, *Current (mA)*, *Element* and *Source type* to be specified. Finally, any other details about the crystal or its diffraction were recorded under the Notes section. Many of the parameters entered in the Setup window are used in data processing and also appear in the CIF file generated after the structure is solved.

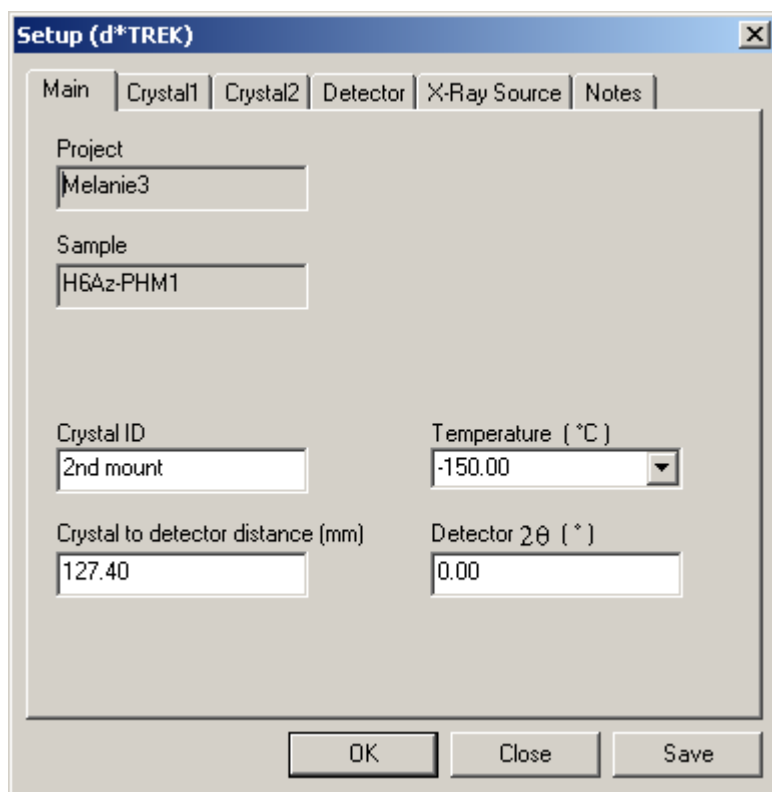


Figure 2.2. d*TREK Setup dialog box in CrystalClear 2.1 showing the Main tab where Temperature and Crystal ID were entered. All other parameters are default.

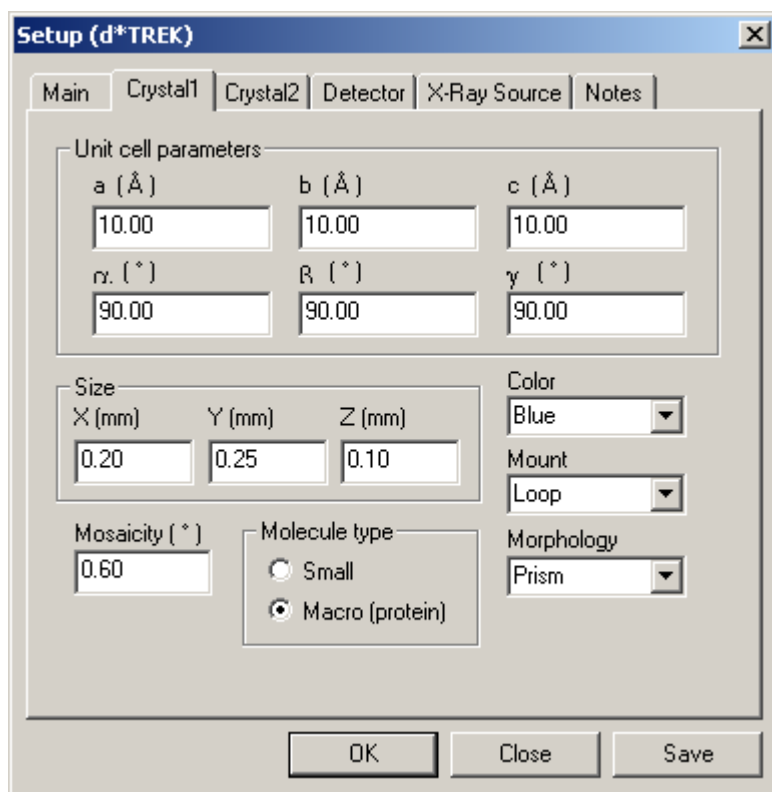


Figure 2.3. d*TREK Setup dialog box in CrystalClear 2.1 showing the Crystal1 tab where Size, Color, Mount and Morphology were entered. All other parameters are default.

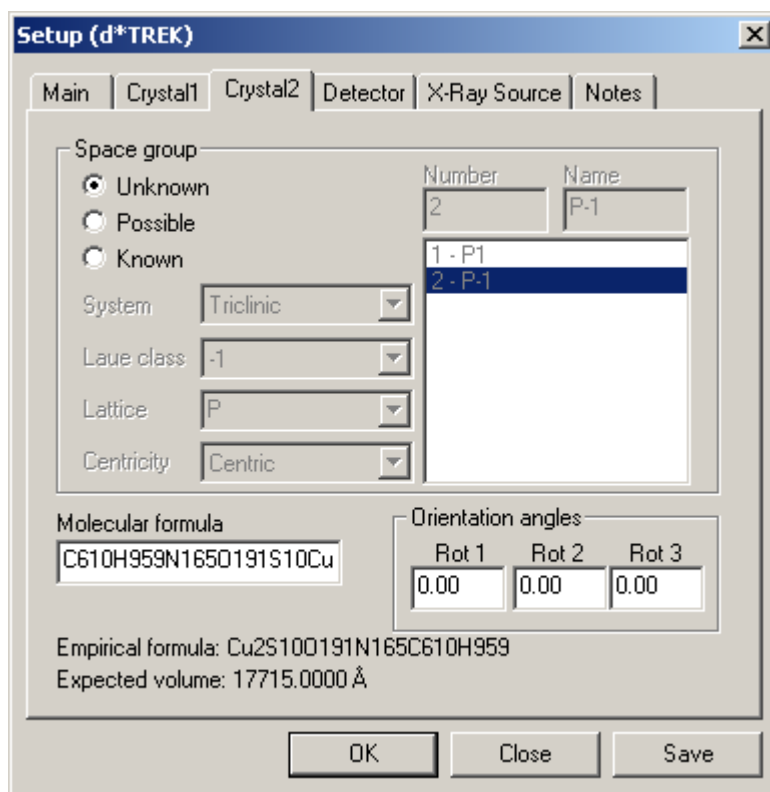


Figure 2.4. d*TREK Setup dialog box in CrystalClear 2.1 showing the Crystal2 tab where Space group and Molecular formula were entered. All other parameters are default.

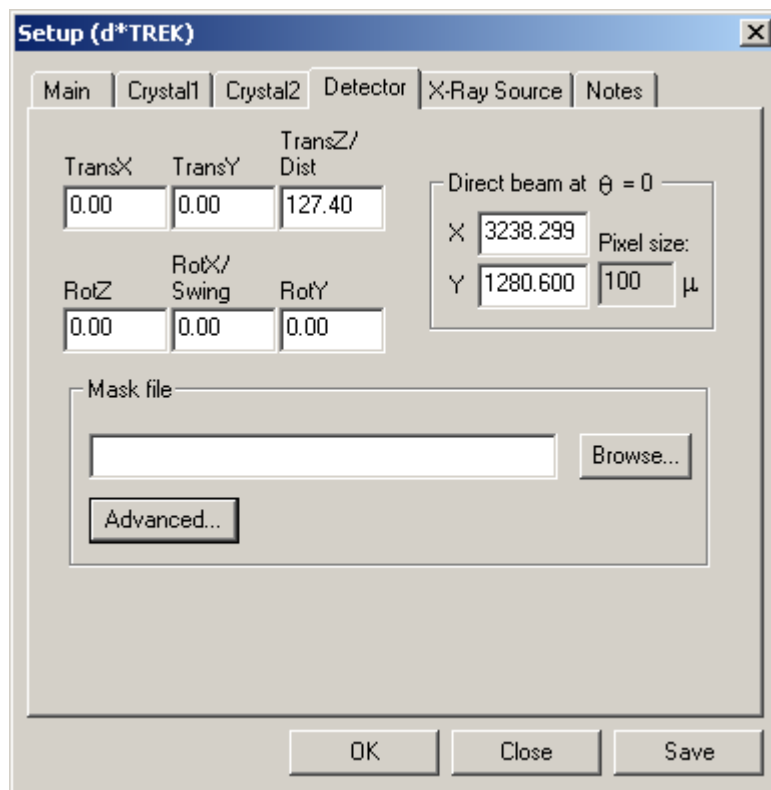


Figure 2.5. d*TREK Setup dialog box in CrystalClear 2.1 showing the Detector tab where no parameters were entered. All parameters are default.

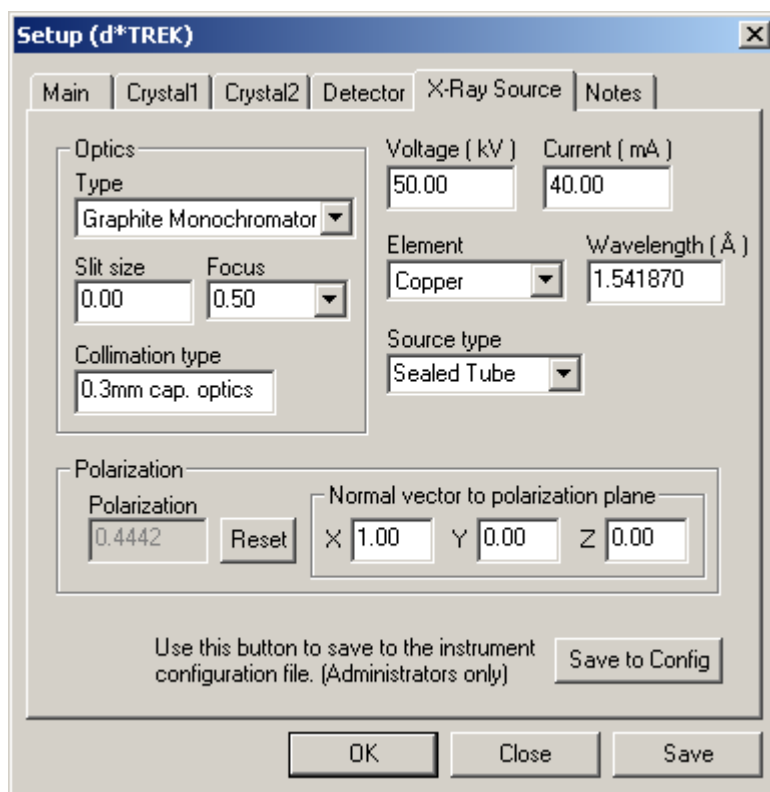


Figure 2.6. *d*TREK Setup dialog box in CrystalClear 2.1 showing the X-Ray Source tab where Optics Type, collimation type, Voltage (kV), Current (mA), Element, and Source type were entered. All other parameters are default.*

Once all Setup parameters were entered and the crystal was centered in the x-ray beam, the crystal was exposed to copper X-ray radiation and two diffraction images were collected. An example of two typical screen images is shown in Figure 2.7. The two image files each contain their own header that records any experimental settings such as beam location, image dimensions, exposure time, and oscillation width.

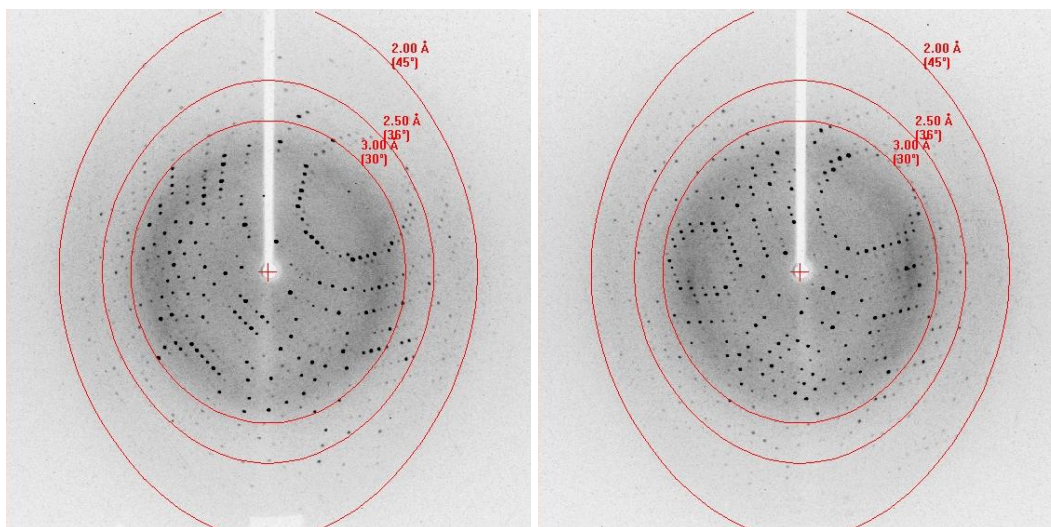


Figure 2.7. Typical screen images that are 0.5° in oscillation width. These images were taken 90° apart.

The first step in determining the unit cell involves locating reflections on the two screened images. All reflection peaks have an associated intensity relative to average background noise. The ratio of intensity to the standard deviation of average background noise (denoted I/Sigma , I/SigI or $I/\sigma I$ in CrystalClear) is one parameter used in finding reflections, seen in Figure 2.8. The default I/Sigma value is 5.00 which was typically used but can be raised or lowered as needed. This value is significant because any reflections having an I/Sigma value below the designated cutoff are not considered spots and therefore, not included in the reflection list that the software passes on to indexing (33). Many protein crystallographers use I/Sigma values of 2 or 3 for clean diffraction images. For images with many spots including twins or noise, it may be best to use I/Sigma values of 10 to be sure that only the strongest spots are used in unit cell determination.

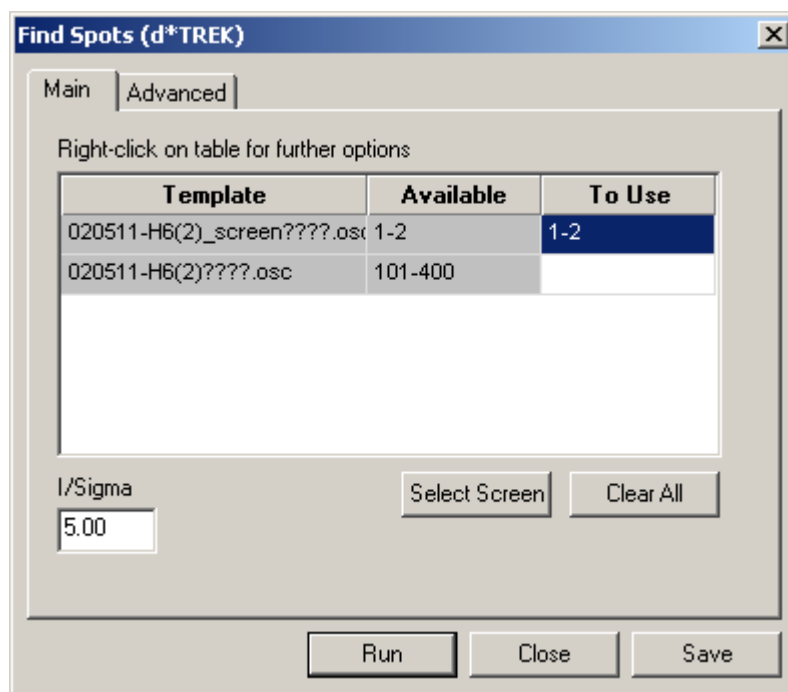


Figure 2.8. The Main tab of the d*TREK Find Spots dialog box in CrystalClear 2.1. The dialog box pictured is an example of typical parameters used when screening a crystal.

Other advanced settings associated with finding spots can also be changed depending on the size and intensity of the reflections produced during screening. These settings shown in Figure 2.9 were always used when finding spots.

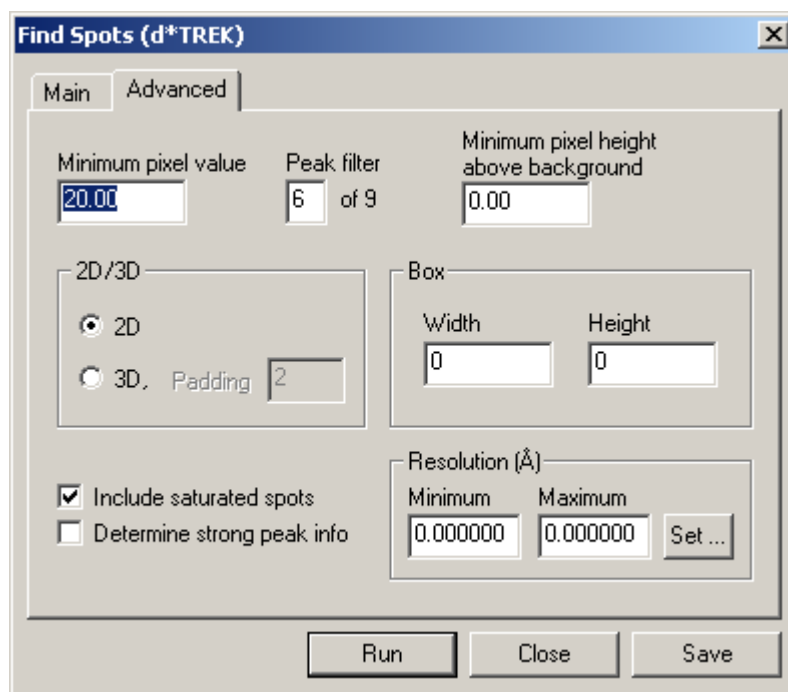


Figure 2.9. The Advanced tab of the d*TREK Find Spots dialog box in CrystalClear 2.1. This dialog box shows default parameters that are typically used when screening a crystal.

Once reflections have been located, a list of spot locations is passed on to the indexing stage where a unit cell, space group, orientation and Bravais lattice type are determined for the crystal being screened. The Index spot function in CrystalClear 2.1, shown in Figure 2.10, was used for this purpose. The *Unknown* option under space group was always selected so that the software was not biased in predicting any particular unit cell. A *Maximum* resolution may be specified which only allows spots out to a certain resolution to be included in indexing. The *User chooses solution* option was always selected because this instructs the software to show a list of possible unit cells after indexing is complete (33).

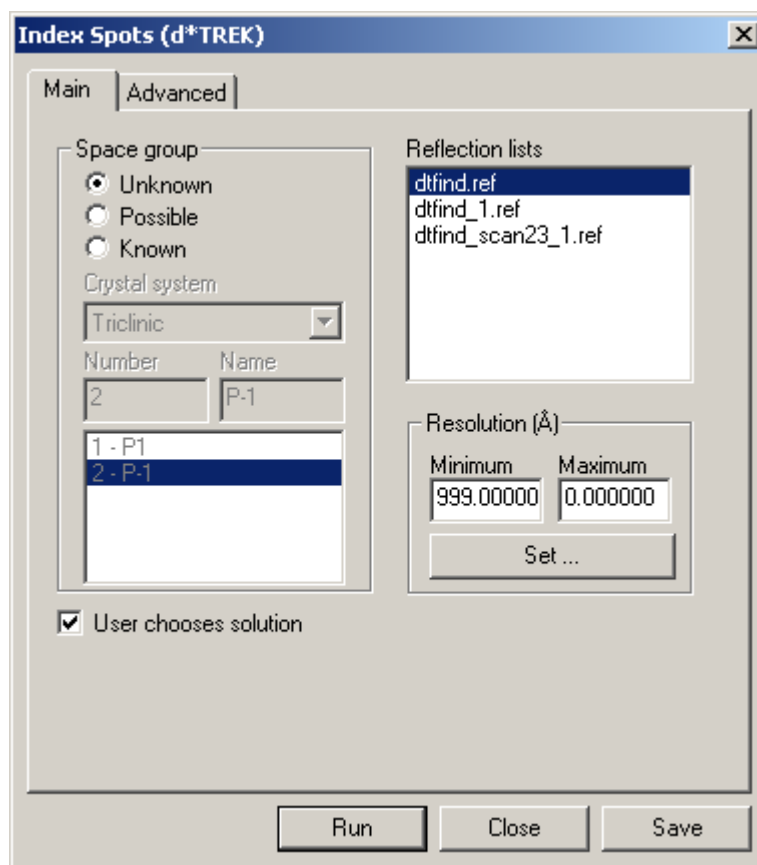


Figure 2.10. The Main tab of the d*TREK Index Spots dialog box in CrystalClear 2.1. Space group, reflection lists, and resolution limits can be specified for indexing.

Parameters on the Advanced tab shown in Figure 2.11 are default and were rarely changed because indexing is especially robust in d*TREK (33). *Max cell length* was not changed to anything other than 0 because doing so tended to limit the number of resulting solutions listed. Decreasing the *Max residual* also resulted in fewer unit cell options to choose from when indexing was complete. By changing *I/SigI*, reflections that do not exceed that cutoff value were excluded from indexing. The *Deice* option was selected so that reflections appearing where ice rings normally show diffraction were not used in indexing. Lastly, it was common for the *Beam check* to remain deselected, but indexing results are not negatively impacted opting to check the beam. When the beam was checked, a radius of 25 was searched for a valid radius of 20. If no better beam location was detected, the software used the location listed in the image headers.

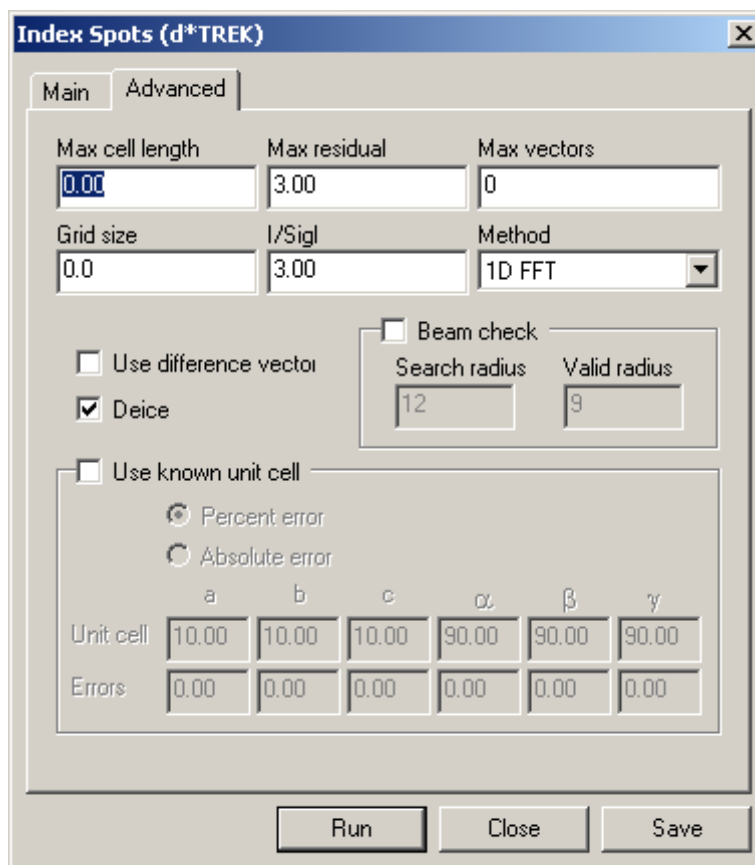


Figure 2.11. The Advanced tab of the d*TREK Index Spots dialog box in CrystalClear 2.1. The default parameters displayed were always used when screening a crystal.

Because the *User chooses solution* option was selected, a window displaying several unit cells opened after indexing was complete, as seen in Figure 2.12. Solutions were listed in order of symmetry, the most symmetrical unit cell appearing first on the list. The software highlighted the most probable solution and orientation. Occasionally, the unit cell that the software deemed the most probable solution was not the correct cell. A unit cell that had a *Least Sq* (least square residual) value of 2.5 or greater was not typically a valid cell (33).

Index Results

Choose a solution:

Soln	Least Sq	Spacegrp	Bravais	Lattice	a	b	c	Volume	α	β	γ
11	0.09	16	orthorh	P	50.17	54.87	85.91	236500	90.00	90.00	90.00
13	0.06	3	monocli	P	54.87	50.17	85.91	236500	90.00	90.04	90.00
14	0.00	1	triclin	P	50.17	54.87	85.91	236500	90.04	90.02	90.04

Orientations

ID	Residual	Rot1	Rot2	Rot3
1	0.0	-117.0	-45.3	88.8
2	0.0	117.0	45.3	-91.2
3	0.0	63.0	-45.3	88.8
4	0.0	-63.0	45.3	-91.2

OK Cancel

Figure 2.12. Sample Index Results where solutions are listed in order of symmetry. The highlighted solution and orientation are results that the software deems most probable.

Once a unit cell solution and orientation matrix were selected, refinement of these values as well as detector and source positions was performed. The refinement reduced differences found between the observed reflection locations and the locations that were calculated using the indexing solutions. The refinement software (Figure 2.13) was always set to *Refine on Images* rather than refining the reflection list generated after finding spots. The same screen images used to find spots (like those in Figure 2.7) were used in refinement. The option to constrain the unit cell according to symmetry was always selected to prevent the software from changing the symmetry of the cell when refining its dimensions. The refinement *Macro Most* was used because it was recommended by Rigaku. This macro has designated parameters such that resolution *Min* and *Max* are set to 0.00, $I/\sigma(I)$ to 5.00, and *Cycles* to 100. It also has designated rejection limits of 0.5 for $X(mm)$ and $Y(mm)$ and 1.0 for $Rot.(deg)$ (33). The *Macro All* can be used as it allows certain rotation and translation values between the detector and source to be refined, however, this refinement is not essential since only two screen images are used (34).

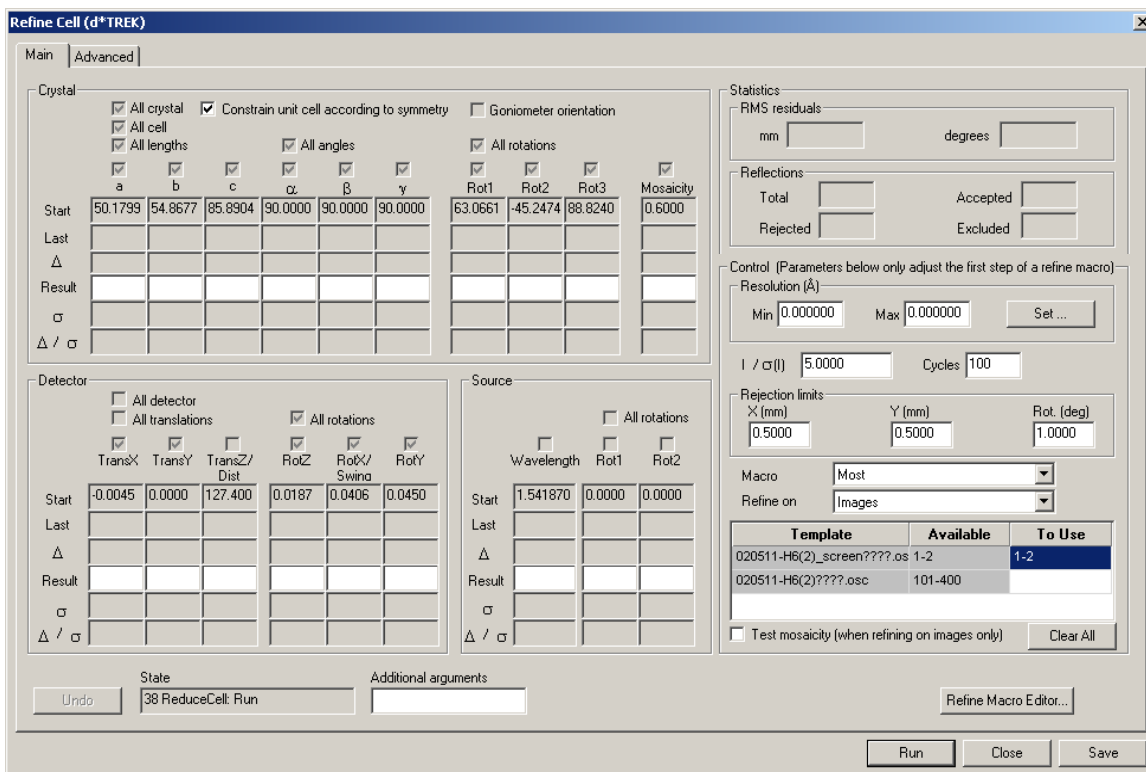


Figure 2.13. The Main tab of the d*TREK Refine Cell window in CrystalClear 2.1. The parameters shown are default with the exception of Refine on Images where both screen images are used.

There are few advanced refinement parameters, none of which were changed from what is shown in Figure 2.14. The *Global refinement* option was not used since it has been disabled in this software version. *Wide slice refinement* is defaulted at none because the screen images taken do not qualify as wide slice since they are smaller than three degrees (33).

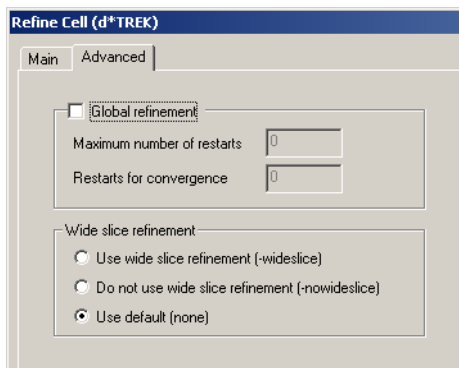


Figure 2.14. Default parameters on the Advanced tab of the d*TREK Refine Cell window in CrystalClear 2.1.

Refinement commenced by hitting *Run*. As multiple rounds of refinement took place, the mosaicity and the ratio of rejected reflections out of total reflections were monitored. When these values did not change appreciably, refinement was considered complete. This typically took three to five runs of refinement. Because the software refined on images, it was possible to graphically see specific reflections that were rejected on the screen images (surrounded by a red circle). With the newly refined unit cell dimensions and positions for the detector and source, calculated reflections could then be predicted and subsequently displayed on the screen images.

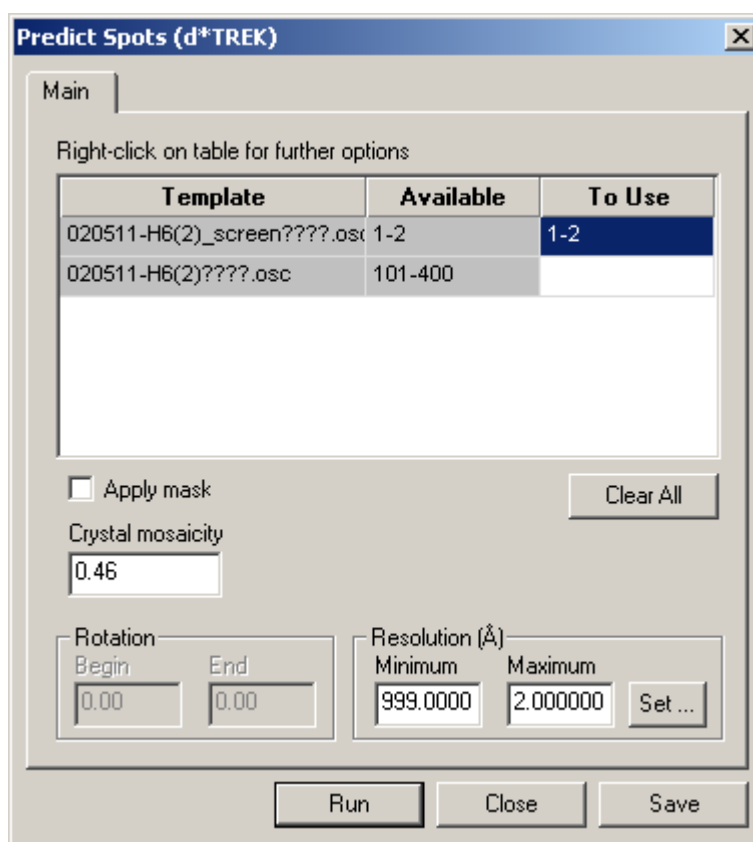


Figure 2.15. The Main tab of the *d*TREK Predict Spots* window in *CrystalClear 2.1*. A Maximum resolution of 2.0 Å was often used for screen images.

The last stage of screening was calculating predicted reflection positions and comparing them to the actual, observed locations. As demonstrated in Figure 2.15, screen images 1-2 were used and the *Crystal mosaicity* value was automatically carried over from refinement. A *Maximum* resolution was usually entered depending on how well the crystal diffracted. A value of 2.00 was routinely used since many crystals that

were screened did not have spots beyond 2.0 Å. If spots were visible beyond 2.0, the resolution maximum was lowered appropriately.

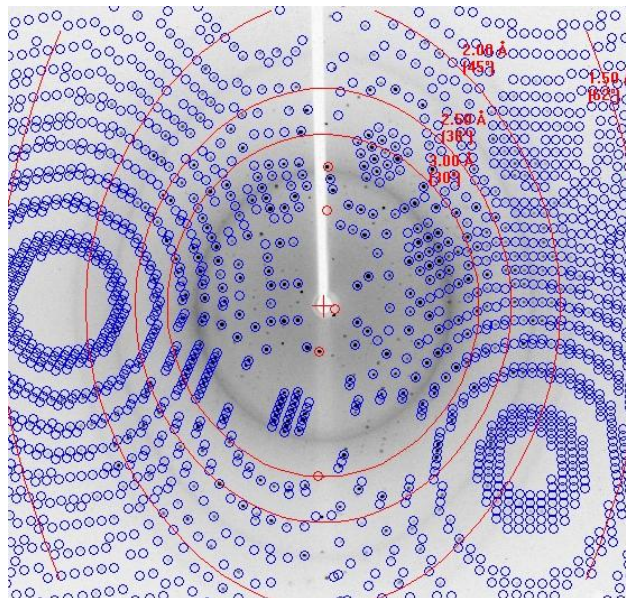


Figure 2.16. Sample prediction results. Mosaicity and refined unit cell parameters are deemed correct due to the accurate predictions for almost all reflections.

Calculated reflections are based on the unit cell and orientation matrix and visually appear on the screen images (Figure 2.16). The images were compared to the calculated reflections (blue circles) to identify the degree to which the software could predict the location of the spots on the images. If most predicted circles were centered on spots, the unit cell assigned to the crystal was deemed correct. (Not all of the spots will be predicted because some are not Bragg reflections.) The prediction also indicated whether or not the CrystalClear software accurately estimated mosaicity. An appropriate mosaicity was estimated when almost all spots were predicted. When only some of the spots were accurately predicted, the mosaicity estimation was low, while a high mosaicity estimate was indicated by many predictions that were observed at locations where no actual spot existed.

Chapter 2.5 Judging Criteria and General Results/Discussion

Predicting a unit cell accurately largely depends on the quality of several factors such as reflection intensity, resolution, crystal mosaicity and to some extent, the lack of

ice rings. These four criteria were the basis for characterizing the diffraction quality of the crystals. High-quality crystals were those that had intense reflections, resolution beyond 2.0 Å, mosaicities less than 0.60 and very minimal to no ice rings. All criteria, with the exception of ice rings, relate to the degree of order inherent in the crystal.

The ratio of reflection intensity relative to background noise is expressed as I/σ and is one criterion that determines the strength of the data. While screening crystals, it was commonly observed that large crystals gave more intense reflections upon diffraction, which is consistent with the findings of Fox & Karplus (35). Theoretically, the intensity of a given reflection is directly related to the number of x-rays that are scattered and converge at the image plate. Because a larger crystal contains more repeating units and takes up more of the beam, more x-rays will be scattered. Thus, reagent conditions were optimized to obtain large crystals for screening in order to avoid collecting weak data.

There are several ways that protein growth can be controlled to yield sizeable crystals, and within a particular buffer system, trends occur that allow for such optimization. The clearest example of reagent optimization is shown in Figure 2.17, where a similar azurin mutant (with only 1 amino acid difference from Az-PHM) was crystallized using two different molecular weights of polyethylene glycol (PEG). There is a distinct difference in size between crystals grown in PEG-4000 (columns 1, 3 and 5) and those grown in PEG-8000 (columns 2, 4 and 6). With the exception of the type of PEG used, drop conditions are identical between columns 1 & 2, between columns 3 & 4 and between columns 5 & 6. It is unmistakable that for this particular buffer system, crystals grown with PEG-8000 are larger than those grown with PEG-4000. The concentration of PEG also affects size. Moving across a pair of columns in a given row, the concentration of PEG decreases which causes a fewer number of large crystals. The number of crystals in a given well is proportional to the size of the crystals. Optimizing conditions that produced large crystals ensured that an incorrect unit cell prediction was not because of weak reflections.

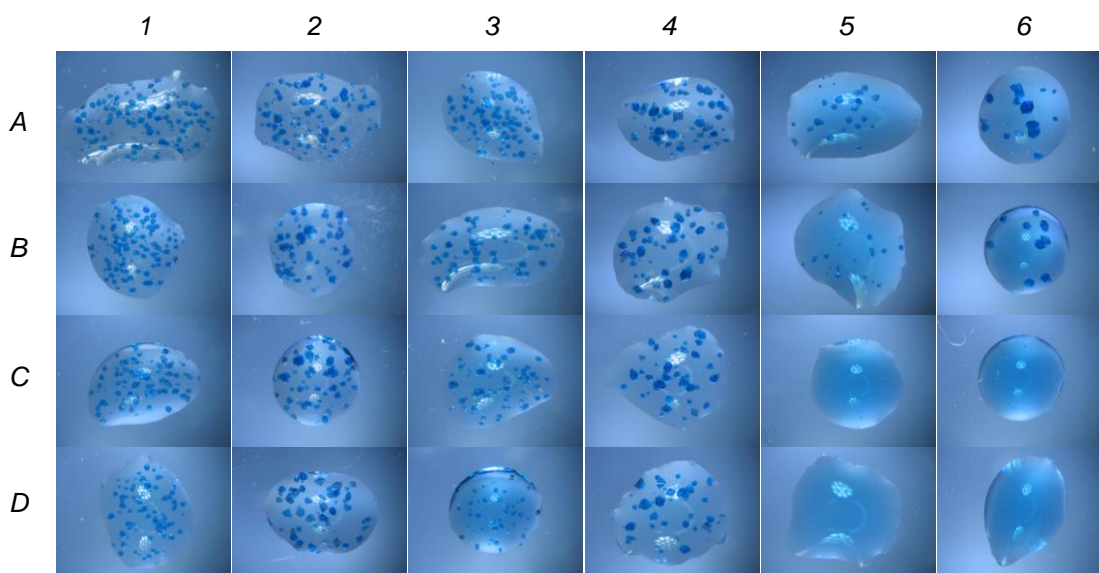


Figure 2.17. Box #58 at ~5 days of an azurin variant with 1 residue difference than Az-PHM. Conditions are identical within column pairs (1 & 2, 3 & 4, and 5 & 6) with the exception of the type of PEG used. PEG-4000 was used in columns 1, 3, and 5 whereas PEG-8000 was used in columns 2, 4 and 6. The concentration of PEG decreases across the row from left to right.

There are two probable reasons for the growth of large crystals in a solution containing a small PEG concentration. When a drop has more water, it takes longer to equilibrate with the reservoir solution below which means that saturation of the drop happens gradually. Thus, fewer nucleation sites result which correlates with the growth of larger crystals. It could also be that because there is less PEG in the drop, there is more mobile water and Az-PHM is not as concentrated. This would also result in less nucleation. In either case, once saturation is reached and nucleation begins, crystals start to grow and the concentration of protein decreases even though the volume of the drop gets smaller (35). This seems plausible because Wilson *et al.* found that as the number of crystals in a given drop decreased, their average size increased (36). Fehrer also found that when crystals grow quickly they tend to be numerous and small (37).

Consistent with these findings by Wilson and Fehrer, imidazole concentration can also be varied to optimize crystal nucleation and size. For this particular buffer, when the concentration of imidazole was decreased, nucleation and crystallization occurred within hours, resulting in many tiny crystals. The photographs in Figure 2.18 from Box #65 were taken after ~24 days. The concentration of imidazole in the drop increases from row A (5 mM) down to Row D (20 mM). With the exception of imidazole, drops within

a given column have identical reagent concentrations. It is clear that the presence of imidazole decreases the number of nucleation sites, resulting in fewer and larger crystals.

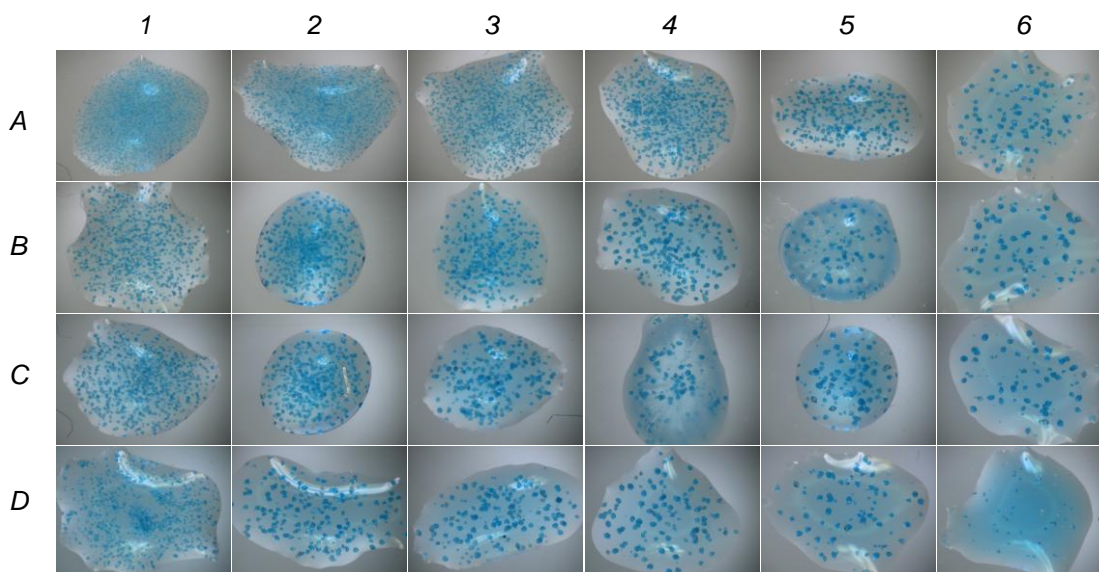


Figure 2.18. Box #65 at ~24 days of Az-PHM in imidazole at pH 8.0. Conditions are identical within columns with the exception of the amount of imidazole used. Imidazole concentration is consistent across a row but increases down a column. PEG concentration decreases across a row from left to right but is consistent down a column.

Growing and screening large crystals provides strong reflection data, however, one of the trade-offs of obtaining strong data is large mosaicity values. This is consistent with what was observed. Mosaicity, the second criterion, is a measure of the degree of overall disorder within a crystal. The root of this word, mosaic, is a great description of this concept because it suggests that although a crystal may appear to be one crystal, it is really composed of little crystals regularly referred to as mosaic blocks. Large crystals have higher mosaicities simply because there is more opportunity for disorder. More abstractly put, if mosaic blocks do not stack properly early on in crystal formation, even small disorder can be exaggerated as the crystal grows larger.

In order to combat the problem of high mosaicities in the Az-PHM crystals, two approaches were used. The first approach worked toward growing crystals more slowly, and the second called for dehydration of the crystals that had already formed. In order to cause slower crystal formation, protein drops were diluted with water, which increased the total volume of the drop. Typically, a volume of 2 μ L was added to the drops, which

contained 2 μL of protein and 2 μL of the reservoir solution. Larger drops equilibrate at a slower rate and as water is lost from the drop slowly over time, the protein does not reach saturation as quickly. Diluting Az-PHM protein drops with water caused crystals to increase in size over a longer period of time, but there was no statistical drop in mosaicity. This may be because the protein drops were only diluted by 2 μL of water at the most. Increasing the volume of the drop by a greater amount may lead to even larger crystals with smaller mosaicities.

Because the first approach was largely unsuccessful, a technique called dehydration was used. In theory, dehydrating crystals after they have formed removes excess water and causes mosaic blocks within the crystal to shift to become more ordered. Mosaicity values collected from crystals with and without dehydration are shown in Table 2.5 below. The dehydrated Az-PHM crystals show a marked improvement in mosaicity values when compared to those that were not dehydrated.

Table 2.5. Mosaicity comparison of crystals from two wells with identical conditions. Crystals in well C1 were dehydrated and show much lower mosaicities than crystals from well D2 which were not dehydrated.

Box #, well #	Crystal	Dehydrated?	Mosaicity (°)
68, D2	Y13	No	-----
	Y14	No	0.55
	Y15	No	0.99
	Y16	No	0.62
69, C1	BB1	Yes	0.41
	BB2	Yes	0.48
	BB3	Yes	0.48
	BB4	Yes	0.36

The third criterion, resolution, is also directly affected by the order within a crystal. The more ordered a crystal is, the higher the resolution will be. Because high resolution reflections are located where x-rays diffracted at higher angles must converge, the impact of thermal motions and crystal disorder is greater, making these reflections less intense or non-existent. Because of this phenomenon, it is difficult to obtain high resolution data. Resolution can be improved, however, by diffracting crystals at low

temperatures to slow down thermal motions and by growing crystals with more order. Seemingly, crystals that are grown at a slower rate will be more ordered because given time, protein molecules will add to the crystal more purposefully instead of hastily.

Unfortunately, large Az-PHM crystals grown at a slower rate did not diffract to a higher resolution. The best explanation for this is that the large crystals that were grown slowly did not have improved mosaicities, and because crystal disorder has a great impact on high-resolution reflections, there were none that appeared on the diffraction pattern. However, because dehydration gave an improvement in mosaicities, there should also be an improvement in resolution, which is precisely what was observed. Diffraction patterns seen in Figure 2.19, illustrate this improvement. The patterns in these photos come from two wells with the same conditions except for PEG concentration in the drop. Crystals that were dehydrated consistently showed reflections between 2.0-2.5 Å and often times showed reflections beyond 2.0 Å, while the crystals not treated do not show diffraction between 2.0-2.5 Å.

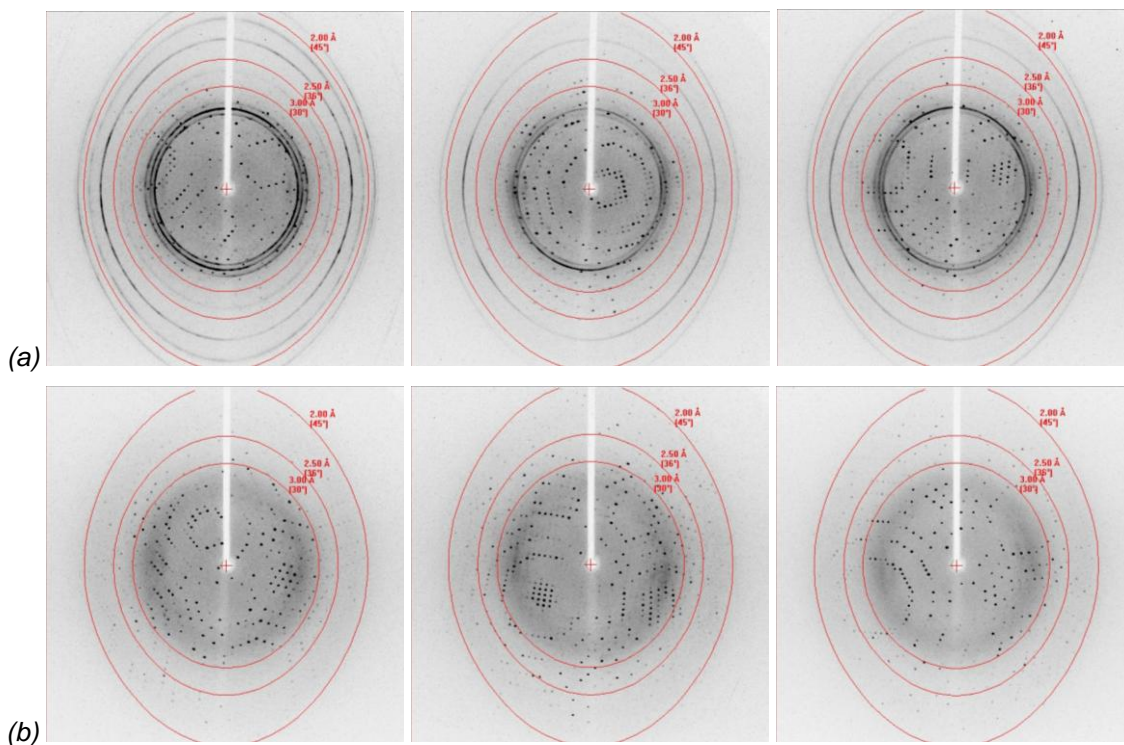


Figure 2.19. Diffraction patterns from crystals grown in (a) Box #70, well B1 and (b) Box #68, well D2. The only difference in crystallography conditions between (a) and (b) is that the drop in (a) contained 2 μ L of reservoir solution with 5% PEG-8000 whereas the drop in (b) contained 7% PEG-8000.

Besides growing large crystals and using dehydration, a final technique called annealing was attempted in which a frozen crystal placed in the nitrogen stream was warmed for short time and then frozen again in the stream. This technique was mildly successful regarding resolution, but unfortunately, mosaicity values increased dramatically. The diffraction patterns in Figure 2.20 show reflections prior to annealing (left) and after annealing (right). The resolution improved from 2.0-2.5 Å, but in almost all cases, mosaicity values doubled.

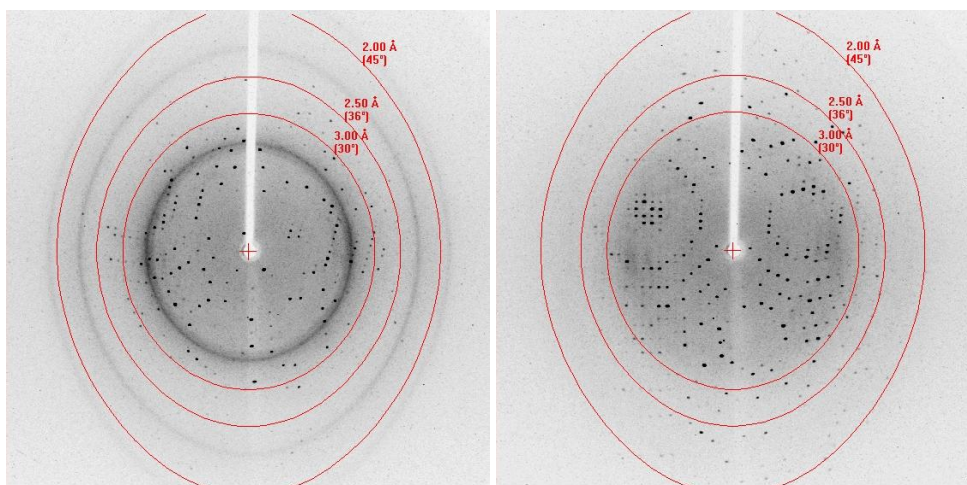


Figure 2.20. Diffraction patterns from Z9 crystal harvested from Box #67, well C1. The pattern on the left was obtained before annealing and the pattern on the right after annealing for 15 seconds.

The fourth and final criterion used to judge a crystal's quality is the presence of ice rings. Ice rings are intense rings visible on a diffraction image due to crystalline ice formation during freezing. Flash-freezing is an important last step in preparing a crystal for screening as crystalline ice can form from water vapor in the air or from liquid in the drop that is looped up during pinning. Sometimes ice rings do not affect the diffraction of the crystal itself, but they can be a hindrance in predicting the unit cell because they appear on the diffraction pattern. Though the CrystalClear software can compensate for ice rings, it is best to avoid them since the software omits all data obscured by ice rings.

To freeze a protein crystal efficiently, the pinned crystal must move from room temperature air to the liquid nitrogen at $-210\text{ }^{\circ}\text{C}$ as quickly as possible. A sharp temperature gradient was created by filling a dewar to the brim in order to provide a uniform transition between temperatures. Typically, this was sufficient to prevent water

vapor from crystallizing. However, preventing liquid in the drop from freezing was more difficult. PEG is a natural glassing agent that immobilizes water, preventing it from forming into crystalline ice. But, because many of the larger protein crystals only required small amounts of PEG to grow, the liquid in the drop often froze upon pinning and freezing crystals. To combat this problem, many different freezing methods were attempted without success, however, one technique called cryoprotection consistently worked at avoiding ice formation from liquid in the drop. At 4 °C, Az-PHM crystals were transferred between a series of three solutions each containing identical concentrations of reservoir reagents with an increase in PEG concentration and the addition of glycerol. The higher concentration of PEG and glycerol provided enough glassing agent to prevent ice formation. A visual comparison of crystals with and without cryoprotection is shown in Figure 2.21. The crystal without cryoprotection (2.21b) has liquid from the drop that was looped during pinning and froze to give crystalline ice which is evident in the crystal's corresponding screen image. The crystal with cryoprotection, however, is clear and ice rings are absent from its screen image.

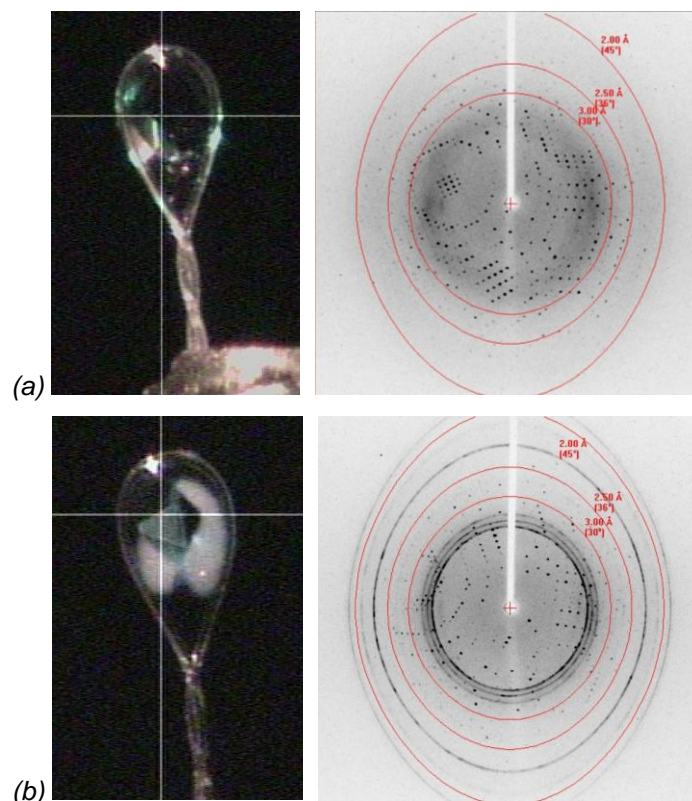


Figure 2.21. (a) Cryoprotected crystal from Box #70, well A3. (b) Crystal from Box #68, well C3 that was not cryoprotected.

After growing and screening hundreds of Az-PHM crystals, trends observed in crystal growth were used to control intensity, mosaicity, resolution and the formation of ice. Crystals were obtained that gave strong intensities, mosaicities below 0.60, and resolutions beyond 2.0 Å in the absence of ice rings by reproducibly growing large Az-PHM crystals that were subsequently dehydrated and cryoprotected. Because diffraction-quality crystals can be selectively grown, the collection of a full data set is possible wherefrom a crystal structure can be obtained.

CHAPTER 3

Chapter 3.1 Introduction

In times past, collecting diffraction data and processing it was no small task. Before the development of modern technology, data collection was a tedious process. Until approximately 50 years ago, weeks were spent collecting images, where each image was collected on a piece of photographic film that required the utmost care in alignment. For those protein crystals that were susceptible to radiation damage, a new crystal was mounted each time an exposure was captured on film. Once all of the images were collected, processing the data took weeks and even months. Reflections on each frame were individually counted and indexed by hand, and were then subsequently “measured” for intensity using a qualitative “eye estimation” scale. After reflections were indexed, complicated mathematics were carried out in order to solve the crystal structure. One two-dimensional Fourier projection took several days of steady calculation. Months and years were spent growing crystals and obtaining a single crystal structure (38).

While growing protein crystals still remains a lengthy and tedious process, great technological advances in diffraction instrumentation and software development have made it possible for crystallographers to collect data with little to no effort involved and process it in a few short hours to obtain a preliminary model. Because modern technology affords the luxury of fast data processing, it has been possible for many researchers to learn crystallography. Hence, understanding the complexities of diffraction and the process by which data is obtained, reduced and modeled is now the rate-limiting step in going from crystals to structures.

The ultimate goal of processing diffraction data is to obtain an electron density map that conveys a three-dimensional depiction of the repeating unit within the protein crystal. In essence, the map is created by summing the waves that are diffracted from a crystal in much the same way that an image is created by a microscope when a lens collects and sums waves of visible light that have been scattered by the object. The waves diffracted from a protein crystal are called structure factors. Because structure factors are waves, they have both amplitudes and phases that must be calculated in order to create an electron density map. Structure factor amplitudes are obtained directly from

the experimental reflection intensities whereas phases must be obtained indirectly, very often through a process called molecular replacement (17).

Powerful crystallographic software programs allow amplitudes and phases to be computed with ease. Two such programs are implemented in this work, namely CrystalClear r2.1 and CCP4 6.1.3. CrystalClear, a software program created by Rigaku, contains algorithms that make data collection a fully automated process. Once diffracted reflections are obtained, CrystalClear integrates them to obtain their intensity information and then takes the experimental intensity data and predicts crystal symmetry. Next, intensities are scaled and those that are symmetry-equivalent are merged to create a list of unique reflection intensities. Later, intensities are converted to structure factor amplitudes, a process carried out by CCP4. CCP4 (Collaborative Computation Project No. 4) is free software for macromolecular crystallography that is capable of molecular replacement, refinement and even validation of a final structural model. Upon calculation of structure factor amplitudes, CCP4 performs molecular replacement, which uses atomic coordinates from a similar protein model to estimate phases. During refinement, phases from the model are coupled to their corresponding structure factor amplitudes. Finally, amplitudes and phases are input into a third program called Coot that computes the Fourier transform of the structure factors and creates an electron density map in three dimensions.

The protocol described in the rest of the chapter contains parameters that best suited the diffraction data that was collected on an Az-PHM crystal grown in Tris buffer. Data is analyzed for quality following each experimental section discussing CrystalClear and CCP4 parameters.

Chapter 3.2 Data Collection and Data Reduction in CrystalClear

Once a crystal is found that diffracts beyond 2.0 Å, has a mosaicity value less than 0.60, has intense reflections and is free of ice, a full data set can be collected. Data was collected on one such crystal, Az-PHM crystal H6, that grew in Box #26, well D6 in a 4 µL hanging drop at an initial concentration of 16.7 mg/mL in 75 mM Tris HCl pH 7.07, 25 mM lithium nitrate, 7.5% polyethylene glycol-2000 and 2.5 mM copper

chloride. The drop was equilibrated against 0.998 mL of the same reagents with double the concentration, the only exception being that the concentration of Tris HCl in the reservoir was 100 mM. The crystal was pinned and frozen without any prior dehydration or cryoprotection. Upon screening the crystal with a 5-minute exposure per frame, it diffracted to 1.5 Å with a mosaicity of 0.46°. A photo of one of the screen images is displayed in Figure 3.1 where there are intense spots visible alongside a few very faint ice rings.

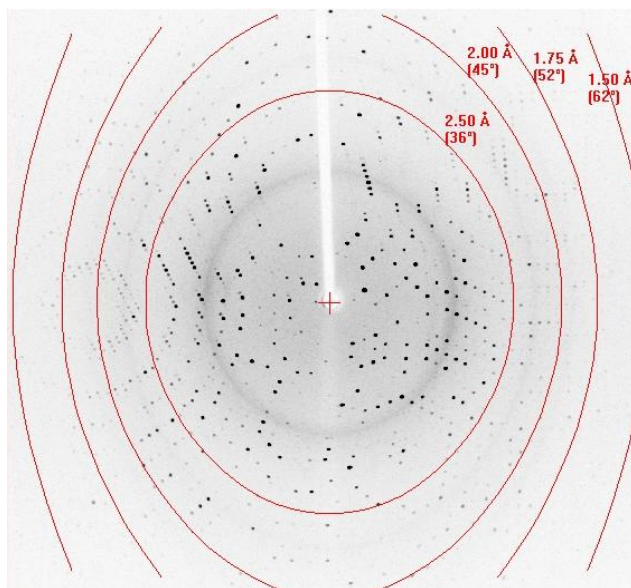


Figure 3.1. Screen image from the Az-PHM crystal H6, which was grown in Box #26, well D6 using Tris HCl buffer at pH 7.07, PEG-2000, lithium nitrate and copper chloride.

A full data set was collected on crystal H6 using a Rigaku R-Axis Rapid II Diffractometer and the d*TREK processing suite in CrystalClear r2.1. Recall that during the screening process (described in Chapter 2.4), the unit cell and crystal orientation are determined. CrystalClear uses this information along with completeness and redundancy conditions in order to identify a strategy for data collection in which the number of images that might otherwise be collected is greatly reduced, saving time and unnecessary radiation of the crystal. The *Completeness* and *Redundancy* conditions are entered by the user in the Main tab of CrystalClear's Strategy window, shown in Figure 3.2, which also allows parameters such as *Search resolution*, *Image rotation width*, and *Anomalous (I^+ , I^-)* to be input. A resolution maximum of 1.2 Å was chosen since it was slightly

greater than the resolution where spots were found on the screen images. The image rotation width is a value that specifies the degree to which the crystal will oscillate during the exposure of one frame. Its value is varied depending on the size of the crystal's unit cell. Because protein crystals typically have large unit cells, a small oscillation range of 0.5° (anything below 3° is considered fine-slice) is commonly used to avoid reflection overlap (33). The target completeness was set to 99% with a 2% tolerance since all (or nearly all) reflections needed to generate a complete set of data (given the crystal's symmetry) should ideally be collected. These reflections are termed "unique" reflections (39). The option to collect *Anomalous* ($I+$, $I-$) was selected because it was questionable whether or not the crystal would give an anomalous signal, and since the intensities can be averaged together in later steps, there was no harm in choosing this option if it was not needed. A minimum redundancy was not specified (33).

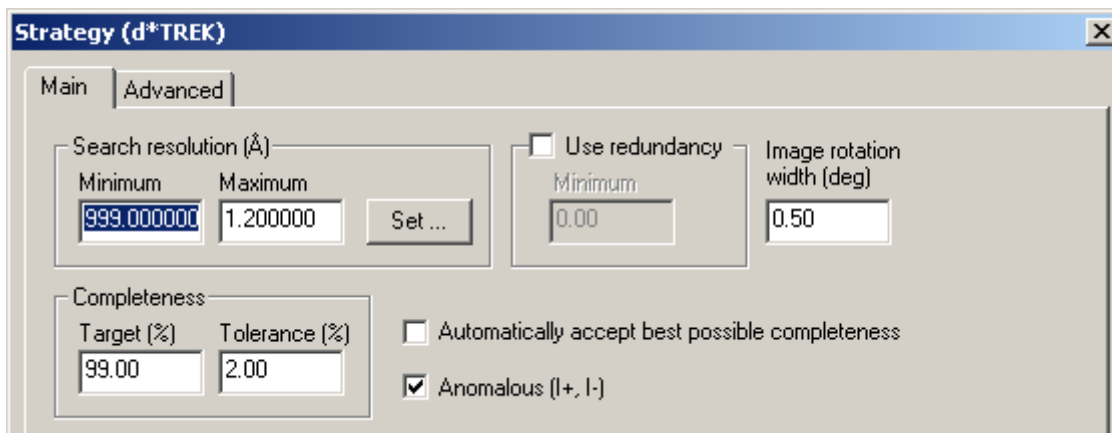


Figure 3.2. The Main tab of the Strategy window from CrystalClear's d*TREK processing suite. Parameters specified here are those used to identify a strategy which was used to collect a full data set on Az-PHM crystal H6 from Box #26.

There are many other strategy parameters located on the Advanced tab, shown in Figure 3.3, but these parameters do not typically require any adjustments as was the case for crystal H6. However, depending on the instrument, it may be necessary to adjust omega rotation angles which place the goniometer cradle in a position that does not interfere with diffraction. This problem was an issue when collecting data on crystal H6, but was avoided by changing the *Start* and *End Angles* in the Collect Images window which appears after the strategy is run.

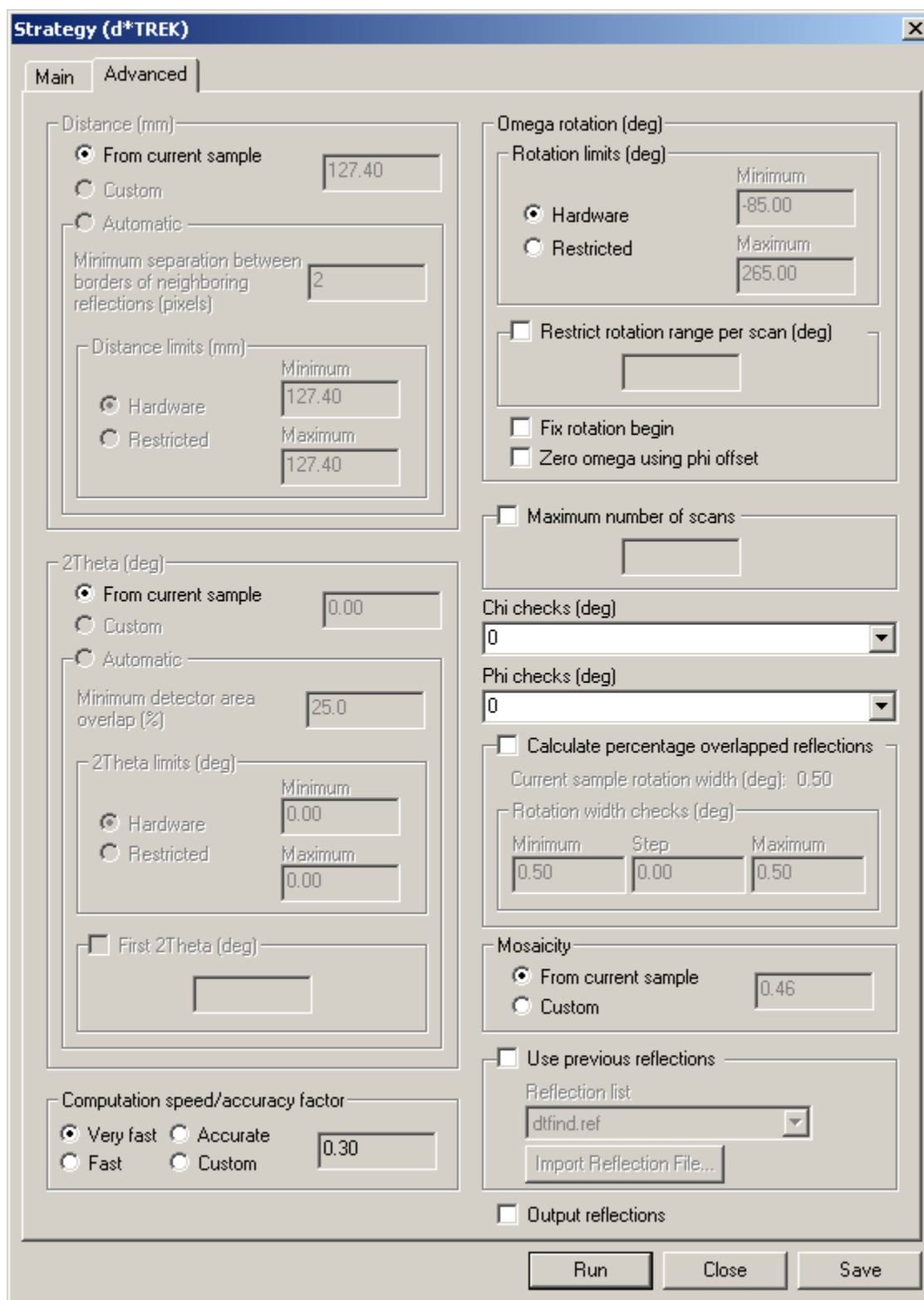


Figure 3.3. The Advanced tab of the Strategy window while using the d*TREK processing suite in CrystalClear 2.1. All parameters designated here were used in the strategy for data collection for Az-PHM crystal H6.

After running strategy, the algorithm calculated the percentage of data completion based on predicted reflections while considering the values specified in the Main tab. If the target percentage is reached, a schedule for data collection appears in the Collect Images window. A schedule contains detector and goniometer settings selected by the strategy algorithm along with the number of images to be collected at a specific angle. The schedule visible in Figure 3.4 shows the settings used to collect data on crystal H6. The *Start* and *End Angles* were adjusted to 5.00 and 155.00, respectively, to prevent the goniometer cradle from casting a shadow in the diffraction images, and the *Exposure Time* was increased to a total of 20 minutes per image. Because the H6 crystal was predicted to be highly symmetric (an orthorhombic unit cell), only one scan was needed to ensure all unique reflections would be collected (33).

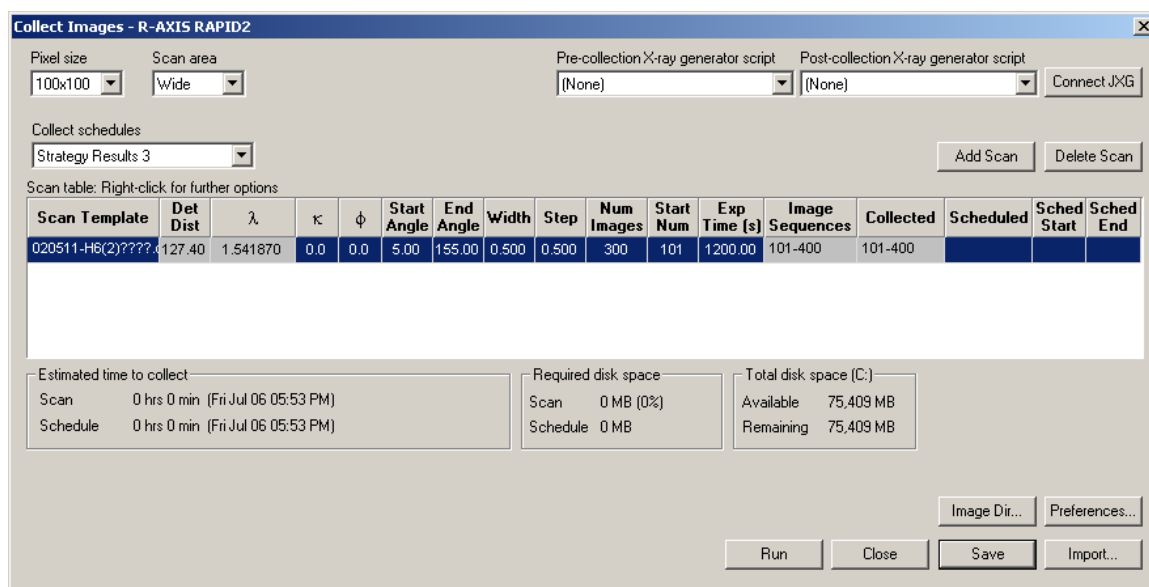


Figure 3.4. The Collect Images window in CrystalClear 2.1. The schedule shown in the figure was used to collect data on crystal H6.

Following the selection of the schedule resulting from strategy, the H6 crystal was exposed to copper K_{α} x-ray radiation (50 kV, 40 mA) through a 0.3 mm capillary optics collimator under a chilled stream of nitrogen gas at -150°C , and a total of 300 diffraction frames were collected. After collecting a full data set of images, the process of data reduction can begin, starting with the integration of reflections in order to obtain intensity information. However, a new unit cell was predicted before integrating. Even though

the crystal's unit cell was already determined during screening, obtaining a more accurate unit cell ensures that integration occurs in the precise location of the spot, in order to avoid integrating background noise. A more accurate unit cell that represents all reflections in the full data set was found when several parameters from the unit cell prediction protocol in Chapter 2.4 were changed, which are described below. Reflections used to determine the unit cell were selected from several images throughout the collection, including the first and last image, instead of spots from two images only. This was done so that the reflections used were an accurate representation of the entire data set (see Figure 2.8). When the reflections were indexed, the location of the beam was checked as there was more opportunity for shifting given that the full data collection took place over several days (see Figure 2.11). The "All" macro was used during refinement of the unit cell instead of the "Most" macro so that the radiation source location was refined along with the detector, crystal and goniometer. When reflections throughout space are used for indexing, everything should be refined. Finally, the number of cycles was changed from 100 to 30 and the X and Y rejection limits were adjusted to 1 (see Figure 2.13). Because CrystalClear's new refinement algorithm includes up to 5 derivative calculations per cycle (instead of 1 which was used in older versions), convergence is reached much faster and 100 cycles is not necessary (34).

After predicting a more accurate unit cell, data reduction began on the full collection of images when all reflections were measured for their intensities through integration. Prior to selecting integration parameters, a mask file was created that defined the shadow of the beam stop which prevented d*TREK from harvesting erroneous intensity information. Parameters for harvesting intensities are shown in the Main tab of the Integrate Reflections window in Figure 3.5, and several of the defaults were consciously changed. First, all images of the collection (which excludes any screen images) were used as the *Template*, and the resolution *Maximum* was set to a value just larger than the resolution on the images where spots are visible. A high maximum was entered since weak data is easily excluded during future scaling and averaging steps. Lastly, a value of 2 for the *Padding* parameter was used which is typical for fine-slice data. Because data from an individual reflection can be located on several images, this

value controls the number of images on either side of the one being integrated that are searched for a reflection's exact position. The *Images per batch*, *Box* and *Batch prefix* values used were default parameters, which are all shown in Figure 3.5 (33).

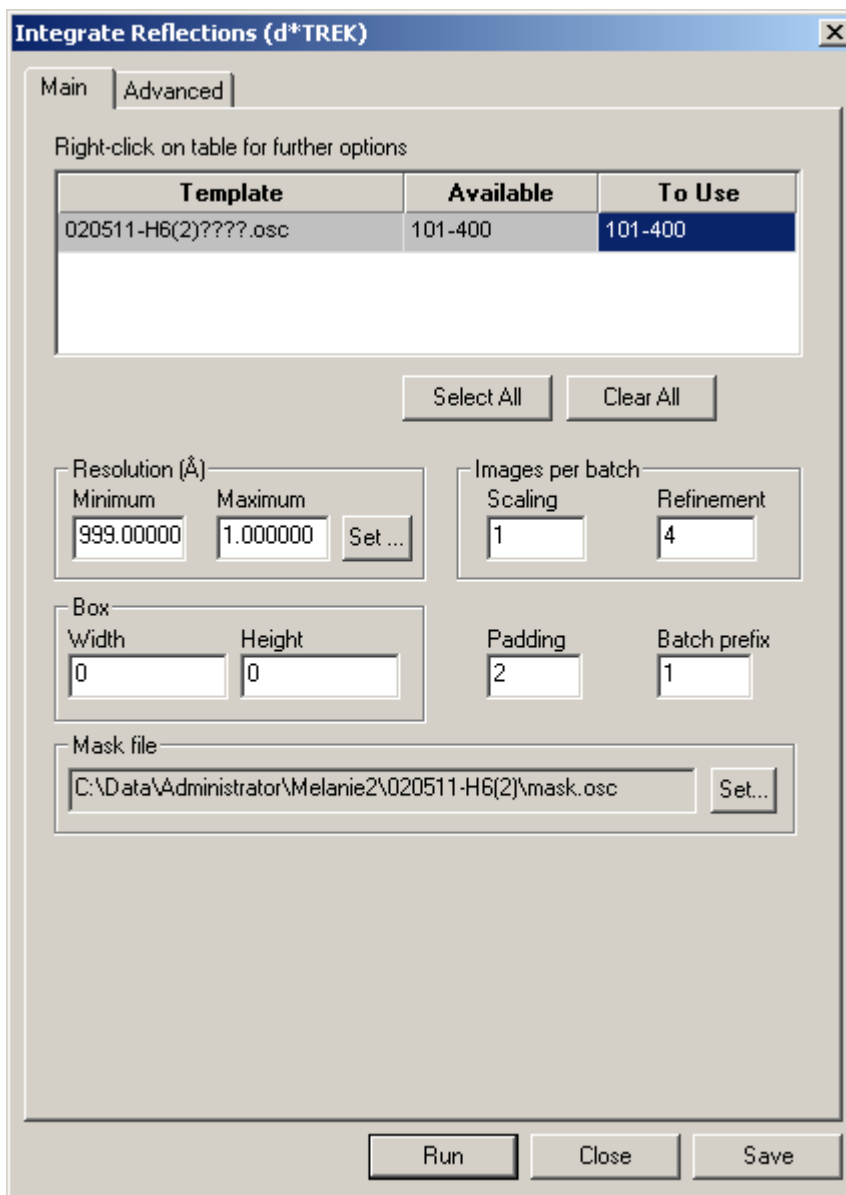


Figure 3.5. The Main tab of the Integrate Reflections window in CrystalClear 2.1 containing parameters used to integrate the diffraction data from Az-PHM crystal H6.

On the Advanced tab of the Integrate Reflections window shown in Figure 3.6, more parameters are available that offer greater flexibility when measuring intensities. Because crystal H6 is a protein crystal, it was advantageous to *Perform profile analysis*

during integration with a *Number* of 50 and *Maximum images* totaling 7. This function creates reflection profiles from well-defined reflections and uses the resulting statistics to best estimate the intensities of weak spots. The *Refinement macro* All was used when refining before integration. Before integration took place on each image, *Find reflections* and *Predict reflections* were both selected before refining using 2 refine batches each, and while finding reflections, only those pixels having an *I/sigma* of 5.00 or greater were included in spots. It is most important to find and predict spots before integration when there are multiple scans present so that any defect in goniometer position after it moves in additional scans does not affect the software's ability to locate the spots and accurately predict and integrate them. Because the prediction depends on the mosaicity value used, it was very important that the *Mosaicity model* have a value of 1 for *Multiply* and a value of 0 for *Add*. When these values were input, the software used the refined mosaicity from the unit cell prediction. For a set of well-predicted images, a high mosaicity used in the integration causes background noise to be integrated and taken for real reflection intensity. Lastly, by opting to *Check overlaps*, any reflections that overlap by a fraction greater than or equal to the value designated are excluded from integration. For H6, any spots that overlapped by $\geq 3\%$ were not harvested. The values for all other parameters not discussed that were used in integrating crystal H6 are shown in Figure 3.6 (33).

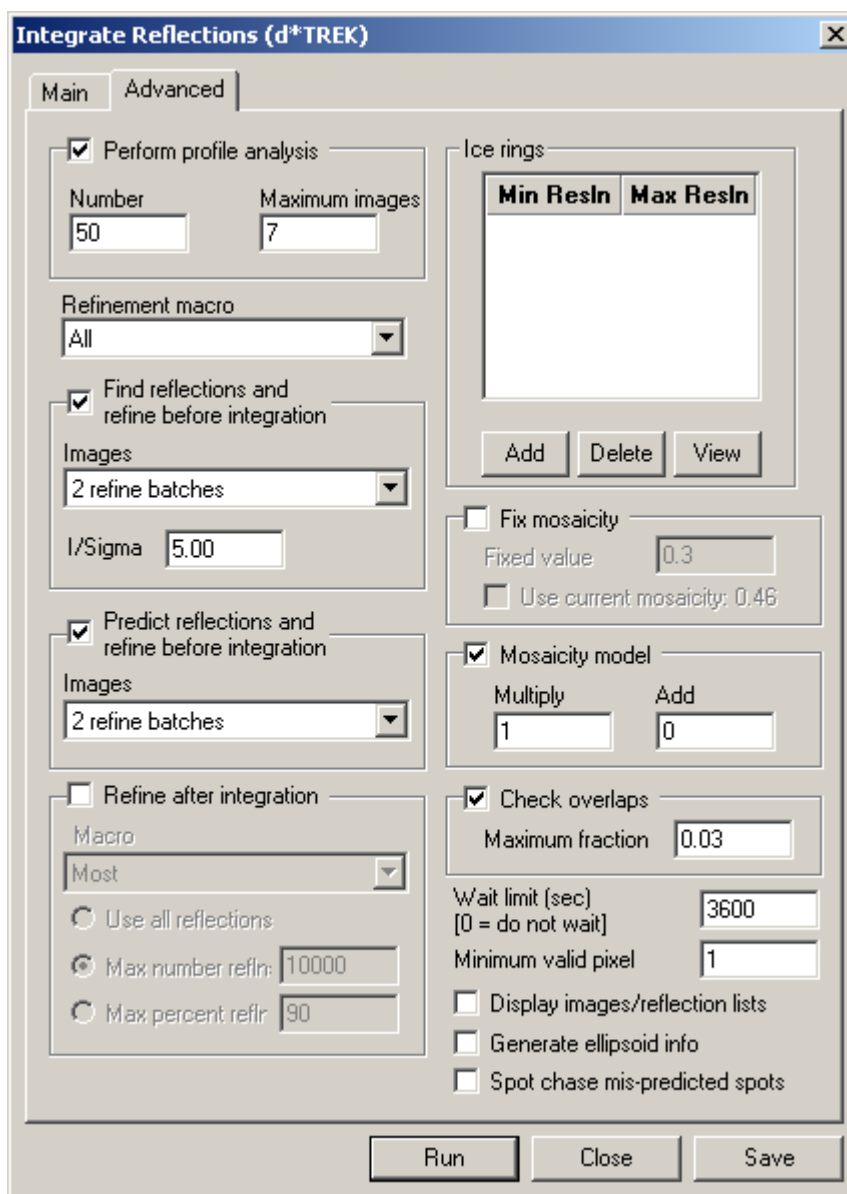


Figure 3.6. The Advanced tab of the Integrate Reflections window in CrystalClear 2.1. The parameters shown in the figure were used in integrating the data from Az-PHM crystal H6.

After the intensity data was harvested, a series of checks were employed to more accurately pinpoint the Laue class, centricity and space group that best characterize the symmetry of crystal H6. It is important to note that although these checks are vital in determining that the correct symmetry is chosen, it is not possible to be sure that the exact symmetry of the crystal has been determined until the electron density map can be viewed (39).

The first, Laue Check, is designed to use the integrated data to determine the Laue symmetry. The data are averaged for each Laue symmetry group and the resulting R_{merge} values for each group are compared. For H6, the parameters used are given in Figure 3.7. The *Reflection list* used, dtprofit.ref, contained the integrated data. Both *Average Bijvoets* and *Batch scale*, using a maximum of 50 reflections per batch, were selected. By choosing to average Bijvoet (or Friedel) pairs, any anomalous signal that is present was removed, avoiding the potential for incorrect symmetry assignment. Batch scale was selected for the same reason, even though the integrated data was all from the same crystal and the crystal was cryocooled to avoid decay. Under the *Highest Laue symmetry*, All classes were considered in determination, and under “Unit cell reduction”, a *Maximum least squares residual* of 0.1 was input which limits the number of possible solutions to only those that are most likely. Lastly, *R-merge* values of 0.08 and 0.15 for *Ideal* and *Poor*, respectively, were used along with an *Incremental increase ratio maximum* of 1.5. By limiting the range of R-merge values, a shorter list of possible Laue classes is produced (33).

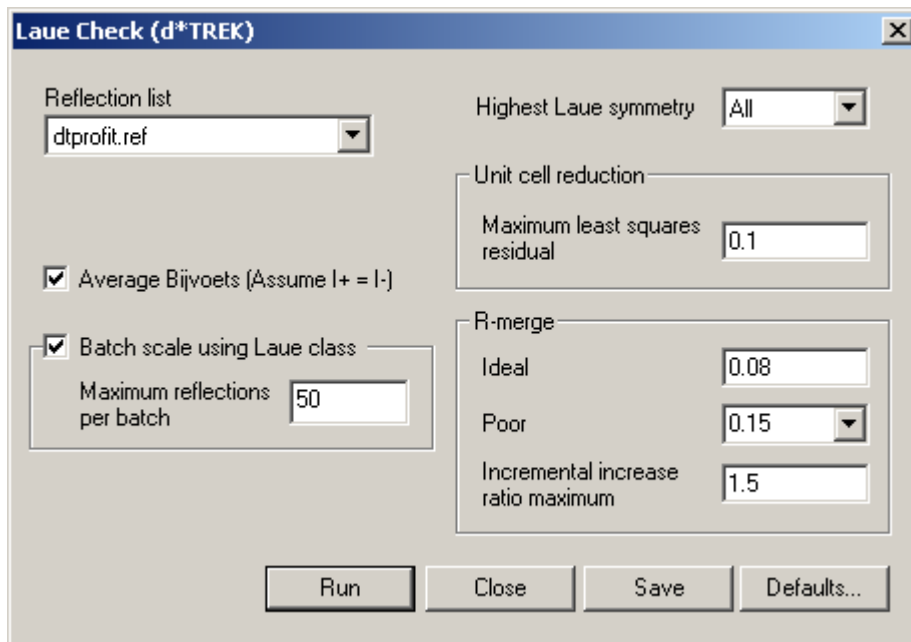


Figure 3.7. Laue Check window containing parameters used for Laue class determination for Az-PHM crystal H6.

The results of the Laue check for crystal H6 can be seen in Figure 3.8 where the solution highlighted in green was the recommended choice for Laue group symmetry. Each Laue group refers to a limited set of crystallographic point groups within a crystal system. Even though the unit cell dimensions already suggested an orthorhombic cell, for which there is only one possible Laue class, all possible unit cells are reconsidered using the integrated data, so that the Laue class is not biased. For crystal H6, Laue group “mmm” was suggested by the software, which is indicative of the orthorhombic crystal system. This group was recommended because it was the solution with the highest symmetry and the one with an R_{merge} value closest to the ideal of 0.08 (33).

Laue Results (d* TREK)

Highlight row to select Laue class:

Laue Class	Unique Axis	Lattice Used	Refln Groups	Calc Mult	Observed Mult	Rmerge	Pass?
-1	-	aP	18197	1.00	2.00	0.05	N/A
2/m	a	mP	70201	2.00	2.24	0.04	[PASS]
2/m	b	mP	85557	2.00	2.15	0.05	[PASS]
2/m	c	mP	98743	2.00	2.13	0.04	[PASS]
mmm	-	oP	94126	4.00	2.75	0.05	[PASS]

OK Cancel

Figure 3.8. Laue Results from Az-PHM crystal H6. The Laue class highlighted in green is the solution with the highest symmetry and R_{merge} closest to the ideal.

The second check uses the integrated data in dtprofit.ref to determine whether or not the space group is centric. Because all naturally-occurring protein molecules are chiral, it was not necessary to perform this check on crystal H6. Nevertheless, centricity was checked and the results can be seen in Figure 3.9.

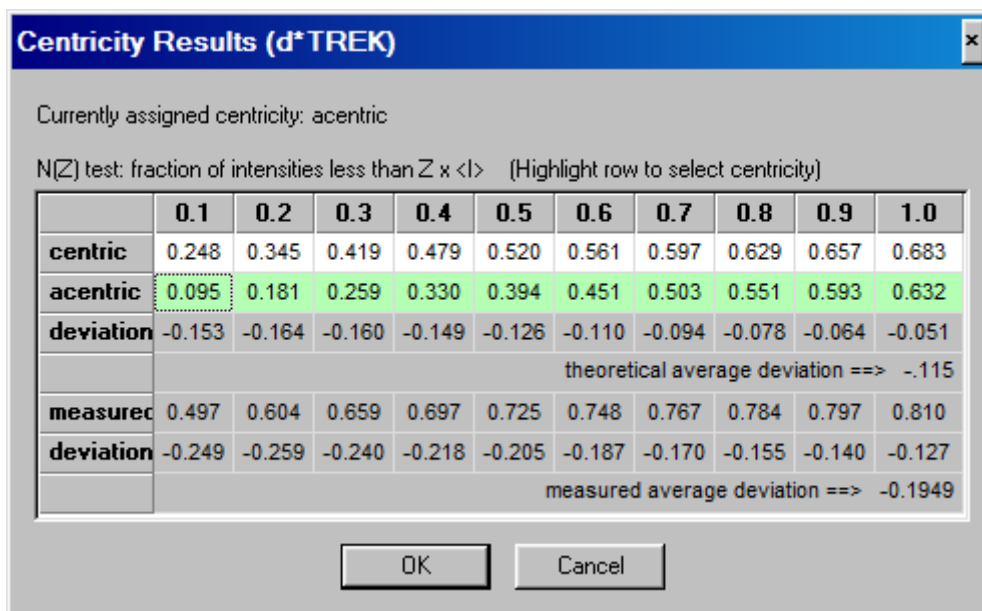


Figure 3.9. Centricity Results from Az-PHM crystal H6. The green highlighted solution is the one recommended by CrystalClear.

The third and final check determines space group by analyzing the reflection data for any systematic absences that are consistent with the laue group and centricity previously assigned. The correct determination largely depends on the I/σ parameter entered by the user. For crystal H6, the integrated data from dtprofit.ref was used and a value of 4 was entered for $\langle I/\sigma \rangle$ tolerance. The resulting data and suggested space group, $P2_12_12_1$, are seen in Figure 3.10. The $\langle I/\sigma \rangle$ tolerance value was important in determining the correct space group since any value less than 4 would have given a different space group. For this data set, it is clear that the $\langle I/\sigma \rangle$ data for $h00$, $0k0$, and $00l$ reflections where h, k and l are odd is small compared to all other $\langle I/\sigma \rangle$ values. These small values indicate areas where no real spots were actually located or in other words, where spots were systematically absent. Defining these locations where no spots were actually integrated allows the space group to be determined based on reflection data (33).

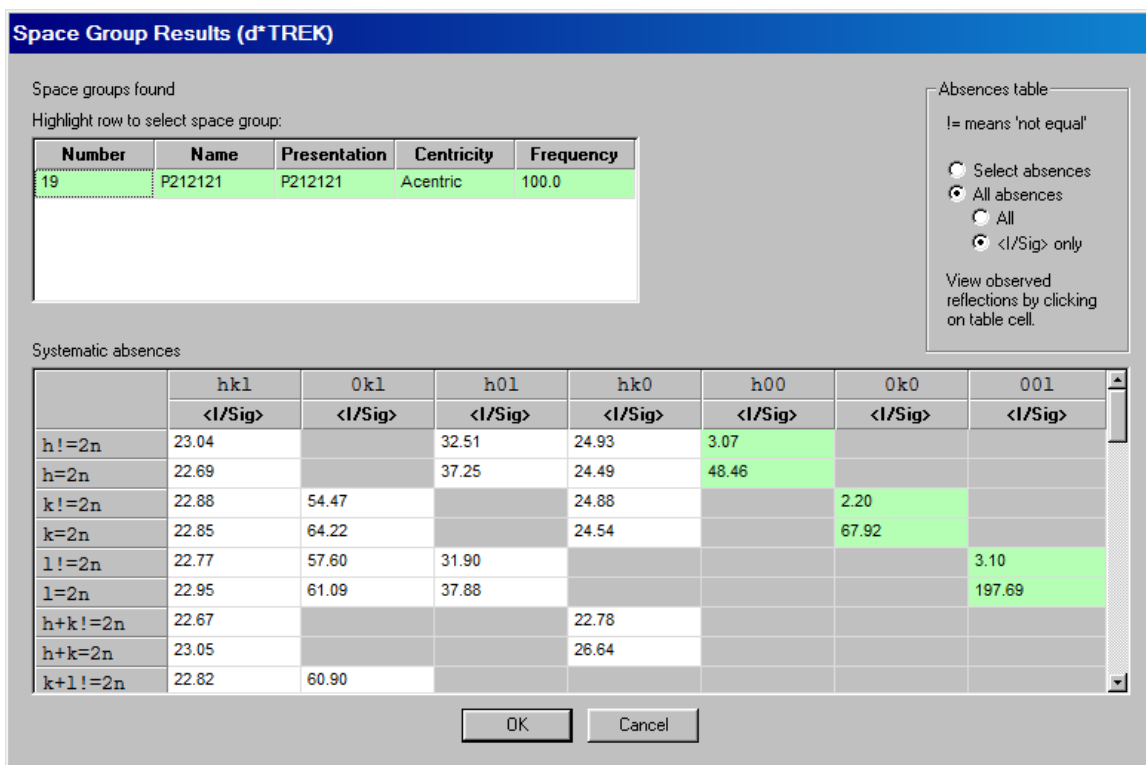


Figure 3.10. Space Group check results from Az-PHM crystal H6. The green boxes indicate systematic absences where the <I/Sig> is low.

After gathering integration data and symmetry characteristics, the final step in data reduction for crystal H6 was carried out by scaling and averaging the integrated intensity information. Integration data is scaled when there is reason to suspect that intensities from different sections of the collection differ. These differences are due to a number of factors including different exposure times, radiation damage, fluctuating source intensity and different absorption of x-rays due to varied paths through the crystal. Besides these systematic errors, random outliers should be rejected and therefore, data correction via scaling is a necessary step in data reduction. After data is scaled, d*TREK averages the intensity data and the output is a file that contains a list of unique reflections. For crystal H6, minimal scaling was required before reflection intensities were averaged. These collection aspects are reflected in some of the parameters used to scale and average the integrated data in dtprofit.ref, which are visible in Figure 3.11. In the scaling model section, *Batch scale* was deselected because the entire crystal was bathed in x-rays and all data was collected from the same crystal at a constant exposure rate. *B-factor* was also deselected because the crystal was cryocooled during the entire data collection,

preventing any major decay. Also in the scaling model section, *Scale I+ and I- separately* was deselected since Friedel pairs do not exist in data with a non-centrosymmetric space group like $P2_12_12_1$, and when selecting *Absorption correction*, the *REQAB* Fourier 8.3 algorithm (40) was chosen. During scaling, *reflections with I/Sigma* < 3.00 were excluded and *Systematic absences* were rejected beforehand. Lastly, the *Resolution Maximum* was set to 1.3 \AA and the *Error model* weights were determined automatically by *d*TREK*. All other parameters were left default as shown.

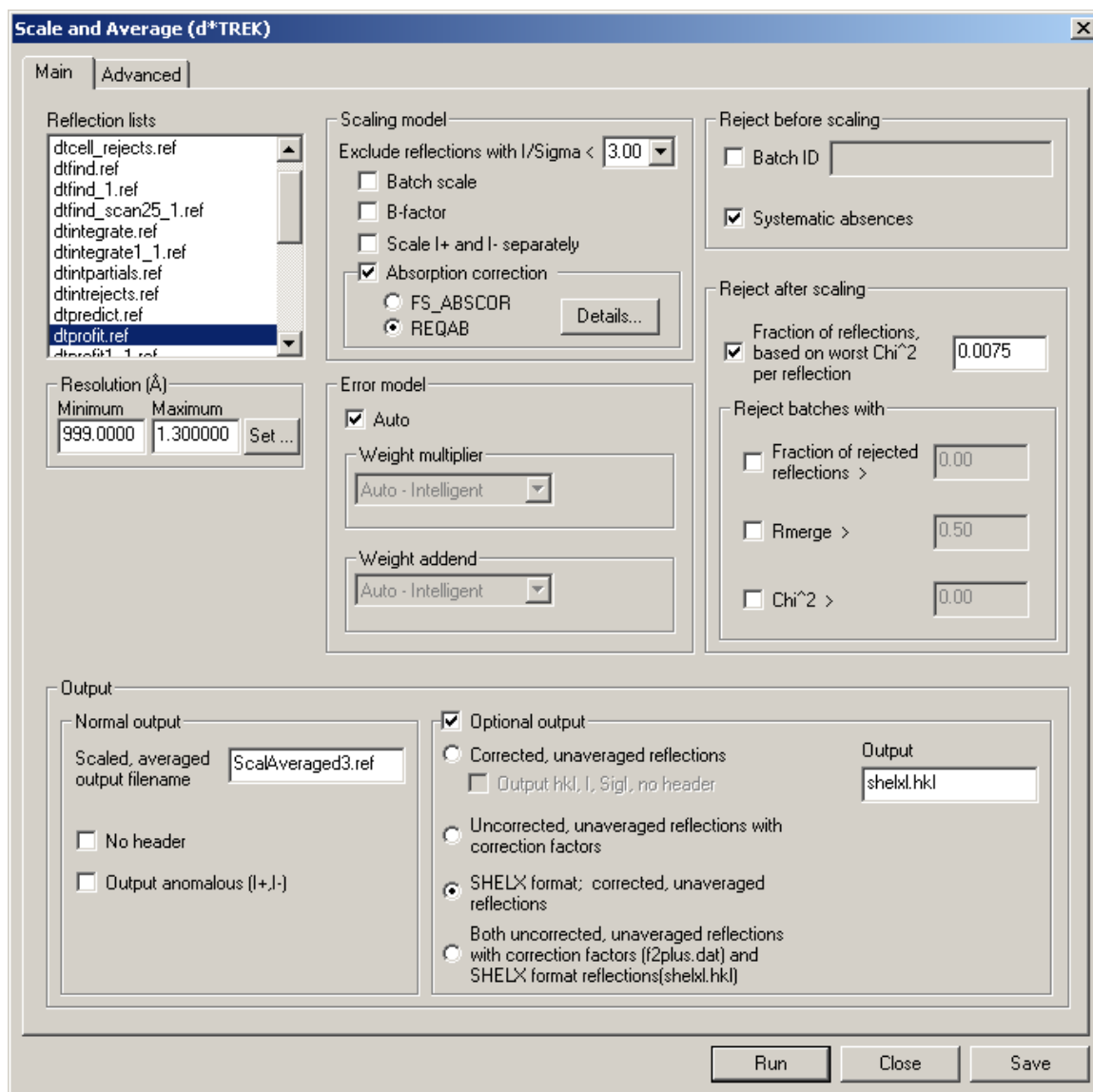


Figure 3.11. The Main tab of the Scale and Average window in CrystalClear 2.1. All parameters shown were used to scale and average integrated data from Az-PHM crystal H6.

Only a few more scale and average parameters exist in the Advanced tab for scale and average, visible in Figure 3.12. All parameters were left as default except for the *Global/per batch range argument(s)*. This value was changed to 0.002 so that the magnitude of the applied scale factors was reduced over the entire data set.

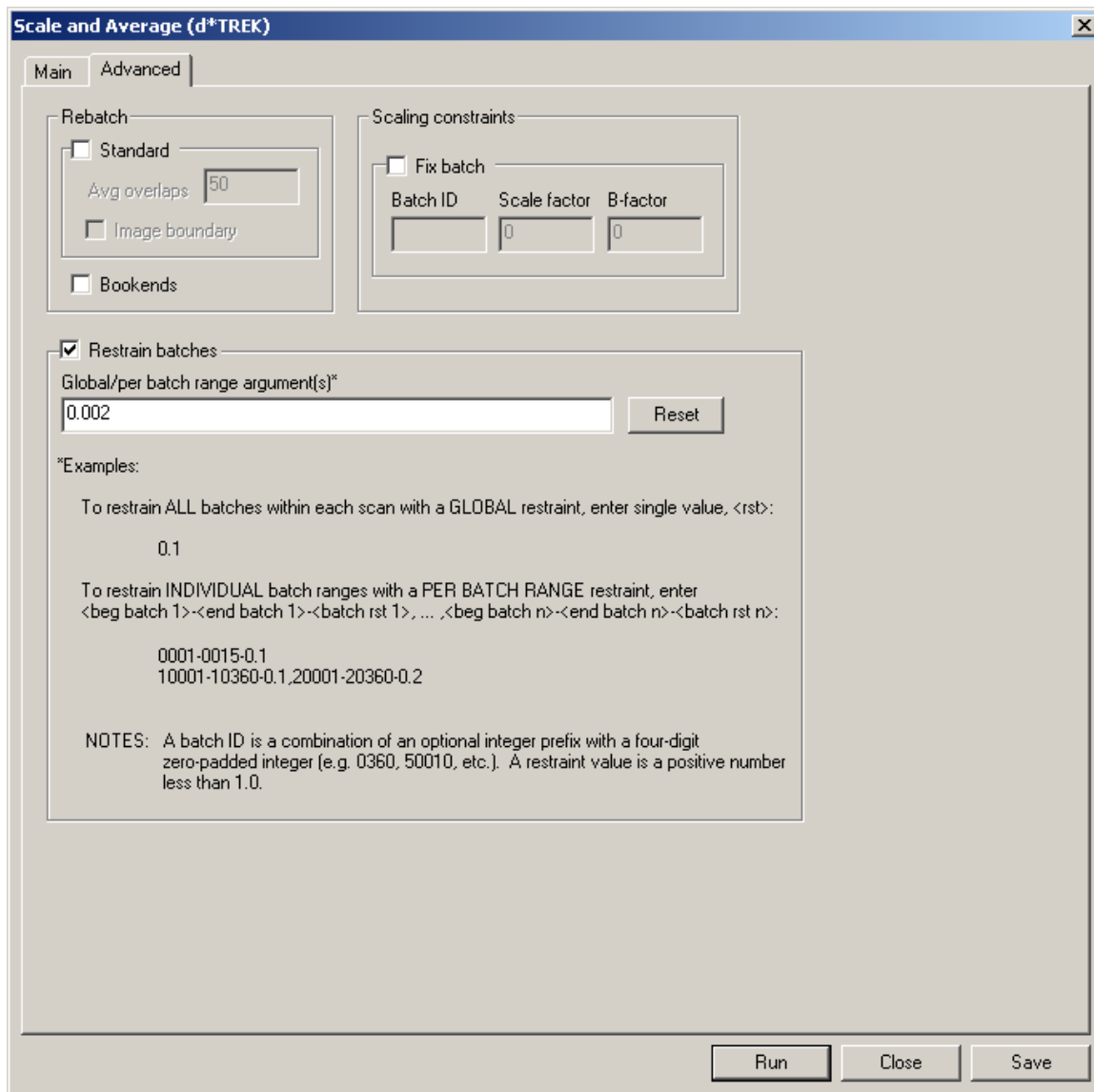


Figure 3.12. The Advanced tab of the Scale and Average window in CrystalClear 2.1. All parameters shown were used to scale and average integrated data from Az-PHM crystal H6.

After scaling and averaging the integrated reflections, statistics are generated which help assess the quality of the diffraction data. Some of the results include merging statistics, completeness, average redundancy, signal strength, resolution and final unit cell

dimensions. The resolution of the data set was limited to 1.3 Å because the average intensity of each reflection in this range had fallen to just over twice the mean standard deviation of the measurements ($I/\sigma I = \sim 2.0$). Hence, the results for Az-PHM crystal H6, given in Table 3.1, reflect statistics for the last resolution shell down to 1.3 Å as well as the entire data set.

Table 3.1. Data collection statistics for the reduction of reflection data from Az-PHM crystal H6 in CrystalClear 2.1. Statistics are given for the entire data set and for the last resolution shell.

	All Resolution Shells	Last Resolution Shell
Space Group	P2 ₁ 2 ₁ 2 ₁	
Unit Cell Dimensions (Å, °)	$a = 50.2, b = 54.9, c = 85.9$ $\alpha = \beta = \gamma = 90$	
Lattice System	Orthorhombic	
Laue Symmetry	mmm	
Mosaicity (°)	0.61	
Total number of reflections	252,950	
Number of unique reflections	52,747	
Resolution Range (Å)	28.05 – 1.30	1.35 – 1.30
Average Redundancy	4.80	2.72
Completeness (%)	89.3	70.7
R_{merge}	0.043	0.356
R_{meas}	0.047	0.437
Output $\langle I/\sigma I \rangle$	15.2	2.3

Completeness is defined as the number of unique reflections measured over the total number of unique reflections possible (possible unique reflections are determined by the symmetry of the crystal) and though it is somewhat lacking at 70.7% for the last resolution shell, the presence of spots was significant enough to warrant inclusion in the data set since $\langle I/\sigma I \rangle$ fell to approximately 2. In Table 3.2, completeness statistics are given by resolution shells. Out of the total number of observed reflections in the last resolution shell, less than 1% of them were rejected, indicating that the spots measured in this shell should not be disregarded. However, including the last resolution shell in the data set may be frowned upon by some since shell completeness is often in the 95th

percentile (39). In hindsight, it may have been advantageous to collect the data using a lower symmetry during strategy to ensure more images were taken and that all unique reflections in this last shell were collected, which would boost completeness not only for the last shell, but also for the entire data set.

Table 3.2. Data collection statistics after the reduction of reflection data from Az-PHM crystal H6 in CrystalClear 2.1. Statistics given are completeness results for the last resolution shell, down to 1.3 Å.

Completeness results for the Last Resolution Shell	
Resolution range	1.35 – 1.30
# of observed rflns	11279
# of rejected rflns	70
Calculated unique rflns	5824
# of unique rflns	4118
# of unique rflns measured once	506
# of unique rflns measured multiple times	3612

In discussing the quality of the data in terms of intensity, it is important to remember that symmetry-equivalent reflections are often measured multiple times and should theoretically have equal intensities. R_{merge} is a statistic that conveys the quality of the intensity measurements made for symmetry-equivalent reflections. It is described as the sum of the absolute differences for all symmetry-related measurements of a particular unique reflection, where the sum of differences from every unique reflection is added together and divided by the sum of the average intensity values of all unique reflections. Though this value is typically reported in crystallographic analysis, it seems to be a poor statistic to use to determine the quality of the intensity data because it is affected by redundancy, a ratio of the total number of intensity measurements to the total number of unique reflections measured (39). If redundancy is high, then R_{merge} will also be high. The statistic R_{meas} is identical to R_{merge} but is independent of redundancy and therefore, a better measure of the quality of intensity measurements (41). The R_{meas} statistic is calculated as follows in CrystalClear,

$$R_{meas} = \frac{\sum_h \sqrt{N/N-1} \sum_i |I_{hi} - \bar{I}_h|}{\sum_h \sum_i \bar{I}_h},$$

where N is the number of observations for unique reflection h , I_{hi} is the i^{th} measurement for unique reflection h , and \bar{I}_h is the average intensity for unique reflection h . R_{meas} for the entire data set is excellent at a value of 0.047, but R_{meas} values increase exponentially until in the outer shell, the error in intensity measurements averages more than 40% of the observed values at 0.437. This is a sensible R_{meas} value because as the diffraction angle gets larger, spots at a higher resolution are more affected by crystal disorder and tend to give more varied intensities (18). Recently, Karplus and Diederichs stated that data should be truncated before $\langle I/\sigma I \rangle$ reaches 2.0 and before R_{merge} or R_{meas} exceeds ~0.6 to 0.8 (42). So, even though R_{meas} is 0.437 for the outer shell, it is well below the R_{meas} cutoff and is another indication that the diffraction data in the last shell should be included in structure determination.

As previously mentioned, redundancy is a statistic describing the number of times a unique reflection is observed. When unique reflections in a data set have high redundancy (or multiplicity), it becomes easy to detect outliers. Detecting outliers accurately also depends on the correct assignment of intensities during integration, making it imperative that detector distances and beam location are properly placed (41). Because the beam location was refined and the average redundancy for most of the resolution shells is moderately high at values between 5 and 8 (see Table 3.3), the scaling and merging algorithm should have had no trouble in detecting outliers. That being said, when considering R_{merge} values no outliers were detected in the data and all frames received the same scale factor, which suggests that the data collected from crystal H6 is high-quality and that the crystal itself suffered no radiation damage during data collection to cause any significant differences in reflection intensities.

Table 3.3. Data collection statistics for the reduction of reflection data from Az-PHM crystal H6 in CrystalClear 2.1. Statistics give redundancy in resolution shells.

Resolution range	Calculated unique rflns	Percent of reflections measured N times, N = (%)						Completeness for shell (%)
		0	1	2	3	4	5-8	
28.05 – 2.80	6213	0.0	0.5	3.2	9.4	9.7	77.2	100.0
2.80 – 2.22	5987	0.2	2.1	3.0	12.3	4.4	78.0	99.8
2.22 – 1.94	5934	0.2	2.4	6.5	11.0	6.8	73.3	99.8
1.94 – 1.76	5873	1.4	2.3	8.2	12.1	13.7	62.4	98.6
1.76 – 1.64	5893	4.5	2.3	9.7	11.5	19.9	52.1	95.5
1.64 – 1.54	5860	7.2	2.1	10.9	9.9	23.8	46.1	92.8
1.54 – 1.46	5852	14.7	3.5	14.6	7.2	24.1	35.8	85.3
1.46 – 1.40	5818	23.2	3.9	23.3	6.9	25.4	17.3	93.7
1.40 – 1.35	5832	28.0	4.8	29.5	4.1	29.5	4.1	72.0
1.35 – 1.30	5824	29.3	8.7	30.8	2.7	28.5	0.0	70.7

Even in keeping the data in the last resolution shell, the statistics for the entire data set are great and promise a high-quality crystal structure. Characteristics of the diffraction, such as mosaicity, suggest that Az-PHM crystal H6 is a well-ordered crystal and the strength of the reflections throughout the data set imply that it is highly probably that the correct Laue class and space group have been chosen. Further confirmation of the excellent quality of the diffraction data will result after estimating phases during molecular replacement and viewing the electron density map.

Chapter 3.3 Intensities to Structure Factors and Molecular Replacement in CCP4

Because the data for Az-PHM crystal H6 was reduced using Rigaku's CrystalClear 2.1 software, it was introduced to the software program CCP4 6.1.3 in the form of a unique list of scaled reflection intensities. Figure 3.13 displays a small portion of the reduced intensity output data from CrystalClear. Each line of the output file includes information for a unique reflection. The location of the reflection is specified by the values for h, k and l (columns 1-3), and the averaged and scaled intensity value for each reflection is also given along with its standard deviation, which are denoted "fIntensity" and "fSigmaI", respectively (columns 4 and 5).

```

nH
nK
nL
fIntensity
fSigmaI
  0    0    4 27753.9 883.621
  0    0    6 5314.16 286.605
  0    0    8 2454.38 190.803
  0    0   10 20753.0 641.960
  0    0   12 275712.0 4416.13
  0    0   14 56.3007 18.2364
  0    0   16 50814.5 1822.22
  0    0   18 27042.6 10548.2
  0    0   20 420541.0 9527.92

```

Figure 3.13. Output file from CrystalClear 2.1 containing a portion of scaled and averaged intensity data for each unique reflection measured in the Az-PHM H6 data set.

The next step used the Import Merged Data function in CCP4 to convert these intensities from CrystalClear to structure factor amplitudes. Of all the parameters used to accomplish this task, shown in Figure 3.14, only a few needed to be modified from their initial default values. The first modification involved *converting scaled data output from d*trek into MTZ format*, a file form that CCP4 reads. Next, the *Use anomalous data* function was deselected since the integrated data, *ScalAveraged3.ref*, contains no anomalous data. To convert intensities to structure factors, the *old Truncate* algorithm was run using the *French&Wilson truncate* protocol (43). While the new structure factors would be written to the output file, *ScalAveraged3.mtz*, the option to *Keep the input intensities in the output MTZ file* was selected though this data was no longer required after conversion. Also, even though the data in *ScalAveraged3.ref* contains a unique list of scaled reflection intensities, it was important to choose to *Ensure unique data & add a FreeR column* to 5% of the data. By flagging a small portion of the structure factors, the data could be used in cross-validation of the model in later steps. Lastly, extra information about the crystal, data set, unit cell dimensions, protein length, space group and radiation wavelength was entered for inclusion in the final MTZ output file before running the Import Merged Data function.

ImportScaled - Import Scaled Data from Denzo or d*trek Initial parameters from E:/CCP4/phm17lta... Help

Job title **Import Merged Data**

Convert scaled data output from **d*trek** into MTZ format

Use anomalous data

Run **old Truncate** to convert intensities to structure factors

Keep the input intensities in the output MTZ file

Ensure unique data & add FreeR column for **0.05** fraction of data. Copy FreeR from another MTZ

Extend reflections to higher resolution:

In **AzPHM_best** **ScalAveraged3.ref** Browse View

Out **AzPHM_best** **ScalAveraged3.mtz** Browse View

Use **dataset name** as identifier to append to column labels

MTZ Project, Crystal, Dataset Names & Data Harvesting

Create harvest file in project harvesting directory

Crystal **H6** belonging to Project **AzPHM_best**

Dataset name **AzPHM**

Extra Information for MTZ File

Space group **19**

Cell dimensions **50.1778** **54.8946** **85.9189** **90.0** **90.0** **90.0**

Data collected at wavelength **1.5419** Angstroms

Estimated number of residues in the asymmetric unit **128**

Edit or Transform Input Data

Set resolution limits to min and max

Scale input intensities by **1.0**

Exclude the following reflections from the output file

Log File Output

Monitor every **10** reflection

Conversion to SFs Parameters

Use **French&Wilson truncate** protocol to generate SFs

Resolution ranges **set by program**

Exclude from Wilson scaling reflections resolution less than **4.0** or greater than

Set volume per atom to **10.0**

Figure 3.14. Import Merged Data parameters used in converting crystal H6's scaled and averaged intensity data into structure factors using CCP4.

A small portion of the resulting MTZ file from the Import Merged Data function is displayed in Figure 3.15. Like the output file ScalAveraged3.ref from CrystalClear, the MTZ file from CCP4 contains reflection locations h, k and l in columns 1, 2 and 3, respectively. Column 4 contains the FreeR flags that mark each reflection depending on the percentage of data withheld from model building. Structure factor amplitudes and their standard deviation are listed in columns 5 and 6, respectively, for each unique reflections from the H6 data set. Recall from Figure 3.14 that structure factor amplitudes were generated using the *French&Wilson truncate* protocol. This algorithm created by French and Wilson (43) is a complex algorithm based on Bayesian statistics that calculates a multiplier by which the data is scaled after the square root is taken. The other option in CCP4, simple square-root, is a straightforward approach that only involves taking the square root of the intensities. For this data, the *French&Wilson* protocol determined that all intensities would be multiplied by 1.569 after taking the square root. So, the structure factor amplitude located at 0, 0, 4 is 261.35 (column 5 in Figure 3.15), which was derived from its intensity of 27753.9 (column 4 in Figure 3.13) by rooting the intensity and then multiplying it by 1.569.

```

LIST OF REFLECTIONS
=====

```

									column
									label
0	0	2	2.00	?	?	?	?		
0	0	4	19.00	261.35	4.17	683.64	21.77		
0	0	6	17.00	114.28	3.09	130.90	7.06	H	H
0	0	8	8.00	77.57	3.04	60.46	4.70	H	K
0	0	10	9.00	226.00	3.50	511.19	15.81	H	L
0	0	12	11.00	823.73	6.60	6791.41	108.78	I	FreeR_flag
0	0	14	18.00	11.15	2.27	1.39	0.45	F	F_AzPHM
0	0	16	4.00	353.51	6.35	1251.68	44.89	Q	SIGF_AzPHM
0	0	18	11.00	231.37	64.94	666.12	259.83	J	IMEAN_AzPHM
0	0	20	13.00	1016.93	11.54	10358.88	234.69	Q	SIGIMEAN_AzPHM

Figure 3.15. Output file from CCP4 containing structure factor amplitudes for each unique reflection measured in the Az-PHM H6 data set.

Upon obtaining structure factor amplitudes, the data was analyzed for solvent content in preparation for molecular replacement. Solvent content is related to the Matthews Coefficient, which is calculated using the cell volume and molecular weight of the crystallized protein. For H6, these parameters were entered in the Cell Content Analysis window seen in Figure 3.16. In the second line of the window, *Calculation of the Matthews coefficient* was for protein only, seeing as H6 did not contain any nucleic

acids. The input *MTZ file*, ScalAveraged3.mtz, containing the structure factor amplitudes was used and because the *Read crystal parameters from MTZ file* option was selected, *Space group*, *Cell dimensions*, and *High resolution limit* were filled in automatically. Lastly, the *Molecular weight of the protein* was 14000, entered in Daltons.

After running the algorithm, the Solvent Content Analysis window shows a table of results (displayed in Figure 3.16) that include the Matthews Coefficient and the corresponding solvent content in percent for a given number of molecules per asymmetric unit (Nmol/asym) along with probabilities indicating the likelihood of each solution. It is clear that there are likely 2 Az-PHM molecules in each unit cell suggested by this solution's the total probability value of 96% and solvent content of 41.83%. Solvent content in protein crystals is typically around 43% and in rare cases has been as low as 27% or as high as 65% (44).

Matthews - Cell Content Analysis Initial parameters from E:/CCP4/phm17ltave/CCP4_DATABASE/3_...

Job title: Calculation for protein only using molecular weight 14000 Daltons

Calculate Matthews coefficient for: protein only

Read crystal parameters from MTZ file

MTZ file: AzPHM_best ScalAveraged3.mtz

Space group: P 21 21 21

Cell a: 50.1778 b: 54.8946 c: 85.9189 alpha: 90.0000 beta: 90.0000 gamma: 90.0000

High resolution limit: 1.300

Use molecular weight: entered in Daltons

Molecular weight of protein or nucleic acid: 14000

Solvent content analysis

Nmol/asym	Matthews Coeff	%solvent	P(1.30)	P(tot)
1	4.23	70.91	0.00	0.03
2	2.11	41.83	0.99	0.96
3	1.41	12.74	0.00	0.00

Cell volume: 236662.766

For given protein molecular weight: 14000 Da

Buttons: Reset, Run Now, Close

Figure 3.16. Cell Content Analysis window in CCP4 containing parameters for the calculation of the matthews coefficient for Az-PHM crystal H6.

The last step of preparation before molecular replacement involves calculation of B-factors which represent thermal disorder and the amount of smearing of electron density due to thermal motion and crystal disorder. The software algorithms BAVERAGE and WILSON accessed through the Analyse Data for MR window, shown in Figure 3.17, calculated B-values for the structure factor amplitudes in *ScalAveraged3.mtz* and for the molecular replacement model *4AZUone.pdb* in order to *Do B value analysis of the experimental data and model structure*. The model in *4AZUone.pdb* is one monomer (chain A) of the structure solved by Nar *et al.* in 1991 and was deposited in the Protein Data Bank with the ID: 4AZU (16). The last parameter entered was the *Number of residues in the asymmetric unit*. This value depends on the results from the solvent content analysis performed previously. Recall that the number of molecules in the asymmetric unit was predicted to be 2. Because Az-PHM is 128 amino acids long, the number of residues in the asymmetric unit is likely 256, which is the value that was entered for this parameter.

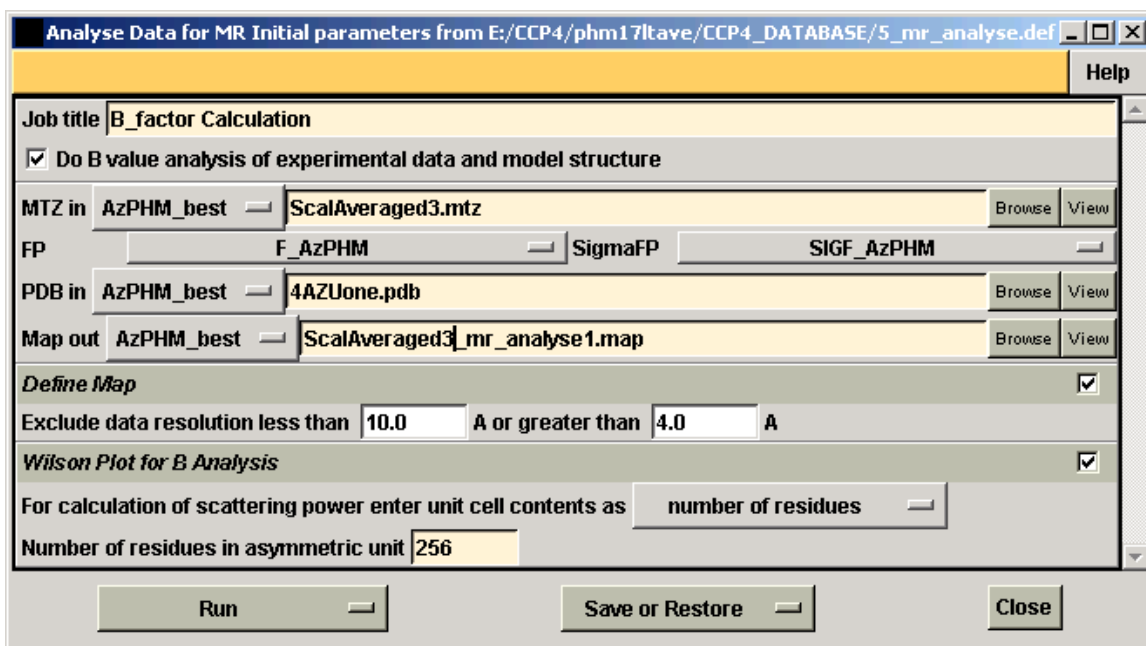


Figure 3.17. Analyse Data for MR window in CCP4 which contains all of the parameters used in calculating B-factors for the experimental structure factor data from Az-PHM crystal H6 and for the accompanying molecular replacement model.

Upon running the analysis software, there were three noteworthy values that were calculated which include B-factors for the experimental data and the molecular replacement model along with a BADD parameter. The B-factor reported for the model (4AZUone.pdb) was 16.656, and for the experimental data it was 13.655. Because the BADD parameter is simply the difference between the two B-values, it follows that its value was -3.001. A low experimental B-factor ensures that the quality of the electron density is preserved (39).

After obtaining the number of molecules per asymmetric unit and the magnitude of the BADD parameter, molecular replacement was initiated using these values. The Molrep – auto MR window in CCP4 (Figure 3.18) includes the values for all the parameters used. Molecular replacement was done *performing rotation and translation functions* to fit the *4AZUone.pdb model* to the experimental structure factor data in *ScalAveraged3.mtz*. All data out to a *maximum resolution* of 1.3 Å was used for fitting. Because Az-PHM has a 3-residue difference from native azurin, there were 3 places in the experimental data where the phases would not be estimated properly. In order to assign the correct name of these three residues to the output file 4AZUone_molrep1.pdb, the option to *Use sequence* was selected and the appropriate sequence file, *AzPHM.seq*, was entered. The file contains one chain of 4AZU's fasta sequence from the Protein Data Bank with proper modifications. All other parameters not discussed are default.

Molrep - Molecular Replacement Initial parameters from E:/CCP4/phm17ltave/CCP4_DATABASE/6_molrep.def

This interface is for version 9.2 of Molrep

Job title Molecular Replacment

Do molecular replacement performing rotation and translation function

Get input structure factors from MTZ file

Input fixed model

Multi-copy search

Use sequence

MTZ in AzPHM_best ScalAveraged3.mtz Browse View

Use Intensities

FP F_AzPHM SIGFP SIGF_AzPHM

Model in AzPHM_best 4AZUone.pdb Browse View

Coords out AzPHM_best 4AZUone_molrep1.pdb Browse View

Experimental Data (Resolution, ANISO, DIFF, BADD, INVER, DSCALE, ...)

Use data to maximum resolution 1.3

minimum resolution

Use default scaling

Apply additional Boverall factor (Badd) -3.001

The Model (SIM, COMPL, SURF, NMR, NCSM, DSCALEM, ...)

Apply set Bvalues related to accessibility & shift to origin to model

If input PDB is for NMR models then use all models in file as single model

Expect fraction completeness of model with fraction similarity to input structure

number of subunits in the model NCSM 0 only for scoring (default from model Self RF)

Model is the map

Search Parameters (NMON, NP, NPT, PST, STICK, LOCK, ...)

Search for 2 monomers in the asymmetric unit

Locked Rotation Function

Use Self Rotation function with 0 peaks from the self-rotation function

Self RF solutions AzPHM_best Browse View

Search for 0 peaks in rotation map and 0 in translation function

Auto pseudo-translation vector

Output the closest of symmetry-equivalent monomers to the coordinates file

Parameter for SEQ

Seq in AzPHM_best AzPHM.seq Browse View

Infrequently Used Parameters (MODE, SAPTF, RAD, PACK, SCORE, LMIN, NOSG)

Change space group Do not change

Use spherically averaged phased translation function with phased rotation function

Use standard RF and TF without rigid body refinement ('fast' mode)

Search radius

Use packing function with translation function

turn off scoring system

number of cycles of RB refinement after TF before TF

Nptd

Lmin

Run Save or Restore Close

Figure 3.18. Molrep – auto MR window containing all parameters used to complete molecular replacment for the experimental data of Az-PHM crystal H6 using phase information from the 4AZU model.

Before viewing the Az-PHM model resulting from molecular replacement, the coordinates were first refined using CCP4's Refmac5 function. During refinement, the position of the atoms in the model structure is refined while accounting for geometric restraints and other restrictions specified by the presence (or absence) of input parameters. These parameters, visible in Figures 3.19 and 3.20, are numerous, so only those that were modified from their default values will be discussed. The first (shown on the second line of the refinement window) gives several options for refinement, and the type of refinement selected depends largely on the resolution of the data. It is common to use restrained refinement following molecular replacement because atoms are refined according to the electron density while using geometric restraints. Another option, unrestrained refinement, does not include geometric restraints (only used in cases of very high resolution) while rigid body refinement only refines the position of the entire asymmetric unit without refining individual atoms (used in cases of low resolution). Because the Az-PHM data is fairly high resolution and had a low thermal B-factor, *TLS & restrained refinement* was performed using *no prior phase information*. TLS, which stands for translation libration and screw-motion, is a type of refinement that takes vibrational motion into account when refining atomic positions. After the type of refinement was selected, the structure factor amplitudes (found in *ScalAveraged3.mtz*) and the Az-PHM model (found in *4AZUone_molrepl.pdb*) were entered after which names for all of the output files were automatically assigned. In the Refinement Parameters section, it was important to *Exclude data with a freer label, FreeR_flag, with a value of 0* in order to set aside 5% of the data for calculation of R_{free} , a statistical value that measures how accurately the model predicts the experimental density. This small fraction of data is also referred to as the "free" set. Looking under the Geometric Restraints section, notice that the option to *make links between all others* has "defined in file only" following it. Another option for this parameter is to select "defined in file or residues are close" and is advantageous to choose if the data are low resolution and there are certain bonds that should be maintained. Non-crystallographic Symmetry (NCS) Restraints were not used in the refinement of H6 data, even though the data suggested that it contained more than one monomer in its asymmetric unit. This is because the

resolution of the data is high, such that NCS restraints were not needed and may in fact be harmful during refinement. The only parameter modified in Figure 3.20 was the type of scaling used, which was changed from *simple* to *Babinet*. This parameter was changed in order to take bulk solvent into account. All other parameters were unchanged.

Run Refmac5 Initial parameters from E:/CCP4/phm17ltave/CCP4_DATABASE/10_refmac5.def

Help

Job title Initial Refinement

Do TLS & restrained refinement using no prior phase information input

no twin refinement

MTZ in AzPHM_best ScalAveraged3.mtz Browse View

FP F_AzPHM Sigma SIGF_AzPHM

MTZ out AzPHM_best ScalAveraged3_refmac3.mtz Browse View

PDB in AzPHM_best 4AZUone_molrep1.pdb Browse View

PDB out AzPHM_best 4AZUone_molrep1_refmac3.pdb Browse View

LIB in AzPHM_best Merge LIBINS Browse View

Output lib AzPHM_best 4AZUone_molrep1.cif Browse View

TLS in (optional) AzPHM_best Create TLSIN Browse View

TLS out AzPHM_best 4AZUone_molrep1_refmac1.tls Browse View

Include keyword file AzPHM_best Browse View

Data Harvesting

Create harvest file in project harvesting directory

Harvest project name AzPHM_best and dataset name AzPHM

TLS Parameters

Number of cycles of TLS refinement 10

Set initial Bfactors to 20.0 (numeric value unimportant)

Add TLS contribution to XYZOUT (B factors and ANISOU lines)

Refinement Parameters

Do 10 cycles of maximum likelihood restrained refinement after TLS refinement

Use hydrogen atoms: use if present in file and output to coordinate file

Resolution range from minimum 46.259 to 1.300

Use automatic weighting Use experimental sigmas to weight Xray terms

Refine isotropic temperature factors

Exclude data with freeR label FreeR_flag with value of 0

Use the free set of reflections for fitting the SigmaA estimate

Setup Geometric Restraints

Checking against dictionary: Rely on user's naming convention

If the following features are found in coordinate file then make restraints to maintain them:

D-peptide cis-peptide Links between symmetry related atoms

Make links between:

Amino acids and DNA/RNA if defined in file only

Sugar-sugar and sugar-peptide if defined in file or residues are close

All others if defined in file only

Setup Non-Crystallographic Symmetry (NCS) Restraints

No NCS restraints are currently defined

Edit list Add NCS restraint

Figure 3.19. Refmac5 parameters used for running the initial round of refinement for Az-PHM crystal H6 data following molecular replacement in CCP4.

Monitoring and Output Options

Output **medium** monitoring statistics

Sigma cutoffs for monitoring levels..

Torsion	Distance	Angle	Planarity	vanDerWaals
10.0	10.0	10.0	10.0	3.0
Chiral	Bfactor	Bsphere	Rbond	NCSr
10.0	10.0	10.0	10.0	10.0

Fcalc	FC	PHICalc	PHIC
mFoDFc	DELFWT	PhimFoDFc	PHDELWT
2mFoDFc	FWT	Phi2mFoDFc	PHWT
FOM	FOM		

Generate weighted difference maps files in **CCP4** format

Scaling

Use **Babinet** scaling

Bulk solvent scaling for resolution range to

Determine scaling using the **working** set of reflections Use experimental sigmas

Calculate the contribution from the solvent region

For the solvent mask calculation:

Increase VDW radius of non-ion atoms by

Increase ionic radius of potential ion atoms by

Shrink the area of the mask by after calculation

For low resolution structures, fix the B values of Babinet's bulk solvent to

Geometric parameters

Restraint	Overall wt	Sigmas			
<input type="checkbox"/> Distance	1.0				
<input type="checkbox"/> Angle	1.0				
<input type="checkbox"/> Bfactor	1.0	main chain bond	main chain angle	side chain bond	side chain angle
<input type="checkbox"/> Plane	1.0	peptide	aromatic		
<input type="checkbox"/> Chiral	1.0	chiral			
<input type="checkbox"/> Torsion	1.0				
<input type="checkbox"/> NCS position	1.0	tight	medium	loose	
NCS Bfactor					
<input type="checkbox"/> VDW contacts	1.0				

Sigmas for types of non-bonding interaction..

Non-bonding H-bonding metal-ion interactions 1-4 atoms in torsion

Set increments for non-bonded interactions..

1-4 atoms in torsion H-bond pair (not hydrogen atom) H-bonded pair (one is hydrogen atom)

Set limits for B values..

Limit B value range from 2.0 to 200.0

Run **Save or Restore** **Close**

Figure 3.20. Refmac5 parameters used for running the initial round of refinement for Az-PHM crystal H6 data following molecular replacment in CCP4.

After 10 cycles of restrained refinement followed by 10 cycles of TLS refinement, R_{factor} and R_{free} are displayed. These R-values play a significant role in determining correct atomic positions in the model because they convey the accuracy of the fit of the Az-PHM model (*4AZUone_molrep1_refmac3.pdb*) to the experimental electron density map (*ScalAveraged3_refmac3.mtz*). R_{factor} is a ratio of the sum of differences between observed and calculated structure factor amplitudes to the sum of observed structure factor amplitudes (39). R_{free} is a similar unbiased statistic that uses a small portion of the reflection data (5% in this case) for “observed structure factor amplitudes” which are excluded during refinement. R_{free} was originally developed after Axel Brünger (45) noticed a bias in R_{factor} that results from using the same reflections for both model building and refinement. After the initial round of refinement for the H6 model and map, Refmac gave an R_{factor} of 32.45% and an R_{free} value of 34.55%. At present, these values are moderately-low for this stage of refinement and hold promise of a well-fitting structure.

At this stage of structure determination, confirmation of the accuracy of the results from much of the work completed in CrystalClear and CCP4 is lacking. However, by simply viewing the preliminary model and corresponding electron density map and evaluating the sheer interpretability of the map and the overall fit of the model, the success of data reduction and molecular replacement be determined. This is accomplished through the use of the macromolecular model-building program, Coot. In this software, an electron density map is created using the coupled structure factor amplitudes and phases that were written into the MTZ file during refinement carried out by Refmac. Coot also overlays the model which can be analyzed for overall fit to the map. In Figure 3.21, a photo containing the map and model leaves no doubt that the electron density surrounds the preliminary model well. This simple observation confirms a great deal: intensities were scaled and merged appropriately, structure factor amplitudes were calculated correctly and the phase estimation in molecular replacement was performed successfully.

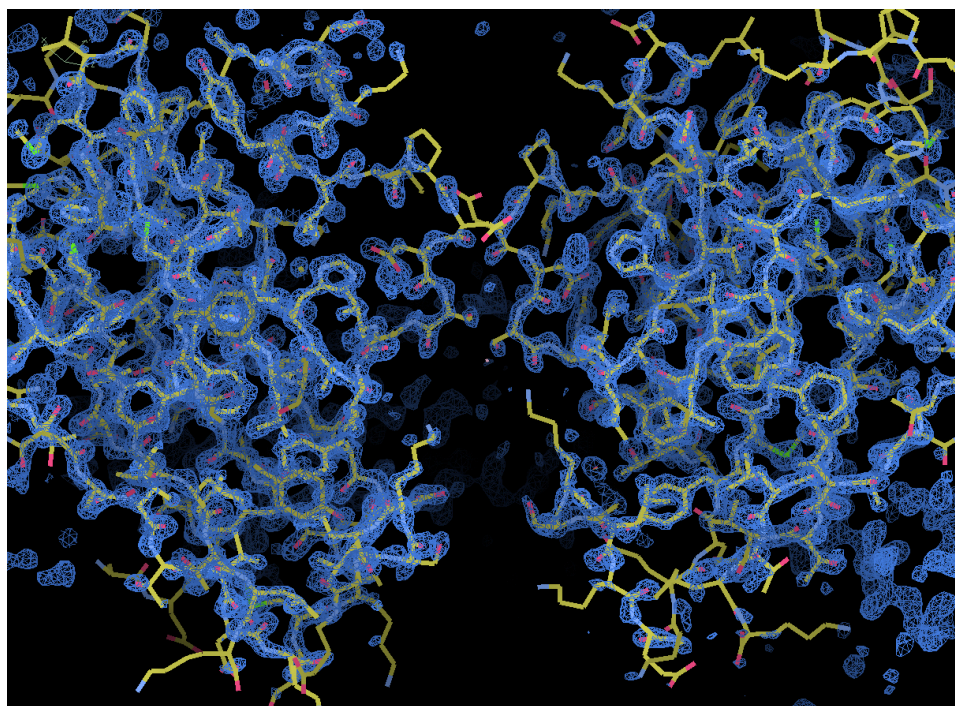


Figure 3.21. Preliminary map and model of Az-PHM resulting from molecular replacement in CCP4.

The second most noticeable feature in Figure 3.21 is the number of molecules in the asymmetric unit. Recall that the solvent content analysis predicted two molecules per unit cell which prompted the MOLREP algorithm to search for two monomers during molecular replacement. In viewing the preliminary structure, the prediction is confirmed because the experimental electron density outlines two distinct Az-PHM molecules that have opposite spatial orientations. The overall fit of the model to the map also validates the choice of Laue class and space group assigned in CrystalClear during data reduction (39).

Upon closer inspection of the structure, electron density surrounds the model equally on all sides throughout a large majority of the structure. A small detailed portion of the structure seen in Figure 3.22 illustrates this point. The high 1.3 Å resolution of the experimental data allows individual atoms in the electron density to be visible, shown most clearly by the phenylalanine residue.

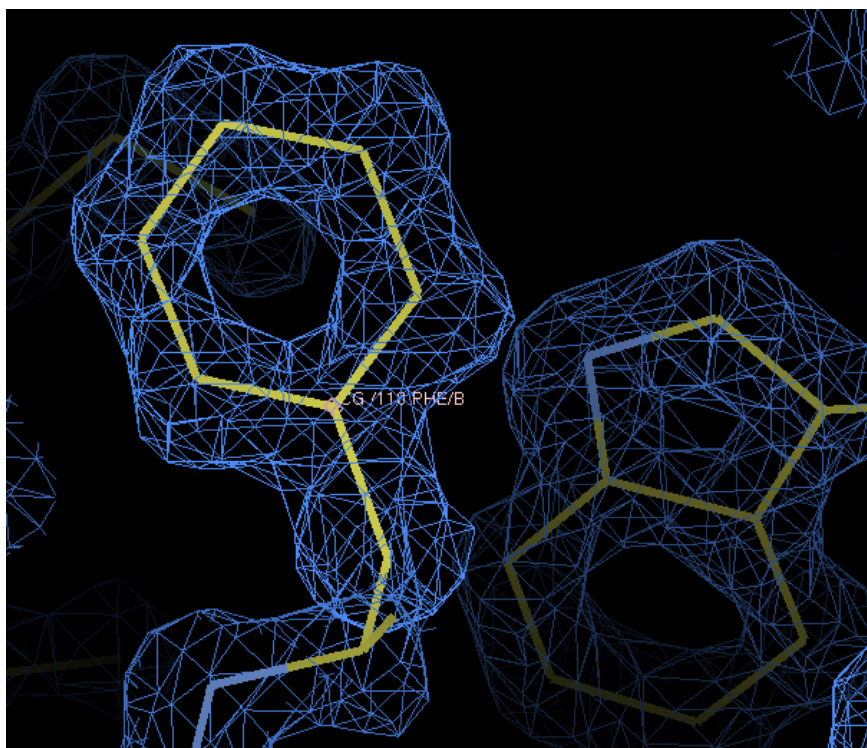


Figure 3.22. Small section of the electron density map and preliminary model from the Az-PHM crystal H6. The electron density map clearly shows individual spheres in the phenylalanine ring.

In looking at a different portion of the structure, seen in Figure 3.23, the identity of the protein is partially confirmed. In this figure, the amino acid in position 14 shows clear electron density in the shape of a histidine residue. Because a crystal structure for wild-type azurin was used for phase estimation in molecular replacement, the model contains a missing part of the histidine residue in position 14. When positions 8 and 16 were inspected, the model showed missing atoms as well. The map showed methionine and histidine density, respectively, confirming that the appropriate mutations were made when creating the Az-PHM protein.

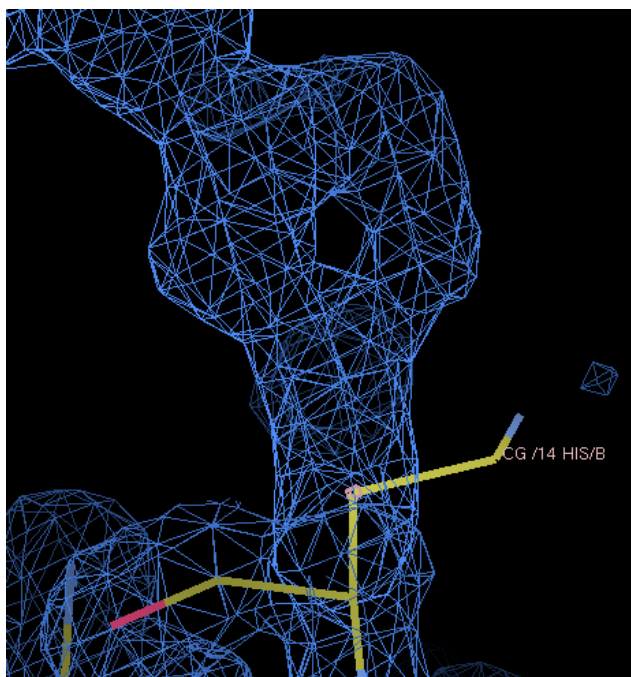


Figure 3.23. Small section of the electron density map and preliminary model from the Az-PHM crystal H6. The electron density map clearly shows electron density for a histidine residue at location 14.

The incorrectly modeled residue can be changed at these three locations within the Coot program to reveal a better model that more accurately describes the true packing of the protein molecules within the crystal. Even without the changes, the overall fit of the preliminary model to the experimentally-determined electron density supports the process and the parameters used in CrystalClear and CCP4 to collect data, reduce it and estimate phases via molecular replacement. The detailed nature of the electron density map shows the high quality of the data obtained and ultimately, the highly-ordered packing of the Az-PHM crystal that was grown.

CHAPTER 4

Chapter 4.1 Introduction

When a preliminary protein model is generated from molecular replacement and initial refinement in CCP4, it is uploaded to a molecular graphics program and compared to the electron density map. Coot (46) is one such program that allows macromolecular crystallographers to easily view their preliminary model against the experimental map and make changes to the model's amino acid sequence, residue conformations and surrounding solvent molecules. Adjustments like these create a more accurate model that better explains the experimental electron density data.

Refining a model in Coot is a tedious process that involves hours of analysis. Changes in the atomic coordinates of the model are typically guided by a difference map, which is created alongside the electron density map. A difference map consists of colored sections of density that indicate areas of positive and negative density that should be modeled and un-modeled, respectively. A crystallographer uses a difference map as an aid in deciding whether an amino acid should be moved or where a water molecule should be added. Scanning the entire difference map can be an arduous task and after a series of adjustments are made to the structure in Coot, the resulting coordinates are input into CCP4 where new coordinates and new phases are calculated upon running a re-refinement. Because the new map is slightly different than the old map, the new coordinates must again be compared to the density and adjusted further if needed. If coordinates are again changed, the model must undergo another bout of refinement in CCP4. This tedious process is known as iterative refinement.

After all residues in the model are properly positioned and iteratively refined, solvent molecules are added to the remaining electron density where appropriate. During the process of adding solvent, the model is again iteratively refined in CCP4 to monitor the effect of the changes to the model. After each round of refinement in CCP4, R-values are monitored (both R_{factor} and R_{free}). If either of the R-values increase after adjustments are made, these changes must be questioned and most-likely discarded if the experimental density does not show a marked improvement.

Changes to the model cease when the fit to the experimental density has been maximized and R-values no longer decrease. R_{factor} and R_{free} are two statistically similar values calculated by CCP4 that guide the process of model refinement by describing the fit of the protein model to the electron density map. R_{factor} is the sum of the differences in observed structure factor amplitudes (F_{obs}) to those calculated from the current model (F_{calc}) all over the sum of observed structure factor amplitudes, which is described by the equation below,

$$R_{\text{factor}} = \frac{\sum_h \left| |F_{\text{obs}}| - |F_{\text{calc}}| \right|}{\sum_h |F_{\text{obs}}|},$$

where h is the set of reflections that is used in creating the map, called the “working” set. This set of reflections is used to create the electron density map and is also used for monitoring the progress of refinement through the calculation of R_{factor} (39). In 1992, Axel Brünger demonstrated the bias associated with using the same set of reflections for refining and monitoring refinement and subsequently, introduced a statistic called R_{free} (45). R_{free} is also known as the “unbiased R_{factor} ” because its calculation is the same as R_{factor} except that the structure factor amplitudes used are from a set of reflections that have been set aside and excluded from the electron density map. This small number of excluded reflections is also known as the “free” set (39).

In setting aside a test set of reflections that was not used to create a protein model, the true predictive power of the final structure can be evaluated. If the model makes accurate predictions of the raw experimental structure factors for the test set, then users of the model can have confidence that it is an accurate depiction of the protein in crystallized form (47). Because R_{free} is one of the only unbiased statistics that judges the model with respect to unused experimental data, it is likely the most powerful validation tool available and therefore, should be given considerable weight in terms of assessing the reliability of the refined model. The Coot software program contains many other tools that validate the model with respect to experimental data, including *Density fit analysis* and *Temperature factor variance analysis* and *NCS differences*.

Coordinate-based methods validate the model by purely assessing the model’s geometry and stereochemistry without considering its fit to the experimental data. Also,

because geometry can be restrained during refinement in CCP4, many of these methods do not provide a true, independent validation of the model, but nonetheless serve to point out potential errors. However, the *Ramachandran plot* is one tool that uses phi and psi torsion angles that are not typically restrained during refinement and as such is a dependable coordinate-based validation tool. Some of the other tools for coordinate validation include side-chain analysis through the *Geometry analysis* function and torsion angle analysis using the *Peptide omega analysis* tool. Stereochemistry is analyzed through the *Incorrect Chiral Volumes* tool while the *GLN and ASN B-factor Outliers* attempts to detect incorrect placements of oxygen and nitrogen atoms in glutamine and asparagine side-chains using temperature factors.

In the sections that follow, the process of model building and refinement of the preliminary Az-PHM model will be discussed. Each of the validation tools mentioned will also be used in determining the quality of the final structure.

Chapter 4.2 Model Building and Refinement in Coot

After performing molecular replacement and initial refinement in CCP4, the Az-PHM model and structure factors were uploaded to the Coot software program where atomic coordinates were adjusted and solvent molecules were fit to un-modeled electron density. Throughout this model building process, only a few changes were made to the model before calculating new phases and thus, a new electron density map in CCP4 (i.e. running the refinement in Refmac5 again). The purpose in refining in this way was to monitor the change in R_{factor} and R_{free} . If any of the individual changes to the model did not produce a decrease in these values, the change was considered inappropriate and unsupported by the experimental data. This careful process was used throughout the Az-PHM model building and refinement process in Coot.

The first changes made to the preliminary model involved residue mutation and copper addition because these structural features were not known by the software to be a part of the Az-PHM protein, because the crystal structure used for molecular replacement only contained one amino acid chain of the wild-type azurin molecule. Copper(II) ions were added in the Type 1 and Type 2 sites of both Az-PHM molecules within the

asymmetric unit using the *Get Monomer* option in Coot, selecting copper ions with the *CU* identifier. Because histidine binds copper strongly, copper(II) ions were also added to the large spherical electron density next to both His83 residues (see Figure 4.1). So, a total of 6 copper atoms were added in the asymmetric unit. Next, electron density in the map supported the expected mutation of residues 8, 14 and 16 in chains A and B to methionine, histidine and histidine, respectively. This was accomplished using the *Mutate & Auto Fit* tool in Coot, where the correct amino acid replaced the old residue and was subsequently fit to the electron density. The new coordinates were then saved and input into CCP4 for TLS & restrained refinement along with the electron density map. The parameters used for refinement with Refmac5 were the same as those shown Figures 3.17 and 3.18 of Chapter 3.3. After refining with Refmac5 in CCP4, both R_{factor} and R_{free} dropped from 32.45% and 34.55% to 27.23% and 30.07%, respectively, indicating that the changes made were indeed warranted. However, after Refmac5 calculated new phases consistent with the newly-refined model, the resulting difference map showed negative density in portions of the copper(II) density, which was most noticeable near the copper(II) ion bound to histidine 83 in chain B (83HisB) as seen in Figure 4.1. Even though negative, red density is present which suggests that too many electrons were placed in that site, it is clear that the copper(II) ion should remain there since it makes chemical sense. There are also many other reasons why some red, negative density may appear in the map. It may be that the density is red because not every protein molecule in the crystal binds copper in this place, leaving a sphere that has less electron density than it theoretically would if all protein molecules bound copper in that space. However a better explanation, and the one that likely applies in this case, comes from paying attention to the locations of the negative density. The red blobs seem to be in a perpendicular direction to the positive, green density where solvent should be bound on either side of the copper(II) ion. In this well known phenomenon, the red density probably appeared in order to compensate for the additional electron density that is localized around the entire copper atom instead of localized around the bonds to the solvent molecules (48).

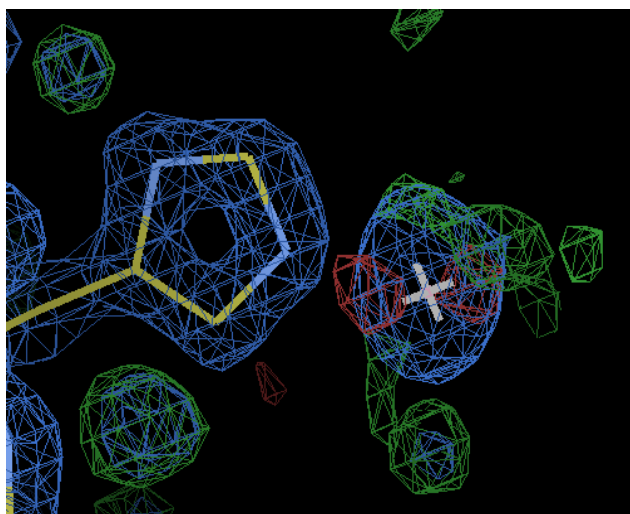


Figure 4.1. Negative, red density on a copper(II) ion next to histidine in chain B, position 83 (83HisB). The negative density seems to be along bonds between 83HisB & copper and between copper and a blob of positive, green density.

Following copper addition and residue mutation, adjustments to individual residue atoms were made with respect to electron density fit and geometry. Residues that did not fit well in the electron density were often identified by a validation feature in Coot called *Density fit analysis*. This interactive tool displayed a color-coded bar graph (Figure 4.2) for each protein monomer, containing a bar for each amino acid residue that described its fit using a score. The score is defined as the average electron density level at the atom centers of the atoms in the residue (49). The farther the score is from 1, the higher the bar appears on the graph. For example, lysine in the 24th position in chain B (24LysB) has a moderately-high, yellow bar with a score of 0.78 (Figure 4.2) that indicates a decent, but not ideal fit to density. Amino acids such as this that had low scores were targeted and adjusted using one of three tools: *Auto Fit Rotamer*, *Rotamers*, or *Real-Space Refine Zone*. The first two options, *Auto Fit Rotamer* and *Rotamers*, fit side-chains to the density automatically or manually, respectively. *Auto Fit Rotamer* automatically chooses the best rotamer from the Molprobity Library developed by Lovell *et al.* (50) that gives the highest electron-density values at the atom centers. *Rotamers*, on the other hand, allows the best side-chain conformation to be selected manually from a generated list, sorted by frequency in the Molprobity database. Lastly, *Real-Space Refine Zone* not only refines atom centers for the highest electron density values but also considers stereochemical restraints when making adjustments to a range of atoms specified by the

user. Before making any adjustments permanent, scores regarding bonds, angles, planes, etc. are displayed that must be acceptable to the user. If desired, the user may move atoms to obtain new geometry scores, before making the adjustments permanent (49).

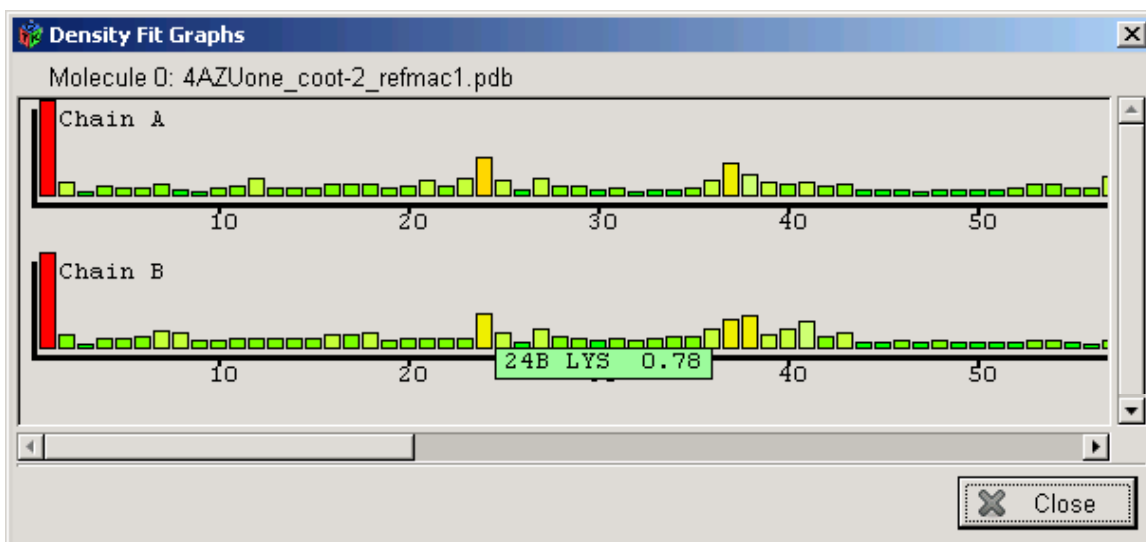


Figure 4.2. Iterative bar graph from the Density fit analysis tool under Coot's validation options. Each bar represents an amino acid in the model and the color and height indicate its fit to the electron density map. chain A and chain B refer to two distinct Az-PHM molecules in the asymmetric unit.

Electron density fit is not the only consideration when making adjustments to atomic coordinates. When residues in a model are adjusted, each residue should fit the density but should also have an acceptable geometry. Like the density tool in Coot, *Geometry analysis* is a validation tool that describes the geometry of the model using a bar graph like the one shown in Figure 4.3. Each bar represents an amino acid in the model, the color and height of which depend on the Z-score of the amino acid in question. Z-values are calculated for bonds, angles and planes by comparing the model to dictionary conformations and are subsequently averaged together, resulting in an overall Z-score. A residue's Z-score is proportional to the height of its corresponding bar (48). For example, the lysine residue at position 24 in chain B has relatively good geometry in terms of its bonds, angles and planes since its bar is green with a moderately-low height due to a Z-score of 5.29. The caption in the graph also indicates that the angle at the 1st carbon of the side-chain (CB) in lysine is the largest contributor to the overall Z-score and should, therefore, be examined more carefully than any bonds or planes.

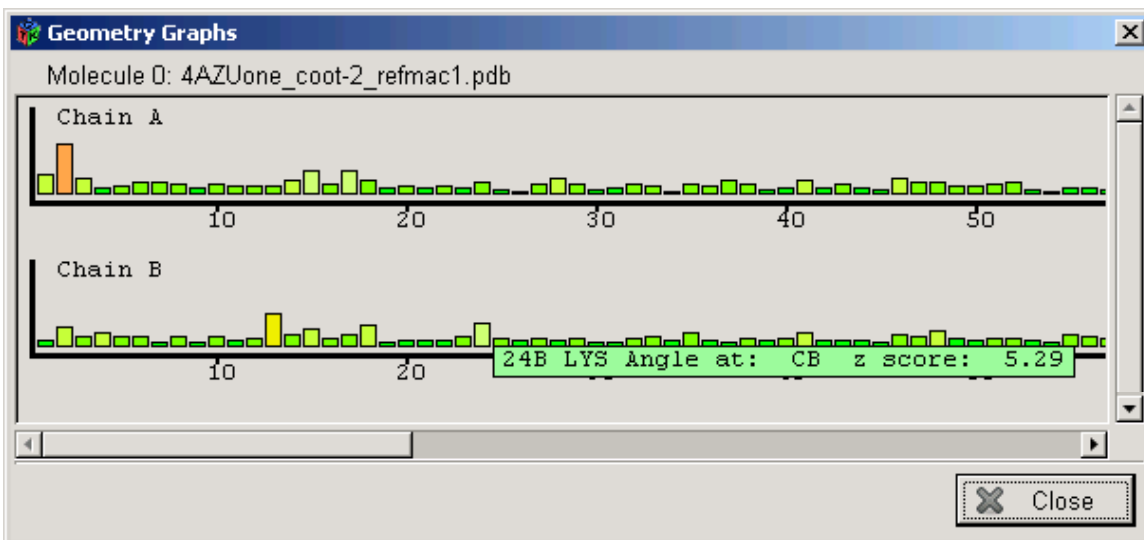


Figure 4.3. Iterative bar graph from the Geometry analysis tool under Coot's validation options. Each bar represents an amino acid in the model and the color and height indicate geometry in terms of angles, bonds, and planes relative to dictionary geometries. chain A and chain B refer to two distinct Az-PHM molecules in the asymmetric unit.

There are two options for addressing geometry in Coot: *Real-Space Refine Zone* and *Regularize Zone*. *Real-Space Refine Zone*, which was previously discussed, refines coordinates while considering density and geometry whereas *Regularize Zone* refines with respect to geometry only. Normally, *Real-Space Refine Zone* was used to refine residues with high Z-scores since *Regularize Zone* frequently moved residues out of their electron density, increasing the R-values after subsequent phase recalculation.

At high resolutions, alternate or split conformations for residues can be modeled if convincing density is located. For example, in Figure 4.4a electron density shows an alternate conformation for sulfur of the methionine in the 8th residue position of chain B (8MetB) which is supported by the negative density surrounding the current position. Another method for identifying density resulting from multiple residue conformations is found in the Coot Extensions menu under Modeling: *Residues with Alt Confs*. After using this function, Coot also identified the density around 8 Met B for potential modeling. Thus, the alternate conformation for 8MetB was modeled with each side-chain occupying 50%, and the new coordinates were then refined in CCP4. Though the post-refinement solution (Figure 4.4b) shows a small amount negative density, a decrease of 0.03% and 0.05% was observed for R_{factor} and R_{free} , respectively. Considering that R_{free} is

a global indicator, it was surprising that such a small local change brought about drop of 0.05%. For this reason and because the negative density on the sulfur disappeared, the alternate coordinates for 8MetB were kept. Even though the *Residues with Alt Confs* function did not find any other density worth investigating, alternate conformations were modeled for 9 other residues, including 6AspA, 57GlnA, 61ThrA, 122LysA, 126ThrA, 4SerB, 27LysB, 34SerB, 126ThrB. Density for these conformations was identified through the use of the *Difference Map Peaks* function in Coot.

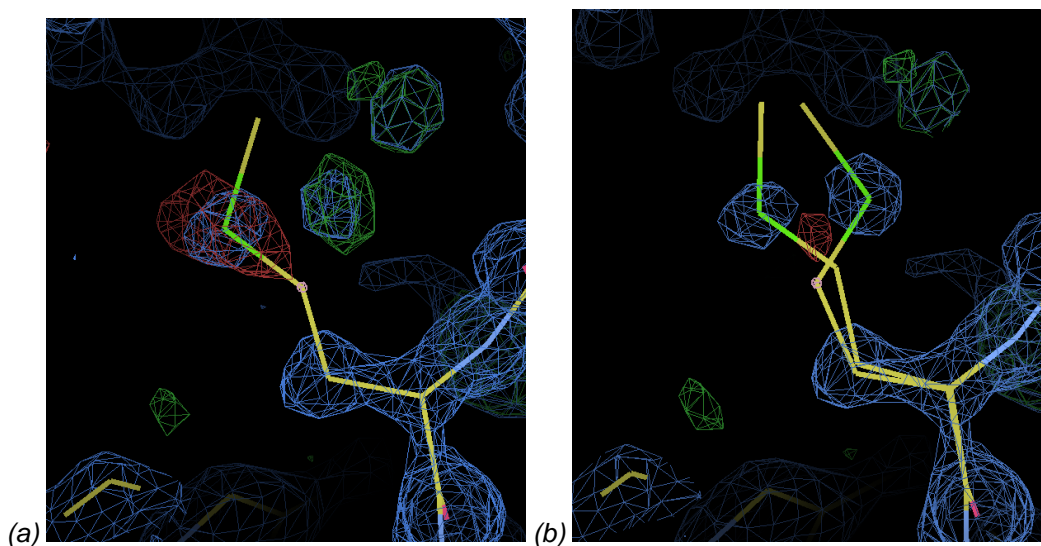


Figure 4.4. (a) Electron density around methionine 8B. This density suggests an alternate conformation should also be modelled. (b) Electron density map and model after an alternate conformation of methionine 8B was modelled.

After all individual amino acid residue adjustments were completed, the next step was to locate large volumes of empty electron density that belong to ligands and other solvent molecules. Using *Difference Map Peaks* was one way to locate these empty areas of electron density. Two other options include the *Find Ligands* function and the validation tool, *Unmodelled blobs*. Because all three functions are slightly different, all three tools were used to place any PEG, nitrate, Tris, calcium and chloride ions that might have been present in the structure. The most sophisticated option, *Find Ligands*, searches the map for a cluster of density and attempts to fit each cluster with ligand(s) specified by the user. This option was employed in the search for solvent molecules in the Az-PHM map and one Tris molecule (identifier: TRS) was located near the copper(II) ion off of 83HisA (Figure 4.5a). After Tris was found and fit to the density, it was merged to the

protein molecule, and before any refinement in CCP4, the Refmac5 parameter *Make links between: All others if* was changed to *defined in file or residues are close*. In changing this parameter, the geometry of the Tris molecule was restrained and any bonding between Tris and the copper was maintained. Following refinement in CCP4, the Tris molecule was kept because both R_{factor} and R_{free} decreased significantly for such a small change (decreased by 0.06% and 0.22%, respectively) and because the electron density map was greatly improved (Figure 4.5b).

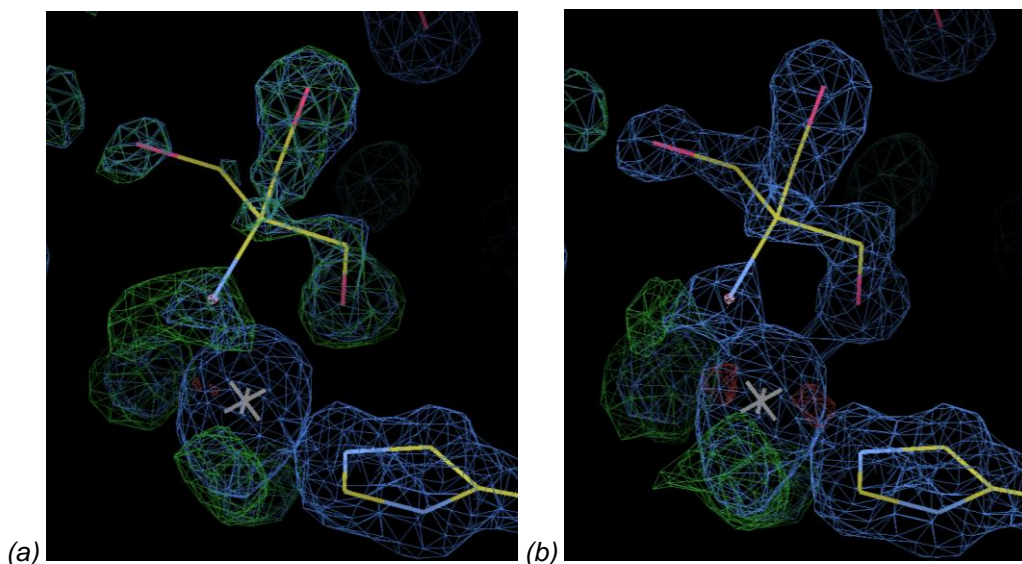


Figure 4.5. (a) Tris molecule fit to positive electron density near 83HisA in Coot. (b) Tris molecule after refinement in CCP4.

In the Az-PHM map, using the *Unmodelled blobs* tool did not reveal any unmodeled electron density, so the *Difference Map Peaks* tool was used. This tool allows the user to locate significant positive and negative peaks (blobs) of density by creating an ordered list according to peak height. Those peaks with the highest deviation from the average are listed first. Using *Difference Map Peaks*, density was located in the Az-PHM map that fit three other Tris molecules and a chloride ion. Chloride ions can be difficult to place in a model because their corresponding density is spherical which can easily be mistaken for a water molecule. The highest peak in the difference map was 21 times the standard deviation (or 21 sigmas) from the average density of the map. The second highest peak had a difference of 12 sigmas after which peak heights decreased steadily. The abnormally high peak at 21 sigmas was one indication that this density did not

belong to a water molecule. Still, both a water molecule (Figure 4.6a) and a chloride ion (Figure 4.6b) were fit in the spherical density and the new maps and resulting B-factors were consulted to determine which molecule the density belonged to. In fitting the water molecule, the new map still showed green density with a height of 7.33 sigmas, and the B-factor was 3.83. After refining the chloride ion, however, the new map showed no leftover density and chloride's B-factor was much more consistent with the rest of the model at 15.77 (recall that the average B-factor of the model is 13.655). Most importantly, though, the chloride ion was the logical choice since chemically, the density was in a pocket within the protein molecule that did not contain any other negatively charged atoms. Also, bond distances to the nearest parts of the protein molecule were between 3.05 and 3.26 Å. These distances are consistent with the chloride distances found in a halide binding protein (51) on the Protein Data Bank (PDB ID: 3SVC) which are between 3.15 and 3.34 Å. Typical hydrogen bond distances, on the other hand, are between 2.4 and 3.2 Å.

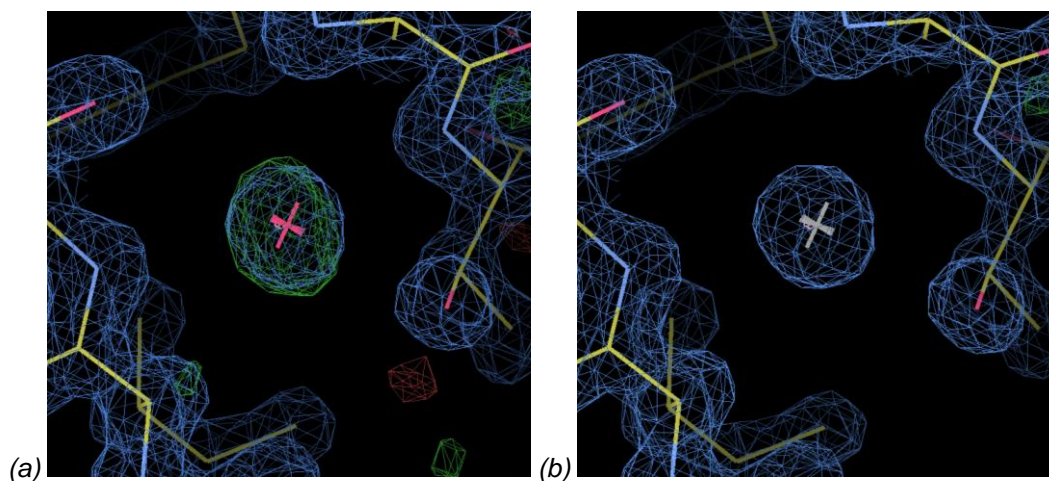


Figure 4.6. Water (a) and chloride ion (b) in a 21 sigmas difference peak. Positive electron density still exists around the water molecule after refinement in CCP4 indicating that water was not the likely reason for the observed electron density.

When all residues in the protein were adjusted and when all other ligand molecules had been added, the last task in Coot involved assigning water molecules to all other significant electron density. A common rule of thumb is that the R_{factor} should be 25% or less at this stage before adding waters. At this point the Az-PHM structure had an R_{factor} of 24.75%, and therefore, adding waters was the next step (52). There are two

ways to accomplish this task: automatically by using the *Find Waters* function in Coot or manually by assigning water molecules using the *Difference Map Peaks* tool. The *Find Waters* function has several advantages: it's a fast, systematic way to add waters and it provides some consistency across many structures in the way water molecules are added (52). However, the function is an automatic software function and could result in filled spots of electron density that do not make chemical sense. For this reason, waters were added to the Az-PHM model manually which allowed more control in the assessment of electron density blobs, where hydrogen-bonding distances could be checked and chemical knowledge could be applied. The difference map was first scanned for the highest peaks and water was assigned to those peaks that were spherical in appearance and within hydrogen bonding distance (2.4 to 3.2 Å) to polar protein atoms or other water molecules. The highest difference map peaks were determined based on their sigma values. The first shell included high peaks from 10-12 sigmas. Anywhere from 10-50 water molecules were typically assigned at a one time, after which they were merged to the structure and refined in CCP4. After refinement, the previously added water molecules were examined for fit to the new electron density map. If new electron density appeared that suggested the water was incorrectly assigned, it was removed from the structure before adding the next set of water molecules. In an ideal scenario like the one seen in Figure 4.7, a spherical blob of positive density will lie within hydrogen bonding distance of several water molecules or protein atoms.

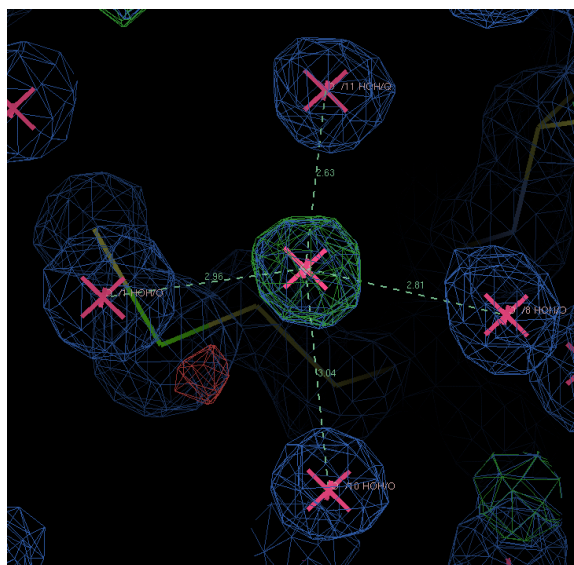


Figure 4.7. Green sphere of positive density fit with a water molecule (red cross in center). The water is ideally located within hydrogen-bonding distance of four other water molecules ~ 2.6 - 3.0 Å away.

After one “bin” of waters was added, the remaining difference map peaks were again studied. The set of electron density peaks in the next highest shell of sigmas was examined. These peaks were fitted with waters, the structure was re-refined and the waters were analyzed to confirm their chemical integrity. This process was repeated iteratively for 20 cycles until a total of 312 waters were added to the Az-PHM structure. Interestingly, after each addition of waters, the sigmas value describing the electron density for the remaining difference map peaks increased. This is a result of the way the sigma values are calculated. As more peaks are added to the model, the standard deviation (or sigma) for the total (assigned and unassigned) electron density decreases, thus the number of standard deviations (or sigmas) increases for un-modeled difference peaks even though the density of the peak remains the same.

Therefore, difficulty arose in deciding when to stop adding water molecules. When many of the significant difference peaks have been assigned, it can be difficult to determine which peaks are water molecules and which peaks are simply due to noise because the difference map starts to reveal more and more noise as their calculated sigmas begin to rise. In this case, crystallographers agree that it is best to err on the side of caution to avoid fitting noise. In determining when no more water molecules should be added, the R_{free} value was consulted since it is a global indicator of the progress of

refinement (52). As more and more bins of water molecules were added, the decrease in R_{free} became less and less substantial. When R_{free} no longer decreased, and noisy peaks became prevalent, adding water molecules ceased and one last refinement cycle was completed.

During the tedious process of adding water molecules, several unexplainable blobs of density appeared that could not be modeled by water, chloride or any of the other reagents used during crystallization. An example of one of these blobs can be seen below in Figure 4.8. Though the peanut-shaped blob did not appear to belong to PEG or nitrate, both of these molecules were used as a model (Figure 4.8a and Figure 4.8b) to fit the density. Both efforts gave decreased R-values after refinement in CCP4 but the fit was terrible, as evidenced by the negative electron density.

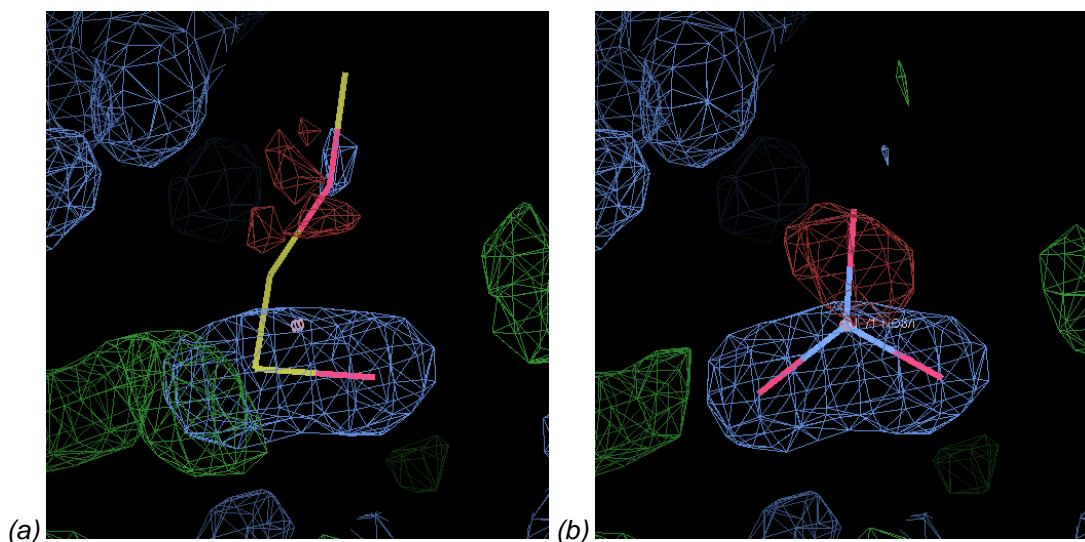


Figure 4.8. A peanut-shaped section of electron density fit with (a) a Polyethylene glycol molecule and (b) a nitrate molecule. Neither of these molecules gave a great fit.

Though modeling of this strange density and others was attempted, all efforts gave poor fits to the experimental density and even though R-values decreased in most cases, these areas could not be modeled in good conscience. This mentality ensured a conservative, yet reliable Az-PHM model where electron density was not “overfit” in an effort to push the R_{factor} down as low as possible (53). The final R_{factor} reached a value of 17.57% and R_{free} had a value of 20.70% in the final Az-PHM model. R_{factor} is expected to be lower than R_{free} because the reflections used in the calculation are not used for

refinement. However, because the diffraction data is high quality, an accurate model would yield structure factors that highly correlate with those that are not used in refinement (i.e. those that are used to calculate R_{free}), yielding similar values for R_{factor} and R_{free} . Kleywegt and Jones claim to obtain small differences in R-values of ~2% with good data sets and a difference as high as ~5-8% with poorer quality data sets and data sets with low resolution (53). The difference between R-values for the final Az-PHM model is ~3%, which suggests that the refined crystal structure adequately models the data without overfitting it.

Chapter 4.3 Validation of Final Model

X-ray crystallography is an exciting field that allows scientists to elucidate the structure of proteins and enzymes in order to better understand how structure influences function. Because inferences regarding function are made from analyzing a crystal structure, it is vital that structures are scrutinized and examined for potential errors before being made available to others. Thus, validation of the final Az-PHM model was essential to be certain that the model contained no major errors that could potentially impede the progress of other scientists who use this structure (47).

A number of structural details were examined using Coot's validation tools to ensure that a proper Az-PHM model was constructed that accurately fits the experimental electron data. In Chapter 4.2, recall that electron density and geometry were taken into account when making adjustments to the residues of the model. These tools used during refinement are validation tools that point out any outliers in the data. A representative portion of the *Density fit analysis* graph for the final model is shown in Figure 4.9. Because most of the residues show a short, green bar in the graph, the model is an excellent overall fit to the electron density map.

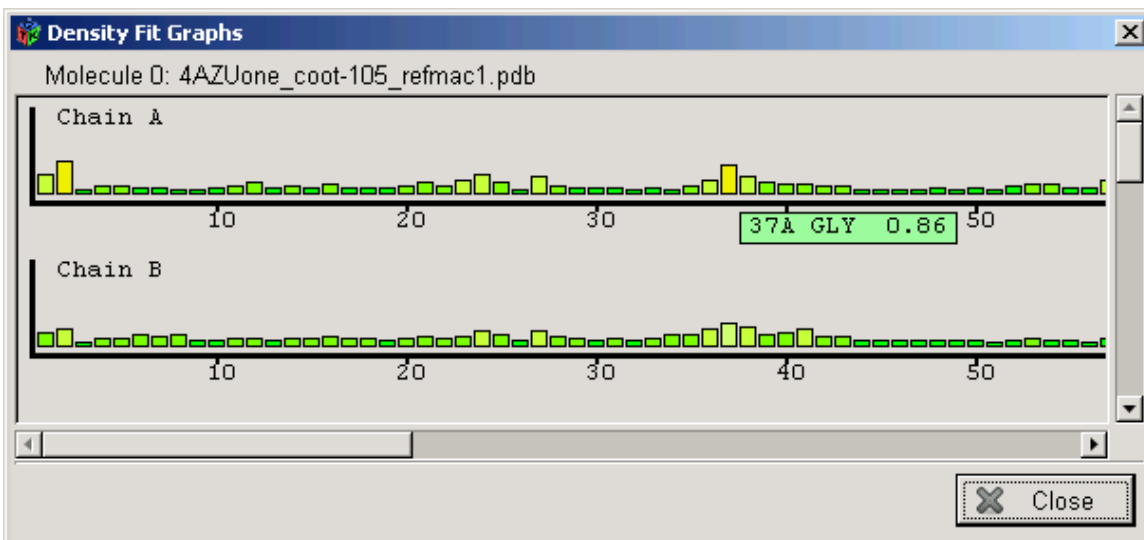


Figure 4.9. Density fit validation graph from Coot for the final model of Az-PHM. Each bar represents an amino acid in the model and the color and height indicate its fit to the electron density map. Green bars indicate excellent fit.

However, when the geometry of the model is considered (a representative portion of the graph is shown in Figure 4.10), it doesn't appear to be a very good structure. Many of the bars are yellow, orange and red, indicating moderate to poor geometry with respect to angles, bonds or planes. All of the amino acids represented by these colored bars were checked against the electron density map and 57 out of 61 residues showed excellent fit. Out of the 4 residues that did not appear to have a good fit, 2 of them gave an increase in R_{free} after modification and CCP4 refinement, and the other 2 did not show any electron density surrounding their side chains but also did not have any negative density either to suggest that their position was incorrect.

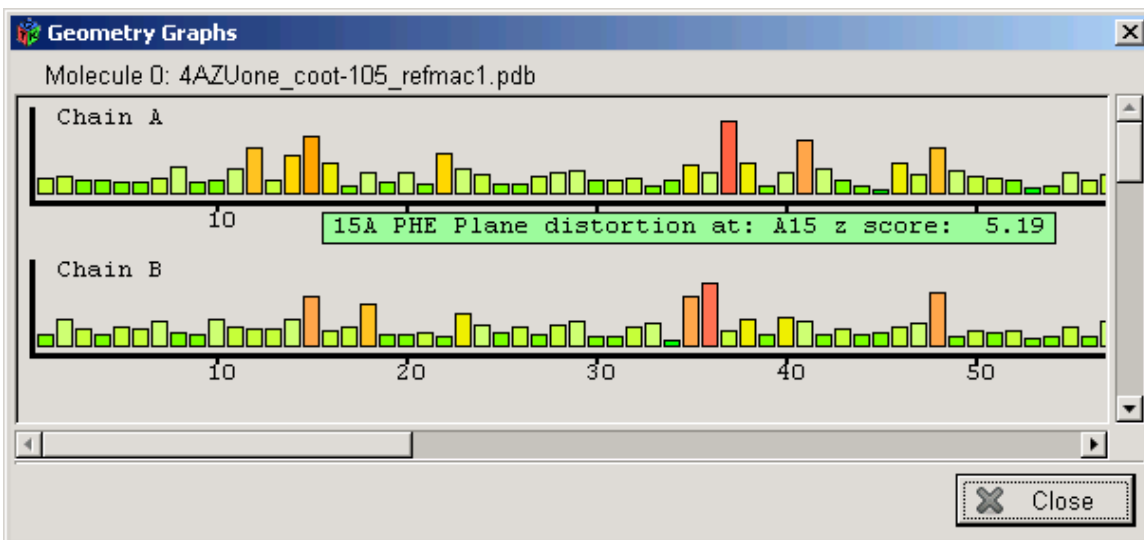


Figure 4.10. Geometry analysis validation graph from Coot for the final model of Az-PHM. Each bar represents an amino acid in the model and the color and height indicate its geometry relative to an average Z-score. Green bars indicate excellent geometry.

Though the geometry analysis appears to give moderate results, the fit of the model to the electron density is excellent. While there seem to be errors in geometry based on theoretically ideal geometries, the experimental data are at a high atomic resolution of 1.30 Å, and therefore, show exactly where atoms should be placed (with only a few exceptions due to unavailable density). To create the best model, recall that coordinates were adjusted and refined iteratively to minimize the values of R_{factor} and R_{free} . Refinement proceeded in this way because R_{free} is a powerful, unbiased statistic that provides a true validation check by evaluating the model on its ability to predict reflections from data that were not used in the creation of the model (47). Therefore, because the experimental density is clear and R-values were minimized, the residues have excellent fit and the resulting, objectionable geometries of some amino acid side-chains must be the result of inherent structural features instead of errors in the model (54). Consequently, the interactions that cause the protein to fold and crystallize in this way must outweigh any strain that results in maintaining the unusual conformations of some side-chains.

Coordinate-based validation methods, like *Geometry analysis* in Coot, are not as ideal as other methods that consider experimental data but nonetheless, can provide assurance that the model agrees with generally-known structural aspects of protein

molecules. Tools in this category are especially valuable if the criterion being used for validation was not restrained during refinement (47, 55). Because Refmac does not refine torsion angles, the model was analyzed using this criterion. The *Ramachandran Plot* function in Coot graphs each amino acid using its torsion angles, phi and psi. Because many of the combinations of phi and psi angles produce steric clashing (56), residues should theoretically only be located on a small portion of the plot. For Az-PHM, the residues in the diagram all fall in preferred or allowed regions (Figure 4.11). The Ramachandran diagram in Coot is interactive in that the “type” of residue selected determines the area of preferred (pink) and allowed (yellow) regions. For example, 44MetA, represented by a blue square with black outline is not a glycine (glycines are indicated by blue triangles) or a proline (prolines are indicated by a blue square with green outline) amino acid and therefore shows the preferred and allowed regions illustrated in Figure 4.11. Of this type of residues, only five (2.07%) fall in “allowed” regions while the rest have “preferred” phi and psi combinations. All proline and glycine residues also fall in “preferred” regions. No amino acids are “outliers”. The five amino acids that do fall in allowed regions include 10AsnA, 10AsnB, 44MetA, 44MetB, and 38AsnB. In viewing these residues in Coot, the electron density map is sharp and confirms their present positions. Emsley *et al.* describe a high quality Ramachandran plot as one having less than 1% outliers and where most residues fall in preferred regions (48). Thus, the Ramachandran plot for Az-PHM indicates that the coordinates result in a high-quality structure.

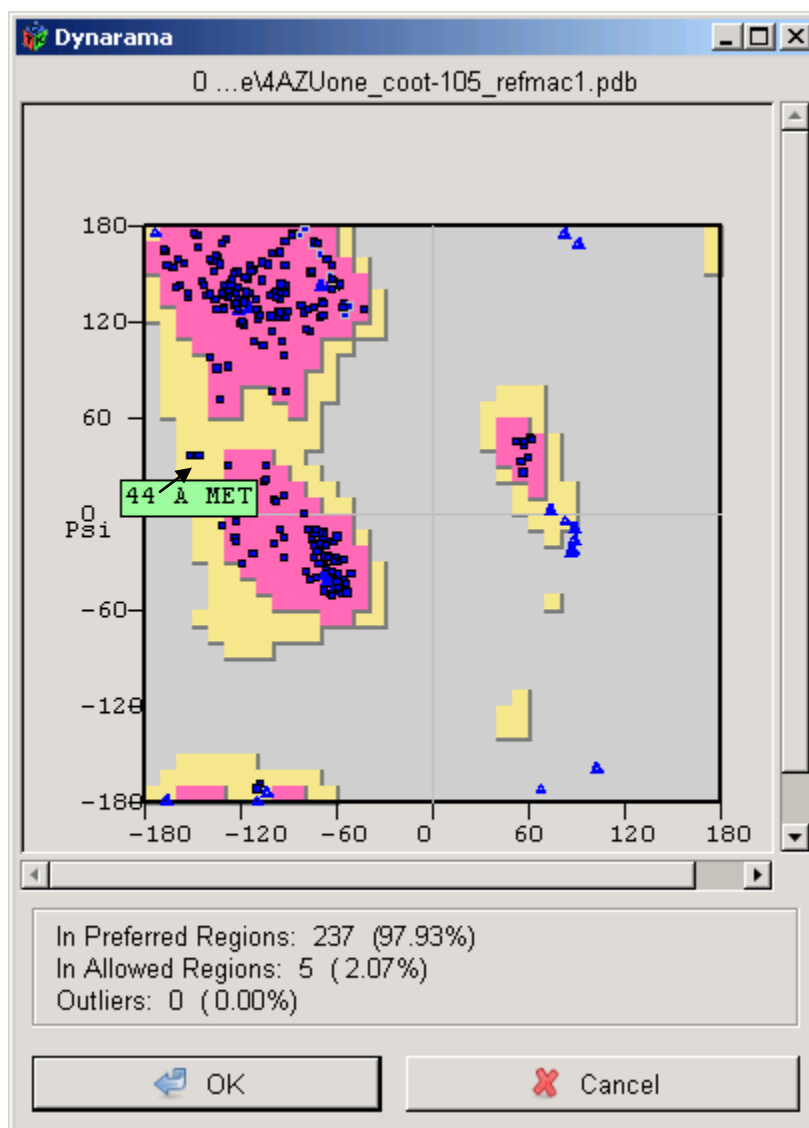


Figure 4.11. Ramachandran Diagram for all residues with preferred and allowed regions indicated for those that are not glycine or proline. The regions in pink are preferred and those in yellow are allowed regions.

The third coordinate-based validation tool used, *Peptide omega analysis*, also examines torsion angles. This function examines the third torsion angle, omega, by creating a bar graph where each bar represents an amino acid. The height and color of a bar is determined by the omega angle's deviation from $\sim 180^\circ$. Because omega measures the angle of the peptide linkage, it is expected to be slightly less than 180° due to its partial double-bond character. Kleywegt suggests that residues that have omega angles outside the range of $180^\circ \pm 20$ should be treated with caution in structures with low resolution (55). A representative portion of the omega graphs for the Az-PHM model is

shown in Figure 4.12. All residues in the model have omega angles in the accepted range with exception of 14HisA and 37GlyA. It seems that both of these residues are in the middle of sharp turns in the backbone, which explains why they are outside of the typical range. Plus, at high resolution, the density clearly indicates that these residues have been modeled clearly, which means that the abnormal omega angles are structural features of the model.

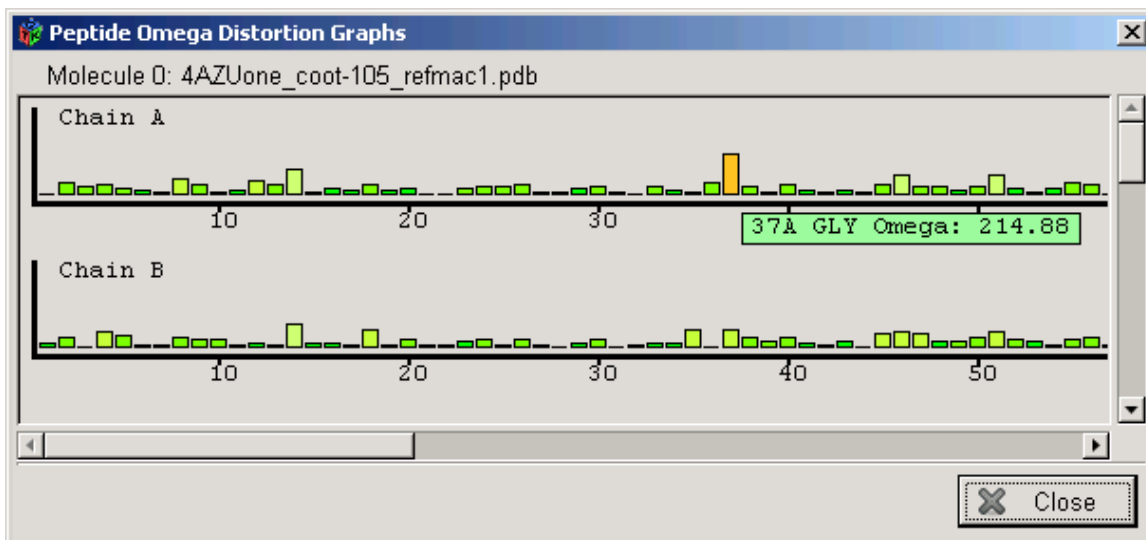


Figure 4.12. Peptide omega analysis graph for the final model of Az-PHM. Short green bars indicate residues that have acceptable omega angles. chain A and chain B refer to two distinct Az-PHM molecules in the asymmetric unit

The fourth coordinate-based method involves analysis of the amino acid side chain geometry through the use of conformation likelihoods. Residues in the Az-PHM model were compared with the Molprobity rotamer probability distribution (50) developed by Lovell and colleagues (48). The *Rotamer analysis* tool in Coot displays a bar graph containing probabilities for every amino acid in the model. The probability values express the likelihood of each amino acid's conformation, as seen in Figure 4.13 for 18AsnB. What is interesting about this particular residue is that it perfectly fits the electron density which is demonstrated in Figure 4.14. So, even though the probability of the rotamer conformation for 18B is only 0.02%, the side-chain exists in this conformation when the protein is in crystalline form. Other residues that give similar probabilities include 70LysB, 122LysA and 122LysB and two of these amino acids show defined density around the atoms of the residues. The lysine residue in chain A does not

show convincing density for its atomic coordinates, however, it does not show negative density either.

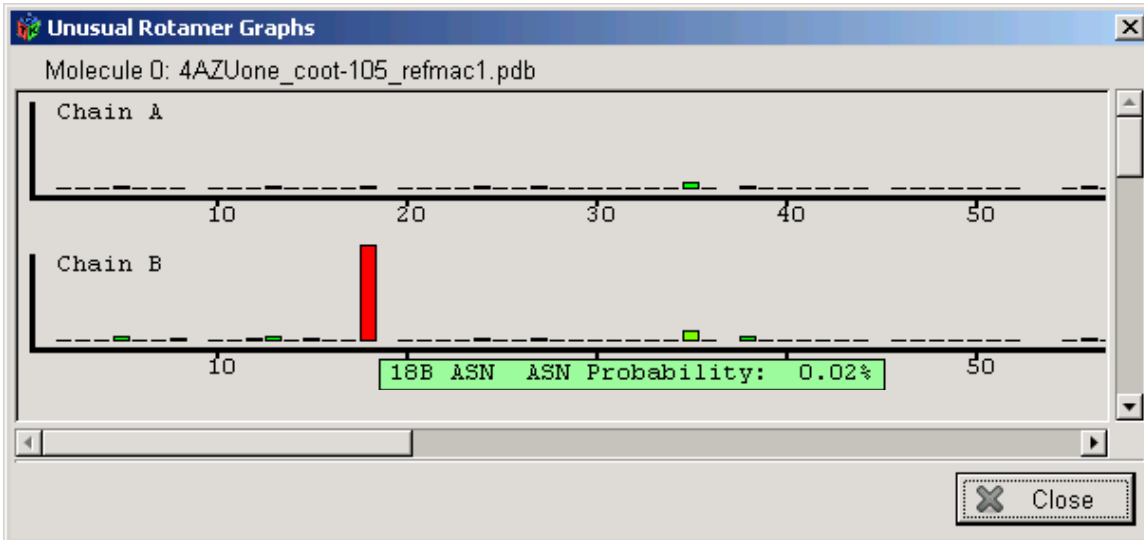


Figure 4.13. Rotamer analysis graphs created in Coot for the final Az-PHM model. Bars are inversely proportional to probability.

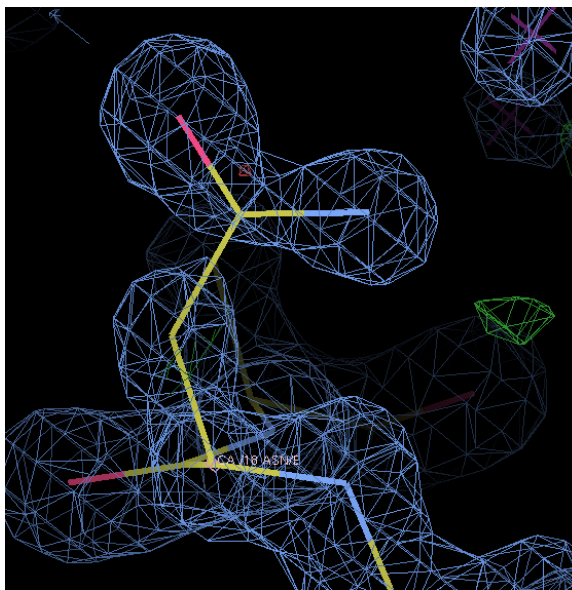


Figure 4.14. Final electron density and structure for the asparagine residue at 18 in chain B in the Az-PHM model. Though the rotamer probability suggested this conformation was 0.02% likely, the electron density shows strong evidence that suggests 18AsnB crystallized this way.

The fifth coordinate-based validation method compares the geometry of each residue in chain A against the same residue of chain B in Coot's *NCS Differences*

function. Because NCS restraints were not used during refinement in CCP4, this function identifies areas with potential error that are worth investigating. The differences in atomic positions between residues are displayed in a bar graph containing r.m.s. deviations (48) after one chain has been translated and superimposed onto another (55). For Az-PHM, a portion of the *NCS Difference* results is shown in Figure 4.15. All residues showing yellow, orange or red bars (r.m.s. ~ 1 or greater) were checked and out of the 12 pairs showing significant differences in atomic position (1, 69, 104, 105, 106, 126 and 127), all but 5 pairs showed electron density to indicate that they were appropriately modeled and that the differences were not errors. For the 5 pairs of residues where differences could not be confirmed by electron density (13, 85, 103, 107 and 128), only 2 showed negative density for the side-chain positions (107 and 128). However, 107GlnA was moved several times to try to the experimental density better and each time, the R_{free} value increased, suggesting that this difference may be a structural feature instead of an error. The other 3 pairs (2 lysine pairs and 1 methionine pair) where atomic positions could not be confirmed did not have clear density to verify their position. In the lysine pairs, electron density was unavailable and for 13MetB, green density in the difference map suggested an alternate conformation should be modeled but could not be because a water molecule with a higher difference peak was too close. Interestingly, most of the pairs showing large differences came from two exterior areas containing turns in the backbone.

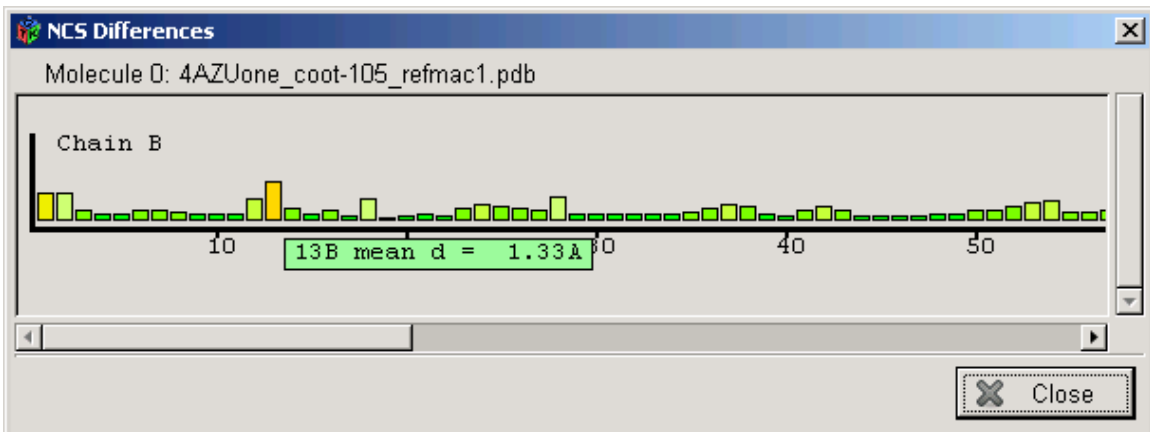


Figure 4.15. Portion of the NCS Differences graph for the final Az-PHM model. Bar height is proportional to the r.m.s. deviations between residue pairs in chains A and B.

The sixth and final tool used in validation of the Az-PHM coordinates described atomic positions in terms of temperature factors. The results of this *Temperature factor variance analysis* function in Coot, a portion of which is visible in Figure 4.16, show high average B-factors for residues in the Az-PHM model that are lengthy and solvent-exposed. Approximately ~70% of these residues are lysine and glutamate amino acids.



Figure 4.16. Results of the Temperature factor variance analysis performed on the final Az-PHM model in Coot.

Lastly, two simple tests for chirality and atom location were conducted in Coot. The test for chirality searched for *Incorrect Chiral Volumes* that would indicate the presence of erroneous D-amino acids and the test for *glutamine/asparagine outliers* attempted to detect incorrectly placed oxygen and nitrogen atoms in glutamine and asparagine side-chains using B-factor analysis. Coot found no incorrect chiral volumes and no glutamine/asparagine outliers. Interestingly, oxygen and nitrogen atoms on a couple of residues were exchanged and the test was conducted again, but did not yield different results, which indicates that this test is not sensitive even with atomic resolution data since the switch went undetected.

Results of all of the validation tools in Coot were examined to assess the quality of the final Az-PHM model and prepare it for submission to the Protein Data Bank (48). After all of the validation tests were completed, the final structure was found to be a good model of the experimental data since it scored well on every test (53). Even though a few

outliers were found when each validation was used, the atomic resolution and quality of the data collected suggest that most if not all of the flagged coordinates were structural features instead of errors. Users of this structure can have confidence that the model is a highly supported illustration of the crystallized Az-PHM protein.

CHAPTER 5

Chapter 5.1 Analysis of Final Az-PHM Crystal Structure

After successfully obtaining an accurate crystal structure, the Az-PHM model was analyzed. The overall structure, shown in Figure 5.1, is a dimer wherein the two Az-PHM molecules are pointed toward each other. A solvent filled channel spans the space between the two molecules. Two of the structure's six copper(II) ions are also located between the two structures. Histidine 83A and 83B each bind a copper in the solvent gap along with Tris molecules. The other four coppers are located in the Type 1 and Type 2 copper active sites.

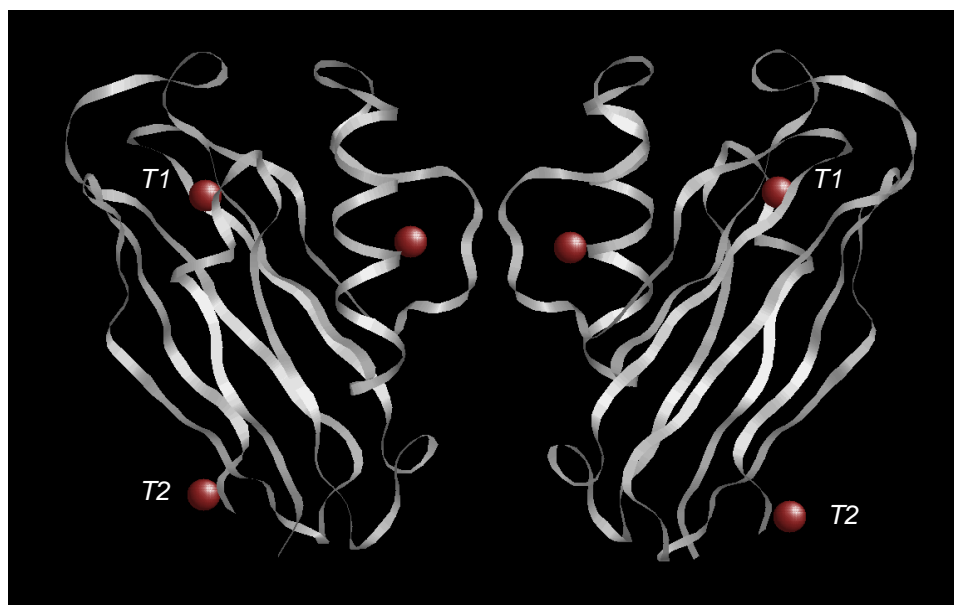


Figure 5.1. Dimerized Az-PHM molecules in the final crystal structure. Copper(II) atoms are shown as maroon spheres. Copper sites are denoted for each monomer.

Upon initial investigation of the final model, it appears that the presence of a third set of copper(II) ions and the dimerized nature of the structure alter perceptions of the Az-PHM variant. It was created as a monomer to bind coppers at the T1 and T2 sites. The two copper ions bound to histidine residues 83A and 83B are located in the center of the final model in Figure 5.1 and are somewhat unique when examining other wild-type azurin structures. The copper ions are separated by a distance of 11.9 Å and are largely solvent-exposed. The other aspect of the final crystal structure that was not anticipated is

dimerization. If it exists in solution, dimerization may affect future physiological studies. In this structure, Az-PHM monomers dimerize due to interactions at the T2 site where one monomer binds the T2 copper ion at His14 and His16, as designed, and the neighboring monomer also interacts with the T2 copper center through Ala1. Several structures of wild-type azurin show copper(II) bound to Ala1 so at high concentrations, it seems probable for Az-PHM dimerization at this residue to occur. However, if dimerization did occur even to a small extent in solution, it follows that EPR studies would show three distinct copper ions, one at the T1 site, one at the T2 site (bound by His14 and 16) and one dimerized copper (bound by His14, His16 and Ala1). Berry *et al.* show that at concentrations of 1 mM, Az-PHM samples only show two types of copper ions at the T1 and T2 sites. Even when excess copper was added to the samples, it did not bind to the protein. A third signal consistent with free copper in solution was observed (15). This EPR data also suggests that the copper(II) ions bound to His83A and 83B do not exist in solution. If they did, the third signal would be consistent with a bound copper instead of free copper(II). Therefore, the third copper ion bound to His83 and the dimerization appear to be artifacts of crystallography rather than structural features.

In creating the biosynthetic Az-PHM model to mimic PHM, it is important that its crystal structure be examined for its structural similarity to the native enzyme's active sites. Recall that PHM has two copper binding sites, one of which is bound by three histidine residues and functions in electron transfer while the other is bound by two histidines and a methionine and functions in catalysis. Since mechanistic studies suggest that an electron is passed from the T1 copper ion to the T2 copper site where catalysis occurs (9), these two copper sites must be analyzed in the Az-PHM model.

The T1 copper site in the Az-PHM model is native to azurin and therefore, should have bonding distances similar to those found in the native protein. In the final Az-PHM structure, the T1 copper shown in Figure 5.2 coordinates two histidine residues and one cysteine residue in a trigonal planar fashion. Bond distances from copper to His46, His117 and Cys112 are ~2.00-2.03 Å, 2.02-2.04 Å and 2.18-2.19 Å, respectively (range limits given for all Az-PHM distances express actual distances from both monomers). In

the wild-type azurin crystal structure, the distance from copper to His46 is 1.99-2.13 Å, from copper to His117 is 1.96-2.11 Å and from copper to Cys112 is 2.17-2.27 Å (ranges given for wild-type azurin cover actual measured distances from all four monomers in the 4AZU crystal structure). The axial ligands that coordinate the T1 center in Az-PHM, Met121 and Gly45, have copper bond distances of 3.27-3.34 Å and 2.61-2.68 Å, respectively in Az-PHM, and similarly, the native distances are 3.05-3.21 Å and 2.84-3.03 Å, respectively.

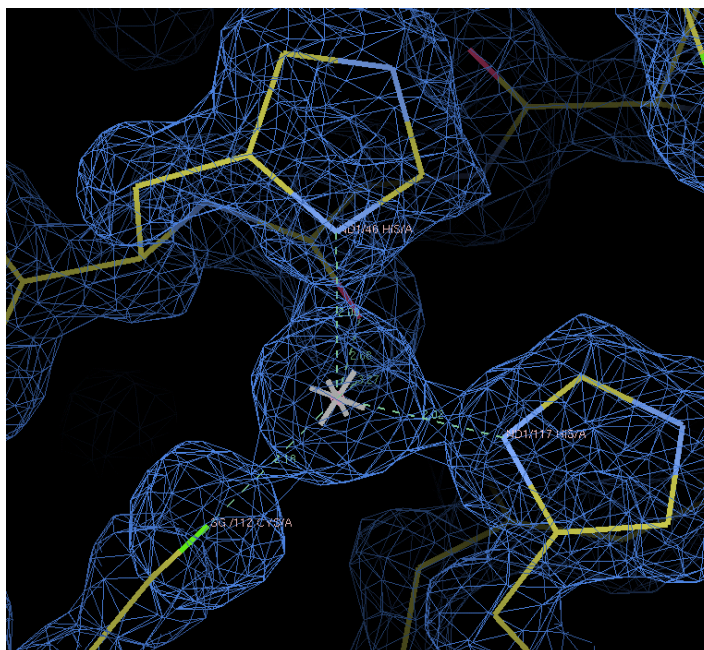


Figure 5.2. Type 1 copper active site in the final Az-PHM crystal structure. The ligands shown include His46, His117 and Cys112 which all coordinate the copper(II) ion in a trigonal planar fashion.

The Az-PHM model was crystallized in order to compare its designed T2 copper site to the native system's catalytic site. Because the azurin scaffold already contained a T1 copper site, only three residues were mutated to incorporate a second copper binding site into the protein. Residues 14 and 16 were mutated to histidines and residue 8 was mutated to a methionine. When viewing the T2 site shown in Figure 5.3, the Az-PHM structure reveals coordination of the copper(II) ion by His14, His16 and Ala1. Unfortunately, the geometry around the T2 copper was compromised due to the way the protein crystallized such that the third ligand, Ala1, is from an Az-PHM molecule in the neighboring asymmetric unit. In the Az-PHM crystal structure, the geometry of the site is

square pyramidal. The His14, His16, Ala1(O), and Ala1(N) distances are 1.97-1.98, 2.07-2.10, 2.04-2.10, and 2.05-2.20 Å, respectively. (Range limits signify actual distances measured for both T2 sites in the asymmetric unit.) All of these atoms lie in a plane with copper while a water molecule binds perpendicular to the plane to create a pyramid. Copper lies a mere ~0.1 Å out of a least-squares plane created by all four residue ligands. The water molecule is positioned above the plane with a distance to the copper(II) ion of 2.42-2.48 Å.

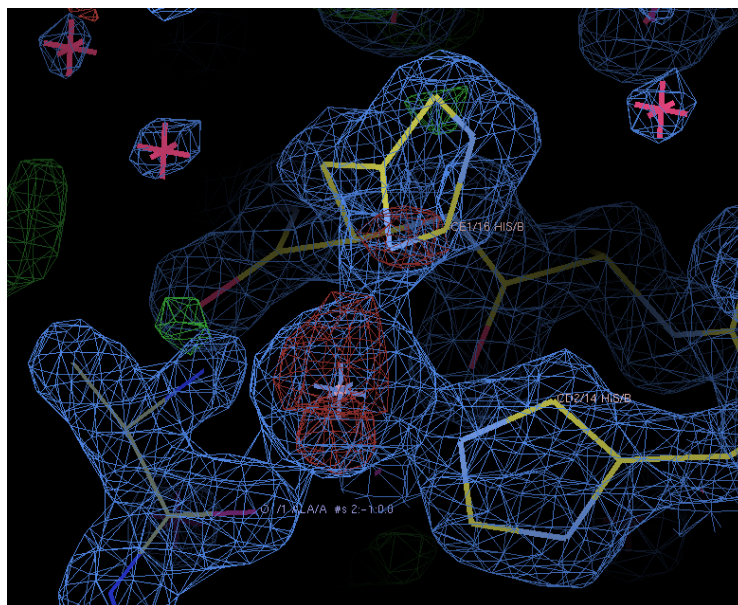


Figure 5.3. Type 2 copper active site in the final Az-PHM crystal structure. The copper(II) ion is bound by His14, His16 and an Ala1 residue from a neighboring Az-PHM molecule.

Because Met8, His14 and His16 were mutated in azurin to create the T2 copper site in Az-PHM, it is necessary to examine the final crystal structure against native azurin in order to determine the effects of the mutations on protein folding. Figure 5.4 shows the overlay of wild-type azurin (PDB ID: 4AZU) in blue and one monomer of the final Az-PHM structure in red. No gross differences exist between the backbones of the structures, which suggests that the three mutations that were made do not affect the ability of the protein to fold properly. Each monomer contains beta-sheet secondary structure with a Greek key motif as well as a looped section containing alpha helices. This also supports the conclusions that many researchers have made concerning the

stability of azurin in the presence of many mutations (13, 14). The only significant difference between the two structures appears at Ala1, where this residue binds the copper ion that bridges two azurin molecules.

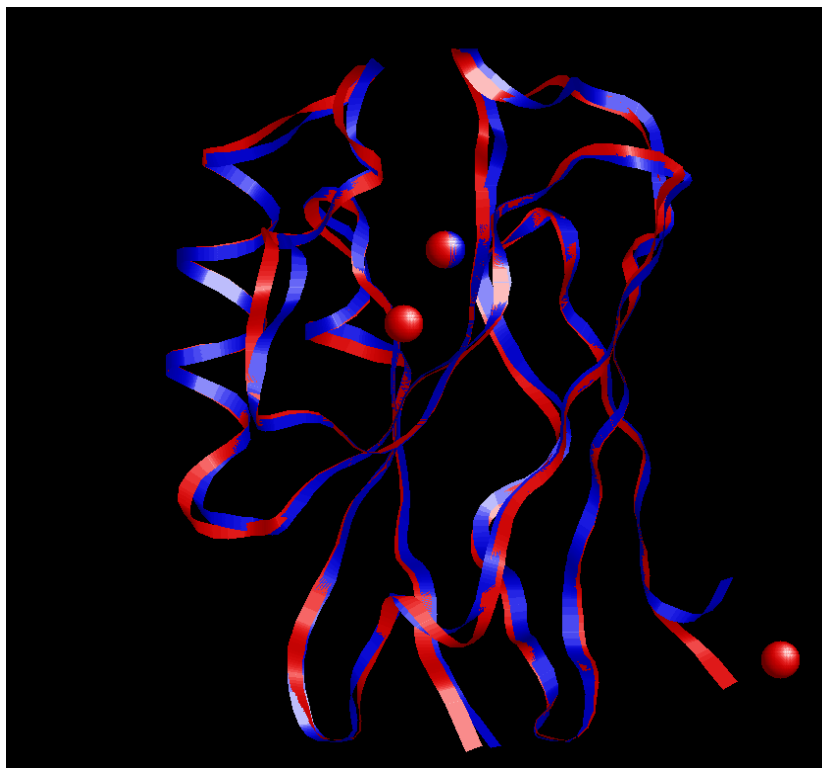


Figure 5.4. Overlay of one monomer of the 4AZU wild-type azurin crystal structure and the final structure of the Az-PHM model. Azurin is in blue and Az-PHM is colored red.

Analysis of the copper binding sites and the overall shape of Az-PHM indicate that the mutations to azurin were successful and the T1 copper center was not disturbed. The T1 and T2 coppers in the model are ~ 14 Å apart which is just slightly larger than the distance of 11.3 Å found in the native PHM crystal structure (7). The coordination mode of the T2 site appears to be an artifact of protein packing such that crystallizing Az-PHM in a different way may lead to a T2 site that is not dimerized with another azurin molecule and therefore, give a better idea of the structure of the protein in solution. With the knowledge gleaned from structural analysis thus far, Az-PHM seems to model the native PHM protein well.

Chapter 5.2 Conclusions and Future directions

To study the mammalian enzyme PHM, a biosynthetic model was created using azurin. The resulting Az-PHM model was successfully purified from *E. coli* cells through lysis with osmotic shock and precipitation, followed by three chromatography columns. The purity of the sample was evaluated using spectroscopic methods and confirmed using SDS-PAGE. Crystals of Az-PHM were successfully grown and crystal properties were tuned using different reagents at various concentrations to achieve the best possible intensity, resolution and mosaicity. One of the diffraction-quality crystals was irradiated with copper K_{α} x-rays to collect a full set of high-resolution images. CrystalClear software from Rigaku was used to assess the symmetry of the crystal and harvest intensity data from the diffraction images. Using the CCP4 software and a process called molecular replacement, the phase-problem was solved and preliminary phases were assigned to the intensity data. After rounds of refinement using Coot, a final crystal structure was produced at 1.3 Å resolution with an R_{factor} of 17.57% and an R_{free} value of 20.78%.

The model's overall structure and its copper active sites were compared to wild-type azurin and native PHM. Structurally, the model seems to mimic PHM for several reasons. First, the Type 1 copper site geometry in Az-PHM was untouched and should therefore function in electron transfer in the same way that PHM's Cu_H site does. Second, the designed site incorporated into azurin binds a copper(II) ion, though the binding in this crystal structure is not ideal since dimerization with another protein molecule occurs. Third, the 14 Å distance between the copper sites in the model is comparable to the native Cu-Cu distance of 11.3 Å in PHM, and finally, the mutations made to create the model did not cause the protein to misfold.

Even though the Type 2 copper site bridged to another molecule in the crystal lattice, the crystal structure has high resolution and will be deposited into the Protein Data Bank. Copper-ligand bond distances for the Type 1 copper site were very consistent between monomers (much more so than those in 4AZU) and thus, may be more accurate. The Type 2 site, though it does not hold all of the information hoped for, does bind a copper and seems distorted merely due to the way the protein molecules dimerized and

packed together during crystallization. Thus, attempts to obtain a diffraction-quality Az-PHM crystal under different conditions will be pursued in the future in order to elucidate the geometry of the designed Type 2 copper site as it exists in solution

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APPENDIX

Information concerning a structure's coordinates, geometry, r.m.s. deviations and refinement details are found in its coordinate file. The coordinate file for the final Az-PHM structure is given below.

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HEADER ELECTRON TRANSPORT(C          23-JUN-93 4AZU
COMPND CONFORMATIONAL TRANSITION INVOLVES A PE
REMARK 3
REMARK 3 REFINEMENT.
REMARK 3 PROGRAM   : REFMAC 5.5.0109
REMARK 3 AUTHORS   : MURSHUDOV,VAGIN,DODSON
REMARK 3
REMARK 3 REFINEMENT TARGET : MAXIMUM LIKELIHOOD
REMARK 3
REMARK 3 DATA USED IN REFINEMENT.
REMARK 3 RESOLUTION RANGE HIGH (ANGSTROMS) : 1.30
REMARK 3 RESOLUTION RANGE LOW  (ANGSTROMS) : 28.05
REMARK 3 DATA CUTOFF          (SIGMA(F)) : NONE
REMARK 3 COMPLETENESS FOR RANGE (%) : 89.23
REMARK 3 NUMBER OF REFLECTIONS      : 50049
REMARK 3
REMARK 3 FIT TO DATA USED IN REFINEMENT.
REMARK 3 CROSS-VALIDATION METHOD      : THROUGHOUT
REMARK 3 FREE R VALUE TEST SET SELECTION : RANDOM
REMARK 3 R VALUE (WORKING + TEST SET) : 0.17730
REMARK 3 R VALUE (WORKING SET) : 0.17572
REMARK 3 FREE R VALUE                : 0.20704
REMARK 3 FREE R VALUE TEST SET SIZE (%) : 5.1
REMARK 3 FREE R VALUE TEST SET COUNT   : 2672
REMARK 3
REMARK 3 FIT IN THE HIGHEST RESOLUTION BIN.
REMARK 3 TOTAL NUMBER OF BINS USED      : 20
REMARK 3 BIN RESOLUTION RANGE HIGH      : 1.300
REMARK 3 BIN RESOLUTION RANGE LOW       : 1.334
REMARK 3 REFLECTION IN BIN (WORKING SET) : 2868
REMARK 3 BIN COMPLETENESS (WORKING+TEST) (%) : 70.86
REMARK 3 BIN R VALUE (WORKING SET) : 0.262
REMARK 3 BIN FREE R VALUE SET COUNT     : 167
REMARK 3 BIN FREE R VALUE                : 0.301
REMARK 3
REMARK 3 NUMBER OF NON-HYDROGEN ATOMS USED IN REFINEMENT.
REMARK 3 ALL ATOMS                      : 2349
REMARK 3
REMARK 3 B VALUES.
REMARK 3 FROM WILSON PLOT (A**2) : NULL
REMARK 3 MEAN B VALUE (OVERALL, A**2) : 14.138
REMARK 3 OVERALL ANISOTROPIC B VALUE.
REMARK 3 B11 (A**2) : 0.00
REMARK 3 B22 (A**2) : 0.00
REMARK 3 B33 (A**2) : 0.00
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REMARK 3 B12 (A**2): 0.00
 REMARK 3 B13 (A**2): 0.00
 REMARK 3 B23 (A**2): 0.00
 REMARK 3
 REMARK 3 ESTIMATED OVERALL COORDINATE ERROR.
 REMARK 3 ESU BASED ON R VALUE (A): 0.058
 REMARK 3 ESU BASED ON FREE R VALUE (A): 0.062
 REMARK 3 ESU BASED ON MAXIMUM LIKELIHOOD (A): 0.037
 REMARK 3 ESU FOR B VALUES BASED ON MAXIMUM LIKELIHOOD (A**2): 1.672
 REMARK 3
 REMARK 3 CORRELATION COEFFICIENTS.
 REMARK 3 CORRELATION COEFFICIENT FO-FC : 0.967
 REMARK 3 CORRELATION COEFFICIENT FO-FC FREE : 0.951
 REMARK 3
 REMARK 3 RMS DEVIATIONS FROM IDEAL VALUES COUNT RMS WEIGHT
 REMARK 3 BOND LENGTHS REFINED ATOMS (A): 2076 ; 0.027 ; 0.021
 REMARK 3 BOND ANGLES REFINED ATOMS (DEGREES): 2815 ; 2.213 ; 1.962
 REMARK 3 TORSION ANGLES, PERIOD 1 (DEGREES): 274 ; 7.027 ; 5.000
 REMARK 3 TORSION ANGLES, PERIOD 2 (DEGREES): 84 ; 39.420 ; 25.952
 REMARK 3 TORSION ANGLES, PERIOD 3 (DEGREES): 373 ; 13.156 ; 15.000
 REMARK 3 TORSION ANGLES, PERIOD 4 (DEGREES): 2 ; 14.198 ; 15.000
 REMARK 3 CHIRAL-CENTER RESTRAINTS (A**3): 315 ; 0.167 ; 0.200
 REMARK 3 GENERAL PLANES REFINED ATOMS (A): 1522 ; 0.014 ; 0.021
 REMARK 3
 REMARK 3 ISOTROPIC THERMAL FACTOR RESTRAINTS. COUNT RMS WEIGHT
 REMARK 3 MAIN-CHAIN BOND REFINED ATOMS (A**2): 1312 ; 1.329 ; 1.500
 REMARK 3 MAIN-CHAIN ANGLE REFINED ATOMS (A**2): 2125 ; 2.042 ; 2.000
 REMARK 3 SIDE-CHAIN BOND REFINED ATOMS (A**2): 764 ; 2.875 ; 3.000
 REMARK 3 SIDE-CHAIN ANGLE REFINED ATOMS (A**2): 680 ; 4.252 ; 4.500
 REMARK 3
 REMARK 3 NCS RESTRAINTS STATISTICS
 REMARK 3 NUMBER OF NCS GROUPS : NULL
 REMARK 3
 REMARK 3 TWIN DETAILS
 REMARK 3 NUMBER OF TWIN DOMAINS : NULL
 REMARK 3
 REMARK 3
 REMARK 3 TLS DETAILS
 REMARK 3 NUMBER OF TLS GROUPS : 2
 REMARK 3 ATOM RECORD CONTAINS RESIDUAL B FACTORS ONLY
 REMARK 3
 REMARK 3 TLS GROUP : 1
 REMARK 3 NUMBER OF COMPONENTS GROUP : 1
 REMARK 3 COMPONENTS C SSSEQI TO C SSSEQI
 REMARK 3 RESIDUE RANGE : A -10 A 9999
 REMARK 3 ORIGIN FOR THE GROUP (A): -10.4640 10.3031 -20.6267
 REMARK 3 T TENSOR
 REMARK 3 T11: 0.0035 T22: 0.0322
 REMARK 3 T33: 0.0189 T12: -0.0002
 REMARK 3 T13: 0.0017 T23: 0.0029
 REMARK 3 L TENSOR
 REMARK 3 L11: 0.5667 L22: 1.2055
 REMARK 3 L33: 1.0449 L12: 0.1447
 REMARK 3 L13: -0.2101 L23: -0.2559
 REMARK 3 S TENSOR

REMARK 3 S11: 0.0070 S12: -0.0142 S13: -0.0031
REMARK 3 S21: -0.0472 S22: -0.0132 S23: 0.0543
REMARK 3 S31: 0.0280 S32: -0.0559 S33: 0.0062
REMARK 3
REMARK 3 TLS GROUP : 2
REMARK 3 NUMBER OF COMPONENTS GROUP : 1
REMARK 3 COMPONENTS C SSSEQI TO C SSSEQI
REMARK 3 RESIDUE RANGE : B -10 B 9999
REMARK 3 ORIGIN FOR THE GROUP (A): -27.2598 28.9550 -1.1153
REMARK 3 T TENSOR
REMARK 3 T11: 0.0462 T22: 0.0064
REMARK 3 T33: 0.0053 T12: -0.0149
REMARK 3 T13: 0.0021 T23: 0.0007
REMARK 3 L TENSOR
REMARK 3 L11: 0.8751 L22: 0.6344
REMARK 3 L33: 1.1725 L12: 0.0514
REMARK 3 L13: 0.6096 L23: 0.3847
REMARK 3 S TENSOR
REMARK 3 S11: -0.0539 S12: 0.0009 S13: -0.0239
REMARK 3 S21: -0.1557 S22: 0.0468 S23: 0.0002
REMARK 3 S31: -0.0760 S32: 0.0062 S33: 0.0071
REMARK 3
REMARK 3
REMARK 3 BULK SOLVENT MODELLING.
REMARK 3 METHOD USED : BABINET MODEL WITH MASK
REMARK 3 PARAMETERS FOR MASK CALCULATION
REMARK 3 VDW PROBE RADIUS : 1.40
REMARK 3 ION PROBE RADIUS : 0.80
REMARK 3 SHRINKAGE RADIUS : 0.80
REMARK 3
REMARK 3 OTHER REFINEMENT REMARKS:
REMARK 3 HYDROGENS HAVE BEEN ADDED IN THE RIDING POSITIONS
REMARK 3 U VALUES : RESIDUAL ONLY
REMARK 3
SSBOND 1 CYS A 3 CYS A 26
SSBOND 2 CYS B 3 CYS B 26
LINKR NE2 HIS A 83 CU CU F 1 HIS-CU
LINKR N ALA A 1 CU CU J 1 ALA-CU
LINKR O ALA A 1 CU CU J 1 ALA-CU1
LINKR ND1 HIS A 46 CU CU C 1 HIS-CU1
LINKR SG CYS A 112 CU CU C 1 CYS-CU
LINKR ND1 HIS A 117 CU CU C 1 HIS-CU1
LINKR N ALA B 1 CU CU G 1 ALA-CU
LINKR O ALA B 1 CU CU G 1 ALA-CU1
LINKR NE2 HIS B 83 CU CU I 1 HIS-CU
LINKR ND1 HIS B 46 CU CU D 1 HIS-CU1
LINKR SG CYS B 112 CU CU D 1 CYS-CU
LINKR ND1 HIS B 117 CU CU D 1 HIS-CU1
LINKR CU CU F 1 O3 TRS E 1 CU-TRS
LINKR CU CU F 1 N TRS E 1 CU-TRS1
LINKR CU CU I 1 N TRS H 1 CU-TRS1
LINKR CU CU F 1 C2 TRS g 1 CU-TRS2
LINKR CU CU F 1 O2 TRS g 1 CU-TRS3
LINKR CU CU F 1 O1 TRS g 1 CU-TRS4
CRYST1 50.178 54.895 85.919 90.00 90.00 90.00 P 21 21 21

SCALE1	0.019929	0.000000	0.000000	0.000000								
SCALE2	0.000000	0.018217	0.000000	0.000000								
SCALE3	0.000000	0.000000	0.011639	0.000000								
ATOM	1	N	ALA	A	1	-14.633	-9.476	-8.661	1.00	19.02	AA	N
ATOM	2	CA	ALA	A	1	-15.132	-8.162	-9.248	1.00	17.94	AA	C
ATOM	3	CB	ALA	A	1	-15.499	-7.147	-8.125	1.00	18.09	AA	C
ATOM	4	C	ALA	A	1	-14.136	-7.558	-10.161	1.00	17.21	AA	C
ATOM	5	O	ALA	A	1	-12.938	-7.781	-9.952	1.00	17.41	AA	O
ATOM	6	N	GLU	A	2	-14.619	-6.849	-11.165	1.00	14.78	AA	N
ATOM	7	CA	GLU	A	2	-13.763	-6.185	-12.139	1.00	17.35	AA	C
ATOM	8	CB	GLU	A	2	-13.605	-6.963	-13.471	1.00	20.52	AA	C
ATOM	9	CG	GLU	A	2	-13.237	-8.476	-13.363	1.00	24.46	AA	C
ATOM	10	CD	GLU	A	2	-11.874	-8.793	-12.718	1.00	35.77	AA	C
ATOM	11	OE1	GLU	A	2	-10.977	-7.898	-12.714	1.00	41.25	AA	O
ATOM	12	OE2	GLU	A	2	-11.701	-9.942	-12.225	1.00	34.43	AA	O
ATOM	13	C	GLU	A	2	-14.168	-4.745	-12.426	1.00	17.02	AA	C
ATOM	14	O	GLU	A	2	-14.531	-4.389	-13.562	1.00	21.18	AA	O
ATOM	15	N	CYS	A	3	-13.924	-3.885	-11.463	1.00	11.30	AA	N
ATOM	16	CA	CYS	A	3	-14.452	-2.484	-11.522	1.00	10.97	AA	C
ATOM	17	CB	CYS	A	3	-15.109	-2.131	-10.174	1.00	10.00	AA	C
ATOM	18	SG	CYS	A	3	-16.539	-3.238	-9.819	1.00	10.95	AA	S
ATOM	19	C	CYS	A	3	-13.372	-1.467	-11.875	1.00	10.61	AA	C
ATOM	20	O	CYS	A	3	-13.411	-0.334	-11.480	1.00	10.61	AA	O
ATOM	21	N	SER	A	4	-12.408	-1.946	-12.663	1.00	10.81	AA	N
ATOM	22	CA	SER	A	4	-11.409	-1.026	-13.163	1.00	11.31	AA	C
ATOM	23	CB	SER	A	4	-10.271	-1.036	-12.202	1.00	12.35	AA	C
ATOM	24	OG	SER	A	4	-9.565	-2.265	-12.277	1.00	15.23	AA	O
ATOM	25	C	SER	A	4	-10.908	-1.387	-14.520	1.00	12.94	AA	C
ATOM	26	O	SER	A	4	-11.143	-2.538	-15.007	1.00	13.32	AA	O
ATOM	27	N	VAL	A	5	-10.196	-0.477	-15.180	1.00	10.03	AA	N
ATOM	28	CA	VAL	A	5	-9.515	-0.782	-16.426	1.00	10.47	AA	C
ATOM	29	CB	VAL	A	5	-10.409	-0.448	-17.628	1.00	12.05	AA	C
ATOM	30	CG1	VAL	A	5	-10.766	1.087	-17.714	1.00	12.84	AA	C
ATOM	31	CG2	VAL	A	5	-9.847	-1.010	-18.917	1.00	17.27	AA	C
ATOM	32	C	VAL	A	5	-8.228	-0.010	-16.467	1.00	10.01	AA	C
ATOM	33	O	VAL	A	5	-8.126	1.115	-15.931	1.00	10.86	AA	O
ATOM	34	N	ASP	A	6	-7.210	-0.614	-17.057	1.00	9.44	AA	N
ATOM	35	CA	AASP	A	6	-5.903	0.070	-17.300	0.50	8.89	AA	C
ATOM	36	CA	BASP	A	6	-5.886	0.037	-17.300	0.50	9.40	AA	C
ATOM	37	CB	AASP	A	6	-4.729	-0.869	-17.056	0.50	8.64	AA	C
ATOM	38	CB	BASP	A	6	-4.756	-0.972	-17.114	0.50	9.33	AA	C
ATOM	39	CG	AASP	A	6	-4.673	-1.425	-15.620	0.50	9.57	AA	C
ATOM	40	CG	BASP	A	6	-4.476	-1.327	-15.644	0.50	12.98	AA	C
ATOM	41	OD1AASP	A	6	-5.328	-0.879	-14.735	0.50	9.04	AA	O	
ATOM	42	OD1BASP	A	6	-4.212	-0.417	-14.878	0.50	9.96	AA	O	
ATOM	43	OD2AASP	A	6	-3.907	-2.440	-15.419	0.50	12.73	AA	O	
ATOM	44	OD2BASP	A	6	-4.371	-2.541	-15.280	0.50	17.94	AA	O	
ATOM	45	C	ASP	A	6	-5.831	0.511	-18.748	1.00	9.51	AA	C
ATOM	46	O	ASP	A	6	-6.126	-0.288	-19.639	1.00	11.28	AA	O
ATOM	47	N	ILE	A	7	-5.545	1.760	-18.933	1.00	8.82	AA	N
ATOM	48	CA	ILE	A	7	-5.528	2.351	-20.273	1.00	9.22	AA	C
ATOM	49	CB	ILE	A	7	-6.559	3.536	-20.323	1.00	10.25	AA	C
ATOM	50	CG1	ILE	A	7	-7.992	3.039	-20.077	1.00	11.65	AA	C
ATOM	51	CD1	ILE	A	7	-8.485	2.072	-21.193	1.00	15.31	AA	C
ATOM	52	CG2	ILE	A	7	-6.481	4.259	-21.713	1.00	12.08	AA	C

ATOM	53	C	ILE	A	7	-4.128	2.938	-20.511	1.00	9.24	AA	C
ATOM	54	O	ILE	A	7	-3.539	3.504	-19.543	1.00	9.25	AA	O
ATOM	55	N	MET	A	8	-3.617	2.808	-21.737	1.00	8.38	AA	N
ATOM	56	CA	MET	A	8	-2.265	3.382	-22.086	1.00	8.90	AA	C
ATOM	57	CB	MET	A	8	-1.356	2.243	-22.602	1.00	8.74	AA	C
ATOM	58	CG	MET	A	8	-0.914	1.275	-21.476	1.00	10.75	AA	C
ATOM	59	SD	MET	A	8	-0.165	-0.212	-22.186	1.00	12.05	AA	S
ATOM	60	CE	MET	A	8	-1.737	-1.051	-22.541	1.00	13.37	AA	C
ATOM	61	C	MET	A	8	-2.462	4.339	-23.261	1.00	9.25	AA	C
ATOM	62	O	MET	A	8	-3.188	3.932	-24.205	1.00	9.48	AA	O
ATOM	63	N	GLY	A	9	-1.803	5.487	-23.203	1.00	7.90	AA	N
ATOM	64	CA	GLY	A	9	-1.708	6.324	-24.424	1.00	8.65	AA	C
ATOM	65	C	GLY	A	9	-0.230	6.496	-24.710	1.00	8.65	AA	C
ATOM	66	O	GLY	A	9	0.568	6.796	-23.826	1.00	9.37	AA	O
ATOM	67	N	ASN	A	10	0.150	6.191	-25.928	1.00	9.92	AA	N
ATOM	68	CA	ASN	A	10	1.573	6.142	-26.251	1.00	8.91	AA	C
ATOM	69	CB	ASN	A	10	1.910	4.741	-26.801	1.00	11.13	AA	C
ATOM	70	CG	ASN	A	10	1.277	4.484	-28.131	1.00	11.35	AA	C
ATOM	71	OD1	ASN	A	10	0.832	5.388	-28.835	1.00	11.46	AA	O
ATOM	72	ND2	ASN	A	10	1.272	3.226	-28.542	1.00	12.42	AA	N
ATOM	73	C	ASN	A	10	2.012	7.327	-27.128	1.00	10.80	AA	C
ATOM	74	O	ASN	A	10	1.271	8.307	-27.350	1.00	10.17	AA	O
ATOM	75	N	ASP	A	11	3.228	7.241	-27.668	1.00	11.00	AA	N
ATOM	76	CA	ASP	A	11	3.775	8.334	-28.499	1.00	9.31	AA	C
ATOM	77	CB	ASP	A	11	5.306	8.253	-28.596	1.00	13.14	AA	C
ATOM	78	CG	ASP	A	11	6.013	8.769	-27.359	1.00	13.84	AA	C
ATOM	79	OD1	ASP	A	11	5.469	9.368	-26.398	1.00	14.47	AA	O
ATOM	80	OD2	ASP	A	11	7.257	8.486	-27.283	1.00	17.89	AA	O
ATOM	81	C	ASP	A	11	3.267	8.307	-29.913	1.00	11.44	AA	C
ATOM	82	O	ASP	A	11	3.595	9.277	-30.664	1.00	12.98	AA	O
ATOM	83	N	GLN	A	12	2.382	7.346	-30.272	1.00	10.46	AA	N
ATOM	84	CA	GLN	A	12	1.761	7.292	-31.623	1.00	12.35	AA	C
ATOM	85	CB	GLN	A	12	1.829	5.899	-32.301	1.00	12.98	AA	C
ATOM	86	CG	GLN	A	12	3.103	5.162	-32.061	1.00	18.78	AA	C
ATOM	87	CD	GLN	A	12	4.212	5.971	-32.465	1.00	15.60	AA	C
ATOM	88	OE1	GLN	A	12	4.179	6.578	-33.620	1.00	18.21	AA	O
ATOM	89	NE2	GLN	A	12	5.302	6.007	-31.615	1.00	20.59	AA	N
ATOM	90	C	GLN	A	12	0.360	7.736	-31.530	1.00	11.43	AA	C
ATOM	91	O	GLN	A	12	-0.389	7.459	-32.394	1.00	13.22	AA	O
ATOM	92	N	MET	A	13	0.039	8.478	-30.473	1.00	11.20	AA	N
ATOM	93	CA	MET	A	13	-1.260	9.182	-30.390	1.00	12.18	AA	C
ATOM	94	CB	MET	A	13	-1.442	10.217	-31.540	1.00	11.56	AA	C
ATOM	95	CG	MET	A	13	-2.478	11.280	-31.145	1.00	11.60	AA	C
ATOM	96	SD	MET	A	13	-2.812	12.425	-32.480	1.00	14.34	AA	S
ATOM	97	CE	MET	A	13	-4.021	11.488	-33.432	1.00	17.08	AA	C
ATOM	98	C	MET	A	13	-2.368	8.103	-30.447	1.00	11.56	AA	C
ATOM	99	O	MET	A	13	-3.347	8.255	-31.127	1.00	12.72	AA	O
ATOM	100	N	HIS	A	14	-2.239	7.028	-29.638	1.00	10.92	AA	N
ATOM	101	CA	HIS	A	14	-3.130	5.853	-29.720	1.00	11.19	AA	C
ATOM	102	CB	HIS	A	14	-2.306	4.753	-30.454	1.00	12.09	AA	C
ATOM	103	CG	HIS	A	14	-2.984	3.412	-30.508	1.00	10.92	AA	C
ATOM	104	ND1	HIS	A	14	-4.089	3.126	-31.308	1.00	14.18	AA	N
ATOM	105	CE1	HIS	A	14	-4.396	1.831	-31.191	1.00	10.41	AA	C
ATOM	106	NE2	HIS	A	14	-3.502	1.251	-30.385	1.00	8.12	AA	N
ATOM	107	CD2	HIS	A	14	-2.630	2.236	-29.932	1.00	10.16	AA	C

ATOM	108	C	HIS	A	14	-3.398	5.351	-28.291	1.00	10.85	AA	C
ATOM	109	O	HIS	A	14	-2.453	5.288	-27.449	1.00	10.61	AA	O
ATOM	110	N	PHE	A	15	-4.664	5.019	-28.021	1.00	9.42	AA	N
ATOM	111	CA	PHE	A	15	-5.036	4.130	-26.921	1.00	7.61	AA	C
ATOM	112	CB	PHE	A	15	-6.431	4.529	-26.363	1.00	9.65	AA	C
ATOM	113	CG	PHE	A	15	-6.546	5.857	-25.611	1.00	8.76	AA	C
ATOM	114	CD1	PHE	A	15	-5.671	6.181	-24.542	1.00	9.75	AA	C
ATOM	115	CE1	PHE	A	15	-5.885	7.389	-23.765	1.00	11.12	AA	C
ATOM	116	CZ	PHE	A	15	-6.983	8.216	-24.023	1.00	11.04	AA	C
ATOM	117	CE2	PHE	A	15	-7.788	7.894	-25.111	1.00	11.58	AA	C
ATOM	118	CD2	PHE	A	15	-7.598	6.713	-25.858	1.00	10.06	AA	C
ATOM	119	C	PHE	A	15	-5.261	2.758	-27.464	1.00	10.18	AA	C
ATOM	120	O	PHE	A	15	-5.905	2.612	-28.524	1.00	10.88	AA	O
ATOM	121	N	HIS	A	16	-4.771	1.721	-26.739	1.00	9.74	AA	N
ATOM	122	CA	HIS	A	16	-5.025	0.397	-27.259	1.00	11.19	AA	C
ATOM	123	CB	HIS	A	16	-4.058	-0.666	-26.660	1.00	12.48	AA	C
ATOM	124	CG	HIS	A	16	-2.606	-0.391	-26.970	1.00	11.17	AA	C
ATOM	125	ND1	HIS	A	16	-2.076	-0.427	-28.248	1.00	11.61	AA	N
ATOM	126	CE1	HIS	A	16	-0.768	-0.130	-28.211	1.00	11.53	AA	C
ATOM	127	NE2	HIS	A	16	-0.469	0.191	-26.962	1.00	12.97	AA	N
ATOM	128	CD2	HIS	A	16	-1.574	-0.014	-26.161	1.00	13.81	AA	C
ATOM	129	C	HIS	A	16	-6.472	-0.078	-27.138	1.00	13.30	AA	C
ATOM	130	O	HIS	A	16	-7.017	-0.857	-28.027	1.00	15.93	AA	O
ATOM	131	N	THR	A	17	-7.154	0.524	-26.187	1.00	8.81	AA	N
ATOM	132	CA	THR	A	17	-8.517	0.164	-25.841	1.00	10.01	AA	C
ATOM	133	CB	THR	A	17	-8.768	0.478	-24.358	1.00	10.24	AA	C
ATOM	134	OG1	THR	A	17	-7.949	-0.411	-23.563	1.00	12.83	AA	O
ATOM	135	CG2	THR	A	17	-10.213	0.299	-23.985	1.00	12.49	AA	C
ATOM	136	C	THR	A	17	-9.492	1.033	-26.685	1.00	10.67	AA	C
ATOM	137	O	THR	A	17	-9.373	2.249	-26.746	1.00	10.41	AA	O
ATOM	138	N	ASN	A	18	-10.412	0.351	-27.375	1.00	9.41	AA	N
ATOM	139	CA	ASN	A	18	-11.456	1.053	-28.158	1.00	10.47	AA	C
ATOM	140	CB	ASN	A	18	-11.647	0.356	-29.534	1.00	9.79	AA	C
ATOM	141	CG	ASN	A	18	-12.073	-1.134	-29.407	1.00	12.97	AA	C
ATOM	142	OD1	ASN	A	18	-11.310	-1.985	-28.884	1.00	11.16	AA	O
ATOM	143	ND2	ASN	A	18	-13.213	-1.512	-30.015	1.00	10.31	AA	N
ATOM	144	C	ASN	A	18	-12.814	1.158	-27.447	1.00	9.36	AA	C
ATOM	145	O	ASN	A	18	-13.590	2.042	-27.864	1.00	10.44	AA	O
ATOM	146	N	ALA	A	19	-13.048	0.346	-26.453	1.00	9.14	AA	N
ATOM	147	CA	ALA	A	19	-14.414	0.272	-25.852	1.00	8.31	AA	C
ATOM	148	CB	ALA	A	19	-15.322	-0.626	-26.633	1.00	9.28	AA	C
ATOM	149	C	ALA	A	19	-14.250	-0.174	-24.433	1.00	9.36	AA	C
ATOM	150	O	ALA	A	19	-13.526	-1.115	-24.178	1.00	11.51	AA	O
ATOM	151	N	ILE	A	20	-14.952	0.478	-23.533	1.00	9.94	AA	N
ATOM	152	CA	ILE	A	20	-14.980	0.143	-22.092	1.00	10.77	AA	C
ATOM	153	CB	ILE	A	20	-14.495	1.322	-21.271	1.00	10.67	AA	C
ATOM	154	CG1	ILE	A	20	-13.011	1.587	-21.602	1.00	9.87	AA	C
ATOM	155	CD1	ILE	A	20	-12.461	2.946	-21.012	1.00	13.55	AA	C
ATOM	156	CG2	ILE	A	20	-14.732	1.025	-19.737	1.00	11.57	AA	C
ATOM	157	C	ILE	A	20	-16.417	-0.209	-21.758	1.00	10.47	AA	C
ATOM	158	O	ILE	A	20	-17.346	0.544	-22.105	1.00	11.86	AA	O
ATOM	159	N	THR	A	21	-16.567	-1.308	-21.023	1.00	11.58	AA	N
ATOM	160	CA	THR	A	21	-17.910	-1.711	-20.531	1.00	13.46	AA	C
ATOM	161	CB	THR	A	21	-18.180	-3.151	-20.910	1.00	15.19	AA	C
ATOM	162	OG1	THR	A	21	-18.242	-3.225	-22.349	1.00	17.57	AA	O

ATOM	163	CG2 THR A 21	-19.488	-3.654	-20.289	1.00	15.26	AA	C
ATOM	164	C THR A 21	-17.833	-1.680	-18.996	1.00	13.88	AA	C
ATOM	165	O THR A 21	-16.982	-2.381	-18.398	1.00	16.67	AA	O
ATOM	166	N VAL A 22	-18.662	-0.823	-18.377	1.00	10.28	AA	N
ATOM	167	CA VAL A 22	-18.691	-0.688	-16.887	1.00	10.41	AA	C
ATOM	168	CB VAL A 22	-19.015	0.705	-16.480	1.00	10.02	AA	C
ATOM	169	CG1 VAL A 22	-19.113	0.779	-14.912	1.00	12.93	AA	C
ATOM	170	CG2 VAL A 22	-17.845	1.704	-17.056	1.00	12.74	AA	C
ATOM	171	C VAL A 22	-19.775	-1.666	-16.371	1.00	10.68	AA	C
ATOM	172	O VAL A 22	-20.946	-1.568	-16.852	1.00	12.04	AA	O
ATOM	173	N ASP A 23	-19.412	-2.582	-15.481	1.00	11.13	AA	N
ATOM	174	CA ASP A 23	-20.385	-3.580	-14.916	1.00	13.43	AA	C
ATOM	175	CB ASP A 23	-19.567	-4.682	-14.197	1.00	15.01	AA	C
ATOM	176	CG ASP A 23	-20.392	-5.755	-13.556	1.00	21.17	AA	C
ATOM	177	OD1 ASP A 23	-21.614	-5.577	-13.280	1.00	24.78	AA	O
ATOM	178	OD2 ASP A 23	-19.715	-6.793	-13.346	1.00	29.18	AA	O
ATOM	179	C ASP A 23	-21.336	-2.790	-13.973	1.00	11.96	AA	C
ATOM	180	O ASP A 23	-20.846	-2.116	-13.049	1.00	11.65	AA	O
ATOM	181	N LYS A 24	-22.662	-2.839	-14.198	1.00	14.49	AA	N
ATOM	182	CA LYS A 24	-23.565	-1.998	-13.383	1.00	15.75	AA	C
ATOM	183	CB LYS A 24	-25.010	-1.917	-13.951	1.00	17.45	AA	C
ATOM	184	CG LYS A 24	-25.633	-3.231	-14.190	1.00	17.66	AA	C
ATOM	185	CD LYS A 24	-27.033	-3.048	-14.792	1.00	20.41	AA	C
ATOM	186	CE LYS A 24	-27.731	-4.375	-14.961	1.00	18.91	AA	C
ATOM	187	NZ LYS A 24	-26.787	-5.466	-15.518	1.00	21.24	AA	N
ATOM	188	C LYS A 24	-23.618	-2.452	-11.907	1.00	14.86	AA	C
ATOM	189	O LYS A 24	-24.167	-1.715	-11.018	1.00	15.50	AA	O
ATOM	190	N SER A 25	-23.066	-3.622	-11.615	1.00	12.95	AA	N
ATOM	191	CA SER A 25	-22.942	-4.045	-10.230	1.00	14.87	AA	C
ATOM	192	CB SER A 25	-22.767	-5.565	-10.188	1.00	14.89	AA	C
ATOM	193	OG SER A 25	-21.507	-5.927	-10.572	1.00	16.79	AA	O
ATOM	194	C SER A 25	-21.817	-3.321	-9.500	1.00	12.12	AA	C
ATOM	195	O SER A 25	-21.756	-3.321	-8.240	1.00	13.48	AA	O
ATOM	196	N CYS A 26	-20.910	-2.681	-10.229	1.00	12.29	AA	N
ATOM	197	CA CYS A 26	-19.872	-1.849	-9.580	1.00	10.69	AA	C
ATOM	198	CB CYS A 26	-18.815	-1.425	-10.623	1.00	9.78	AA	C
ATOM	199	SG CYS A 26	-17.895	-2.794	-11.296	1.00	12.00	AA	S
ATOM	200	C CYS A 26	-20.453	-0.607	-8.975	1.00	10.46	AA	C
ATOM	201	O CYS A 26	-21.173	0.099	-9.626	1.00	13.78	AA	O
ATOM	202	N LYS A 27	-20.110	-0.279	-7.728	1.00	9.23	AA	N
ATOM	203	CA LYS A 27	-20.474	1.026	-7.106	1.00	10.83	AA	C
ATOM	204	CB LYS A 27	-20.326	0.981	-5.574	1.00	11.53	AA	C
ATOM	205	CG LYS A 27	-21.357	0.090	-4.880	1.00	21.53	AA	C
ATOM	206	CD LYS A 27	-22.828	0.591	-4.986	1.00	28.18	AA	C
ATOM	207	CE LYS A 27	-23.776	-0.405	-4.229	1.00	29.85	AA	C
ATOM	208	NZ LYS A 27	-23.131	-1.808	-4.318	1.00	30.84	AA	N
ATOM	209	C LYS A 27	-19.655	2.172	-7.670	1.00	9.71	AA	C
ATOM	210	O LYS A 27	-20.150	3.265	-7.922	1.00	10.17	AA	O
ATOM	211	N GLN A 28	-18.392	1.890	-7.893	1.00	9.61	AA	N
ATOM	212	CA GLN A 28	-17.428	2.907	-8.427	1.00	8.56	AA	C
ATOM	213	CB GLN A 28	-16.522	3.400	-7.345	1.00	10.04	AA	C
ATOM	214	CG GLN A 28	-17.355	4.170	-6.286	1.00	11.30	AA	C
ATOM	215	CD GLN A 28	-16.453	5.075	-5.491	1.00	17.23	AA	C
ATOM	216	OE1 GLN A 28	-15.561	4.601	-4.782	1.00	17.87	AA	O
ATOM	217	NE2 GLN A 28	-16.645	6.397	-5.627	1.00	21.61	AA	N

ATOM	218	C	GLN	A	28	-16.594	2.218	-9.492	1.00	9.27	AA	C
ATOM	219	O	GLN	A	28	-16.478	0.941	-9.483	1.00	10.75	AA	O
ATOM	220	N	PHE	A	29	-16.127	2.988	-10.426	1.00	8.27	AA	N
ATOM	221	CA	PHE	A	29	-15.269	2.466	-11.511	1.00	8.63	AA	C
ATOM	222	CB	PHE	A	29	-16.006	2.580	-12.816	1.00	11.26	AA	C
ATOM	223	CG	PHE	A	29	-15.382	1.856	-13.937	1.00	9.97	AA	C
ATOM	224	CD1	PHE	A	29	-15.413	0.460	-14.038	1.00	8.09	AA	C
ATOM	225	CE1	PHE	A	29	-14.786	-0.220	-15.136	1.00	12.25	AA	C
ATOM	226	CZ	PHE	A	29	-14.108	0.512	-16.084	1.00	9.12	AA	C
ATOM	227	CE2	PHE	A	29	-13.984	1.880	-15.957	1.00	9.79	AA	C
ATOM	228	CD2	PHE	A	29	-14.667	2.569	-14.892	1.00	9.92	AA	C
ATOM	229	C	PHE	A	29	-14.018	3.292	-11.543	1.00	9.00	AA	C
ATOM	230	O	PHE	A	29	-14.026	4.485	-11.299	1.00	9.00	AA	O
ATOM	231	N	THR	A	30	-12.914	2.604	-11.792	1.00	8.32	AA	N
ATOM	232	CA	THR	A	30	-11.566	3.237	-11.753	1.00	8.44	AA	C
ATOM	233	CB	THR	A	30	-10.714	2.623	-10.674	1.00	9.14	AA	C
ATOM	234	OG1	THR	A	30	-11.384	2.764	-9.396	1.00	11.09	AA	O
ATOM	235	CG2	THR	A	30	-9.316	3.270	-10.568	1.00	10.54	AA	C
ATOM	236	C	THR	A	30	-10.875	2.993	-13.076	1.00	9.15	AA	C
ATOM	237	O	THR	A	30	-10.751	1.889	-13.610	1.00	9.05	AA	O
ATOM	238	N	VAL	A	31	-10.335	4.103	-13.628	1.00	9.02	AA	N
ATOM	239	CA	VAL	A	31	-9.495	4.070	-14.817	1.00	8.63	AA	C
ATOM	240	CB	VAL	A	31	-9.966	5.086	-15.912	1.00	10.15	AA	C
ATOM	241	CG1	VAL	A	31	-8.966	5.216	-17.069	1.00	13.06	AA	C
ATOM	242	CG2	VAL	A	31	-11.390	4.700	-16.333	1.00	12.07	AA	C
ATOM	243	C	VAL	A	31	-8.092	4.395	-14.354	1.00	8.77	AA	C
ATOM	244	O	VAL	A	31	-7.822	5.473	-13.766	1.00	9.13	AA	O
ATOM	245	N	ASN	A	32	-7.129	3.524	-14.721	1.00	8.46	AA	N
ATOM	246	CA	ASN	A	32	-5.718	3.716	-14.375	1.00	8.57	AA	C
ATOM	247	CB	ASN	A	32	-5.144	2.467	-13.722	1.00	7.88	AA	C
ATOM	248	CG	ASN	A	32	-5.987	2.085	-12.501	1.00	7.87	AA	C
ATOM	249	OD1	ASN	A	32	-6.030	2.804	-11.490	1.00	9.53	AA	O
ATOM	250	ND2	ASN	A	32	-6.576	0.902	-12.580	1.00	10.21	AA	N
ATOM	251	C	ASN	A	32	-5.025	4.011	-15.721	1.00	8.78	AA	C
ATOM	252	O	ASN	A	32	-4.769	3.088	-16.513	1.00	9.14	AA	O
ATOM	253	N	LEU	A	33	-4.623	5.280	-15.914	1.00	7.98	AA	N
ATOM	254	CA	LEU	A	33	-3.966	5.696	-17.158	1.00	8.67	AA	C
ATOM	255	CB	LEU	A	33	-4.423	7.098	-17.508	1.00	9.68	AA	C
ATOM	256	CG	LEU	A	33	-3.940	7.717	-18.803	1.00	9.52	AA	C
ATOM	257	CD1	LEU	A	33	-4.412	6.952	-20.033	1.00	12.48	AA	C
ATOM	258	CD2	LEU	A	33	-4.316	9.167	-18.922	1.00	10.70	AA	C
ATOM	259	C	LEU	A	33	-2.472	5.729	-17.019	1.00	8.45	AA	C
ATOM	260	O	LEU	A	33	-1.950	6.175	-16.012	1.00	9.07	AA	O
ATOM	261	N	SER	A	34	-1.754	5.271	-18.044	1.00	7.84	AA	N
ATOM	262	CA	SER	A	34	-0.286	5.367	-18.108	1.00	9.06	AA	C
ATOM	263	CB	SER	A	34	0.417	4.036	-17.707	1.00	9.06	AA	C
ATOM	264	OG	SER	A	34	0.127	3.059	-18.686	1.00	10.25	AA	O
ATOM	265	C	SER	A	34	0.132	5.857	-19.464	1.00	10.21	AA	C
ATOM	266	O	SER	A	34	-0.677	5.860	-20.424	1.00	10.53	AA	O
ATOM	267	N	HIS	A	35	1.380	6.357	-19.472	1.00	10.92	AA	N
ATOM	268	CA	HIS	A	35	1.922	6.955	-20.733	1.00	10.47	AA	C
ATOM	269	CB	HIS	A	35	2.097	8.453	-20.564	1.00	11.96	AA	C
ATOM	270	CG	HIS	A	35	2.454	9.150	-21.849	1.00	11.40	AA	C
ATOM	271	ND1	HIS	A	35	3.351	8.655	-22.814	1.00	12.57	AA	N
ATOM	272	CE1	HIS	A	35	3.410	9.530	-23.826	1.00	11.20	AA	C

ATOM	273	NE2 HIS A 35	2.622	10.576	-23.530	1.00	10.28	AA	N
ATOM	274	CD2 HIS A 35	1.988	10.333	-22.311	1.00	9.41	AA	C
ATOM	275	C HIS A 35	3.260	6.279	-20.958	1.00	12.00	AA	C
ATOM	276	O HIS A 35	4.306	6.766	-20.502	1.00	15.23	AA	O
ATOM	277	N PRO A 36	3.287	5.187	-21.721	1.00	10.94	AA	N
ATOM	278	CA PRO A 36	4.566	4.509	-22.018	1.00	14.36	AA	C
ATOM	279	CB PRO A 36	4.115	3.137	-22.509	1.00	17.31	AA	C
ATOM	280	CG PRO A 36	2.762	3.270	-22.989	1.00	17.68	AA	C
ATOM	281	CD PRO A 36	2.138	4.349	-22.115	1.00	12.48	AA	C
ATOM	282	C PRO A 36	5.228	5.289	-23.124	1.00	17.18	AA	C
ATOM	283	O PRO A 36	4.600	6.256	-23.658	1.00	20.62	AA	O
ATOM	284	N GLY A 37	6.443	4.839	-23.501	1.00	19.22	AA	N
ATOM	285	CA GLY A 37	7.159	5.820	-24.449	1.00	22.44	AA	C
ATOM	286	C GLY A 37	7.623	7.224	-23.874	1.00	22.56	AA	C
ATOM	287	O GLY A 37	7.527	7.494	-22.649	1.00	22.71	AA	O
ATOM	288	N ASN A 38	8.082	8.137	-24.734	1.00	16.77	AA	N
ATOM	289	CA ASN A 38	9.150	9.003	-24.274	1.00	15.33	AA	C
ATOM	290	CB ASN A 38	10.449	8.715	-25.040	1.00	14.98	AA	C
ATOM	291	CG ASN A 38	10.885	7.242	-24.968	1.00	22.39	AA	C
ATOM	292	OD1 ASN A 38	11.218	6.642	-26.005	1.00	33.69	AA	O
ATOM	293	ND2 ASN A 38	10.915	6.662	-23.757	1.00	27.41	AA	N
ATOM	294	C ASN A 38	8.955	10.497	-24.297	1.00	14.63	AA	C
ATOM	295	O ASN A 38	9.801	11.219	-23.786	1.00	15.81	AA	O
ATOM	296	N LEU A 39	7.819	10.967	-24.828	1.00	13.03	AA	N
ATOM	297	CA LEU A 39	7.634	12.425	-25.006	1.00	13.24	AA	C
ATOM	298	CB LEU A 39	6.869	12.739	-26.299	1.00	14.89	AA	C
ATOM	299	CG LEU A 39	7.618	12.316	-27.551	1.00	12.62	AA	C
ATOM	300	CD1 LEU A 39	6.643	12.409	-28.721	1.00	18.12	AA	C
ATOM	301	CD2 LEU A 39	8.735	13.315	-27.821	1.00	16.41	AA	C
ATOM	302	C LEU A 39	6.943	13.079	-23.788	1.00	9.47	AA	C
ATOM	303	O LEU A 39	6.030	12.479	-23.201	1.00	12.60	AA	O
ATOM	304	N PRO A 40	7.350	14.284	-23.458	1.00	11.73	AA	N
ATOM	305	CA PRO A 40	6.769	14.959	-22.313	1.00	11.74	AA	C
ATOM	306	CB PRO A 40	7.709	16.108	-22.059	1.00	15.39	AA	C
ATOM	307	CG PRO A 40	8.241	16.407	-23.310	1.00	14.79	AA	C
ATOM	308	CD PRO A 40	8.375	15.119	-24.086	1.00	13.78	AA	C
ATOM	309	C PRO A 40	5.312	15.510	-22.591	1.00	12.24	AA	C
ATOM	310	O PRO A 40	4.864	15.572	-23.772	1.00	10.79	AA	O
ATOM	311	N LYS A 41	4.637	15.923	-21.526	1.00	10.03	AA	N
ATOM	312	CA LYS A 41	3.255	16.275	-21.600	1.00	10.77	AA	C
ATOM	313	CB LYS A 41	2.570	16.352	-20.241	1.00	14.89	AA	C
ATOM	314	CG LYS A 41	2.861	17.479	-19.474	1.00	17.49	AA	C
ATOM	315	CD LYS A 41	1.947	17.387	-18.195	1.00	18.06	AA	C
ATOM	316	CE LYS A 41	2.534	18.330	-17.264	1.00	16.95	AA	C
ATOM	317	NZ LYS A 41	1.872	18.344	-15.929	1.00	13.11	AA	N
ATOM	318	C LYS A 41	3.006	17.491	-22.448	1.00	10.88	AA	C
ATOM	319	O LYS A 41	1.908	17.621	-22.958	1.00	10.38	AA	O
ATOM	320	N ASN A 42	3.986	18.398	-22.564	1.00	9.40	AA	N
ATOM	321	CA ASN A 42	3.705	19.614	-23.333	1.00	11.34	AA	C
ATOM	322	CB ASN A 42	4.567	20.848	-22.925	1.00	12.46	AA	C
ATOM	323	CG ASN A 42	5.983	20.677	-23.154	1.00	16.90	AA	C
ATOM	324	OD1 ASN A 42	6.443	19.674	-23.618	1.00	14.70	AA	O
ATOM	325	ND2 ASN A 42	6.738	21.744	-22.801	1.00	20.08	AA	N
ATOM	326	C ASN A 42	3.689	19.365	-24.842	1.00	9.80	AA	C
ATOM	327	O ASN A 42	3.207	20.250	-25.598	1.00	11.29	AA	O

ATOM	328	N	VAL	A	43	4.136	18.220	-25.307	1.00	10.48	AA	N
ATOM	329	CA	VAL	A	43	4.116	17.890	-26.750	1.00	10.84	AA	C
ATOM	330	CB	VAL	A	43	5.470	17.890	-27.459	1.00	9.82	AA	C
ATOM	331	CG1	VAL	A	43	6.162	19.296	-27.369	1.00	12.44	AA	C
ATOM	332	CG2	VAL	A	43	6.317	16.763	-26.921	1.00	11.23	AA	C
ATOM	333	C	VAL	A	43	3.359	16.588	-27.072	1.00	8.37	AA	C
ATOM	334	O	VAL	A	43	3.063	16.300	-28.201	1.00	9.76	AA	O
ATOM	335	N	MET	A	44	3.095	15.798	-26.023	1.00	9.30	AA	N
ATOM	336	CA	MET	A	44	2.431	14.509	-26.234	1.00	9.57	AA	C
ATOM	337	CB	MET	A	44	3.428	13.383	-26.658	1.00	10.78	AA	C
ATOM	338	CG	MET	A	44	2.843	12.100	-27.175	1.00	13.29	AA	C
ATOM	339	SD	MET	A	44	1.642	12.219	-28.526	1.00	11.99	AA	S
ATOM	340	CE	MET	A	44	2.732	12.697	-29.870	1.00	12.13	AA	C
ATOM	341	C	MET	A	44	1.634	14.122	-24.987	1.00	9.73	AA	C
ATOM	342	O	MET	A	44	1.556	12.935	-24.604	1.00	11.12	AA	O
ATOM	343	N	GLY	A	45	1.043	15.123	-24.349	1.00	9.07	AA	N
ATOM	344	CA	GLY	A	45	0.231	14.870	-23.186	1.00	9.38	AA	C
ATOM	345	C	GLY	A	45	-1.009	14.036	-23.509	1.00	8.83	AA	C
ATOM	346	O	GLY	A	45	-1.633	14.200	-24.577	1.00	9.39	AA	O
ATOM	347	N	HIS	A	46	-1.385	13.166	-22.538	1.00	7.59	AA	N
ATOM	348	CA	HIS	A	46	-2.658	12.464	-22.646	1.00	8.18	AA	C
ATOM	349	CB	HIS	A	46	-2.468	10.941	-22.983	1.00	9.72	AA	C
ATOM	350	CG	HIS	A	46	-1.684	10.677	-24.227	1.00	7.90	AA	C
ATOM	351	ND1	HIS	A	46	-1.949	11.275	-25.466	1.00	8.83	AA	N
ATOM	352	CE1	HIS	A	46	-1.086	10.769	-26.356	1.00	9.73	AA	C
ATOM	353	NE2	HIS	A	46	-0.316	9.870	-25.754	1.00	9.82	AA	N
ATOM	354	CD2	HIS	A	46	-0.670	9.799	-24.410	1.00	9.87	AA	C
ATOM	355	C	HIS	A	46	-3.439	12.569	-21.371	1.00	9.04	AA	C
ATOM	356	O	HIS	A	46	-2.872	12.492	-20.256	1.00	9.64	AA	O
ATOM	357	N	ASN	A	47	-4.745	12.687	-21.516	1.00	8.45	AA	N
ATOM	358	CA	ASN	A	47	-5.638	12.450	-20.416	1.00	8.32	AA	C
ATOM	359	CB	ASN	A	47	-6.225	13.800	-19.889	1.00	9.57	AA	C
ATOM	360	CG	ASN	A	47	-6.919	14.648	-20.957	1.00	9.54	AA	C
ATOM	361	OD1	ASN	A	47	-7.191	14.175	-22.043	1.00	9.34	AA	O
ATOM	362	ND2	ASN	A	47	-7.206	15.939	-20.657	1.00	10.37	AA	N
ATOM	363	C	ASN	A	47	-6.716	11.486	-20.862	1.00	9.54	AA	C
ATOM	364	O	ASN	A	47	-6.811	11.030	-22.032	1.00	9.29	AA	O
ATOM	365	N	TRP	A	48	-7.531	11.043	-19.903	1.00	9.05	AA	N
ATOM	366	CA	TRP	A	48	-8.713	10.214	-20.208	1.00	8.25	AA	C
ATOM	367	CB	TRP	A	48	-8.561	8.870	-19.480	1.00	8.60	AA	C
ATOM	368	CG	TRP	A	48	-9.708	7.904	-19.795	1.00	8.47	AA	C
ATOM	369	CD1	TRP	A	48	-9.749	6.946	-20.755	1.00	8.28	AA	C
ATOM	370	NE1	TRP	A	48	-10.959	6.230	-20.714	1.00	9.07	AA	N
ATOM	371	CE2	TRP	A	48	-11.705	6.746	-19.662	1.00	8.04	AA	C
ATOM	372	CD2	TRP	A	48	-10.981	7.809	-19.082	1.00	9.16	AA	C
ATOM	373	CE3	TRP	A	48	-11.512	8.463	-17.935	1.00	10.25	AA	C
ATOM	374	CZ3	TRP	A	48	-12.790	8.092	-17.520	1.00	8.59	AA	C
ATOM	375	CH2	TRP	A	48	-13.446	7.077	-18.146	1.00	10.13	AA	C
ATOM	376	CZ2	TRP	A	48	-12.973	6.432	-19.234	1.00	8.99	AA	C
ATOM	377	C	TRP	A	48	-9.906	11.005	-19.663	1.00	8.65	AA	C
ATOM	378	O	TRP	A	48	-9.944	11.318	-18.439	1.00	9.23	AA	O
ATOM	379	N	VAL	A	49	-10.813	11.370	-20.553	1.00	9.20	AA	N
ATOM	380	CA	VAL	A	49	-11.929	12.254	-20.213	1.00	9.01	AA	C
ATOM	381	CB	VAL	A	49	-11.734	13.620	-20.994	1.00	9.05	AA	C
ATOM	382	CG1	VAL	A	49	-12.912	14.642	-20.642	1.00	10.87	AA	C

ATOM	383	CG2 VAL A 49	-10.391	14.247	-20.571	1.00	10.08	AA	C
ATOM	384	C VAL A 49	-13.201	11.570	-20.661	1.00	8.46	AA	C
ATOM	385	O VAL A 49	-13.221	11.023	-21.799	1.00	9.17	AA	O
ATOM	386	N LEU A 50	-14.283	11.625	-19.841	1.00	8.53	AA	N
ATOM	387	CA LEU A 50	-15.535	10.954	-20.157	1.00	9.13	AA	C
ATOM	388	CB LEU A 50	-15.825	10.008	-18.983	1.00	10.01	AA	C
ATOM	389	CG LEU A 50	-17.131	9.144	-19.128	1.00	9.71	AA	C
ATOM	390	CD1 LEU A 50	-17.060	8.202	-20.241	1.00	12.74	AA	C
ATOM	391	CD2 LEU A 50	-17.335	8.367	-17.821	1.00	11.39	AA	C
ATOM	392	C LEU A 50	-16.631	11.973	-20.253	1.00	9.86	AA	C
ATOM	393	O LEU A 50	-16.749	12.824	-19.365	1.00	9.49	AA	O
ATOM	394	N SER A 51	-17.424	11.864	-21.300	1.00	8.28	AA	N
ATOM	395	CA SER A 51	-18.647	12.712	-21.420	1.00	7.55	AA	C
ATOM	396	CB SER A 51	-18.367	14.037	-22.204	1.00	9.61	AA	C
ATOM	397	OG SER A 51	-18.109	13.800	-23.594	1.00	9.96	AA	O
ATOM	398	C SER A 51	-19.680	11.896	-22.149	1.00	8.69	AA	C
ATOM	399	O SER A 51	-19.393	10.828	-22.727	1.00	9.29	AA	O
ATOM	400	N THR A 52	-20.887	12.451	-22.259	1.00	8.90	AA	N
ATOM	401	CA THR A 52	-21.832	11.910	-23.256	1.00	10.68	AA	C
ATOM	402	CB THR A 52	-23.200	12.538	-23.209	1.00	9.81	AA	C
ATOM	403	OG1 THR A 52	-23.043	13.915	-23.540	1.00	11.55	AA	O
ATOM	404	CG2 THR A 52	-23.822	12.345	-21.817	1.00	10.60	AA	C
ATOM	405	C THR A 52	-21.276	12.093	-24.684	1.00	11.09	AA	C
ATOM	406	O THR A 52	-20.430	12.962	-24.909	1.00	11.23	AA	O
ATOM	407	N ALA A 53	-21.728	11.270	-25.612	1.00	11.12	AA	N
ATOM	408	CA ALA A 53	-21.321	11.478	-26.999	1.00	13.00	AA	C
ATOM	409	CB ALA A 53	-21.967	10.365	-27.883	1.00	15.58	AA	C
ATOM	410	C ALA A 53	-21.760	12.859	-27.494	1.00	12.44	AA	C
ATOM	411	O ALA A 53	-21.022	13.501	-28.240	1.00	13.56	AA	O
ATOM	412	N ALA A 54	-22.940	13.322	-27.088	1.00	11.86	AA	N
ATOM	413	CA ALA A 54	-23.364	14.635	-27.498	1.00	11.31	AA	C
ATOM	414	CB ALA A 54	-24.781	14.931	-26.968	1.00	12.74	AA	C
ATOM	415	C ALA A 54	-22.471	15.777	-27.079	1.00	10.03	AA	C
ATOM	416	O ALA A 54	-22.343	16.789	-27.807	1.00	13.10	AA	O
ATOM	417	N ASP A 55	-21.782	15.595	-25.948	1.00	9.66	AA	N
ATOM	418	CA ASP A 55	-20.935	16.700	-25.386	1.00	9.50	AA	C
ATOM	419	CB ASP A 55	-20.976	16.672	-23.899	1.00	9.37	AA	C
ATOM	420	CG ASP A 55	-22.262	17.266	-23.316	1.00	11.67	AA	C
ATOM	421	OD1 ASP A 55	-22.857	18.121	-24.053	1.00	14.34	AA	O
ATOM	422	OD2 ASP A 55	-22.616	16.905	-22.170	1.00	11.74	AA	O
ATOM	423	C ASP A 55	-19.439	16.628	-25.849	1.00	9.73	AA	C
ATOM	424	O ASP A 55	-18.716	17.556	-25.545	1.00	10.86	AA	O
ATOM	425	N MET A 56	-19.008	15.491	-26.418	1.00	9.18	AA	N
ATOM	426	CA MET A 56	-17.574	15.274	-26.674	1.00	9.56	AA	C
ATOM	427	CB MET A 56	-17.326	13.955	-27.384	1.00	11.57	AA	C
ATOM	428	CG MET A 56	-15.880	13.822	-27.821	1.00	10.70	AA	C
ATOM	429	SD MET A 56	-15.561	12.215	-28.600	1.00	12.60	AA	S
ATOM	430	CE MET A 56	-13.857	12.584	-29.120	1.00	14.76	AA	C
ATOM	431	C MET A 56	-16.992	16.401	-27.508	1.00	10.50	AA	C
ATOM	432	O MET A 56	-15.912	16.927	-27.139	1.00	9.41	AA	O
ATOM	433	N GLN A 57	-17.608	16.809	-28.620	1.00	10.32	AA	N
ATOM	434	CA AGLN A 57	-16.933	17.805	-29.470	0.50	10.79	AA	C
ATOM	435	CA BGLN A 57	-16.946	17.752	-29.479	0.50	10.35	AA	C
ATOM	436	CB AGLN A 57	-17.639	18.084	-30.807	0.50	12.75	AA	C
ATOM	437	CB BGLN A 57	-17.723	17.861	-30.809	0.50	12.00	AA	C

ATOM	438	CG	AGLN	A	57	-16.822	19.043	-31.735	0.50	13.64	AA	C
ATOM	439	CG	BGLN	A	57	-17.689	16.617	-31.812	0.50	9.06	AA	C
ATOM	440	CD	AGLN	A	57	-15.457	18.422	-32.119	0.50	19.29	AA	C
ATOM	441	CD	BGLN	A	57	-18.585	16.835	-33.053	0.50	14.60	AA	C
ATOM	442	OE1	AGLN	A	57	-15.437	17.504	-32.944	0.50	22.26	AA	O
ATOM	443	OE1	BGLN	A	57	-19.805	16.818	-32.952	0.50	17.99	AA	O
ATOM	444	NE2	AGLN	A	57	-14.321	18.909	-31.538	0.50	15.50	AA	N
ATOM	445	NE2	BGLN	A	57	-17.960	17.061	-34.223	0.50	14.10	AA	N
ATOM	446	C	GLN	A	57	-16.737	19.093	-28.751	1.00	9.55	AA	C
ATOM	447	O	GLN	A	57	-15.679	19.714	-28.890	1.00	10.26	AA	O
ATOM	448	N	GLY	A	58	-17.756	19.547	-28.012	1.00	9.61	AA	N
ATOM	449	CA	GLY	A	58	-17.601	20.822	-27.284	1.00	10.64	AA	C
ATOM	450	C	GLY	A	58	-16.565	20.713	-26.169	1.00	9.95	AA	C
ATOM	451	O	GLY	A	58	-15.801	21.621	-25.982	1.00	9.96	AA	O
ATOM	452	N	VAL	A	59	-16.473	19.589	-25.455	1.00	9.61	AA	N
ATOM	453	CA	VAL	A	59	-15.432	19.428	-24.437	1.00	9.50	AA	C
ATOM	454	CB	VAL	A	59	-15.609	18.069	-23.679	1.00	8.96	AA	C
ATOM	455	CG1	VAL	A	59	-14.372	17.814	-22.757	1.00	9.93	AA	C
ATOM	456	CG2	VAL	A	59	-16.889	18.096	-22.871	1.00	8.66	AA	C
ATOM	457	C	VAL	A	59	-14.021	19.484	-25.099	1.00	8.50	AA	C
ATOM	458	O	VAL	A	59	-13.098	20.155	-24.588	1.00	10.72	AA	O
ATOM	459	N	VAL	A	60	-13.843	18.827	-26.256	1.00	8.41	AA	N
ATOM	460	CA	VAL	A	60	-12.555	18.824	-26.966	1.00	9.02	AA	C
ATOM	461	CB	VAL	A	60	-12.617	17.829	-28.101	1.00	9.38	AA	C
ATOM	462	CG1	VAL	A	60	-11.404	18.042	-29.075	1.00	11.19	AA	C
ATOM	463	CG2	VAL	A	60	-12.651	16.447	-27.570	1.00	12.07	AA	C
ATOM	464	C	VAL	A	60	-12.185	20.236	-27.454	1.00	8.91	AA	C
ATOM	465	O	VAL	A	60	-11.087	20.743	-27.223	1.00	9.13	AA	O
ATOM	466	N	THR	A	61	-13.107	20.929	-28.110	1.00	10.58	AA	N
ATOM	467	CA	ATHR	A	61	-12.894	22.295	-28.669	0.50	10.27	AA	C
ATOM	468	CA	BTHR	A	61	-12.758	22.228	-28.699	0.50	11.06	AA	C
ATOM	469	CB	ATHR	A	61	-14.154	22.799	-29.379	0.50	9.43	AA	C
ATOM	470	CB	BTHR	A	61	-13.748	22.615	-29.811	0.50	11.57	AA	C
ATOM	471	OG1	ATHR	A	61	-14.353	21.990	-30.525	0.50	11.01	AA	O
ATOM	472	OG1	BTHR	A	61	-15.059	22.579	-29.285	0.50	13.23	AA	O
ATOM	473	CG2	ATHR	A	61	-13.988	24.297	-29.772	0.50	10.46	AA	C
ATOM	474	CG2	BTHR	A	61	-13.654	21.642	-30.940	0.50	8.90	AA	C
ATOM	475	C	THR	A	61	-12.546	23.283	-27.571	1.00	11.38	AA	C
ATOM	476	O	THR	A	61	-11.609	24.079	-27.648	1.00	10.57	AA	O
ATOM	477	N	ASP	A	62	-13.398	23.266	-26.548	1.00	9.41	AA	N
ATOM	478	CA	ASP	A	62	-13.218	24.198	-25.449	1.00	9.36	AA	C
ATOM	479	CB	ASP	A	62	-14.444	24.306	-24.509	1.00	8.64	AA	C
ATOM	480	CG	ASP	A	62	-15.635	24.888	-25.165	1.00	11.73	AA	C
ATOM	481	OD1	ASP	A	62	-15.580	25.625	-26.185	1.00	12.79	AA	O
ATOM	482	OD2	ASP	A	62	-16.728	24.647	-24.613	1.00	11.41	AA	O
ATOM	483	C	ASP	A	62	-11.934	23.881	-24.657	1.00	9.28	AA	C
ATOM	484	O	ASP	A	62	-11.236	24.797	-24.172	1.00	10.11	AA	O
ATOM	485	N	GLY	A	63	-11.615	22.589	-24.578	1.00	10.23	AA	N
ATOM	486	CA	GLY	A	63	-10.413	22.129	-23.897	1.00	9.84	AA	C
ATOM	487	C	GLY	A	63	-9.181	22.714	-24.597	1.00	9.50	AA	C
ATOM	488	O	GLY	A	63	-8.195	23.246	-23.973	1.00	10.30	AA	O
ATOM	489	N	MET	A	64	-9.159	22.630	-25.935	1.00	10.84	AA	N
ATOM	490	CA	MET	A	64	-8.004	23.169	-26.737	1.00	12.57	AA	C
ATOM	491	CB	MET	A	64	-8.197	22.973	-28.235	1.00	13.39	AA	C
ATOM	492	CG	MET	A	64	-7.097	23.730	-28.918	1.00	14.27	AA	C

ATOM	493	SD	MET	A	64	-6.780	23.121	-30.517	1.00	21.17	AA	S
ATOM	494	CE	MET	A	64	-8.146	23.139	-31.305	1.00	13.17	AA	C
ATOM	495	C	MET	A	64	-7.843	24.642	-26.436	1.00	11.92	AA	C
ATOM	496	O	MET	A	64	-6.727	25.095	-26.218	1.00	12.86	AA	O
ATOM	497	N	ALA	A	65	-8.932	25.376	-26.440	1.00	9.79	AA	N
ATOM	498	CA	ALA	A	65	-8.880	26.821	-26.265	1.00	11.59	AA	C
ATOM	499	CB	ALA	A	65	-10.250	27.477	-26.539	1.00	11.76	AA	C
ATOM	500	C	ALA	A	65	-8.431	27.196	-24.858	1.00	11.10	AA	C
ATOM	501	O	ALA	A	65	-7.803	28.221	-24.692	1.00	13.09	AA	O
ATOM	502	N	SER	A	66	-8.725	26.389	-23.859	1.00	10.38	AA	N
ATOM	503	CA	SER	A	66	-8.238	26.646	-22.476	1.00	11.41	AA	C
ATOM	504	CB	SER	A	66	-8.976	25.762	-21.506	1.00	10.82	AA	C
ATOM	505	OG	SER	A	66	-10.325	26.256	-21.456	1.00	17.70	AA	O
ATOM	506	C	SER	A	66	-6.738	26.468	-22.323	1.00	10.55	AA	C
ATOM	507	O	SER	A	66	-6.115	27.127	-21.476	1.00	11.87	AA	O
ATOM	508	N	GLY	A	67	-6.151	25.570	-23.108	1.00	9.35	AA	N
ATOM	509	CA	GLY	A	67	-4.695	25.426	-23.103	1.00	10.07	AA	C
ATOM	510	C	GLY	A	67	-4.110	24.581	-21.998	1.00	8.65	AA	C
ATOM	511	O	GLY	A	67	-4.804	24.042	-21.125	1.00	10.39	AA	O
ATOM	512	N	LEU	A	68	-2.804	24.408	-22.079	1.00	8.85	AA	N
ATOM	513	CA	LEU	A	68	-2.064	23.628	-21.067	1.00	11.15	AA	C
ATOM	514	CB	LEU	A	68	-0.585	23.451	-21.558	1.00	10.95	AA	C
ATOM	515	CG	LEU	A	68	0.337	22.630	-20.627	1.00	9.98	AA	C
ATOM	516	CD1	LEU	A	68	-0.218	21.189	-20.456	1.00	13.46	AA	C
ATOM	517	CD2	LEU	A	68	1.781	22.632	-21.214	1.00	13.67	AA	C
ATOM	518	C	LEU	A	68	-2.162	24.180	-19.647	1.00	10.09	AA	C
ATOM	519	O	LEU	A	68	-2.141	23.359	-18.706	1.00	13.24	AA	O
ATOM	520	N	ASP	A	69	-2.378	25.479	-19.499	1.00	10.75	AA	N
ATOM	521	CA	ASP	A	69	-2.567	26.053	-18.170	1.00	11.98	AA	C
ATOM	522	CB	ASP	A	69	-2.908	27.531	-18.320	1.00	12.29	AA	C
ATOM	523	CG	ASP	A	69	-3.018	28.280	-17.009	1.00	15.20	AA	C
ATOM	524	OD1	ASP	A	69	-2.044	28.423	-16.248	1.00	13.08	AA	O
ATOM	525	OD2	ASP	A	69	-4.058	28.879	-16.679	1.00	15.04	AA	O
ATOM	526	C	ASP	A	69	-3.705	25.359	-17.446	1.00	12.74	AA	C
ATOM	527	O	ASP	A	69	-3.706	25.266	-16.230	1.00	13.92	AA	O
ATOM	528	N	LYS	A	70	-4.688	24.820	-18.189	1.00	11.72	AA	N
ATOM	529	CA	LYS	A	70	-5.913	24.141	-17.616	1.00	10.32	AA	C
ATOM	530	CB	LYS	A	70	-7.120	24.812	-18.235	1.00	12.24	AA	C
ATOM	531	CG	LYS	A	70	-7.152	26.318	-18.052	1.00	13.26	AA	C
ATOM	532	CD	LYS	A	70	-7.029	26.731	-16.600	1.00	14.94	AA	C
ATOM	533	CE	LYS	A	70	-7.441	28.242	-16.467	1.00	16.14	AA	C
ATOM	534	NZ	LYS	A	70	-6.574	29.256	-17.288	1.00	12.36	AA	N
ATOM	535	C	LYS	A	70	-5.884	22.648	-17.917	1.00	12.11	AA	C
ATOM	536	O	LYS	A	70	-6.965	21.976	-17.833	1.00	12.27	AA	O
ATOM	537	N	ASP	A	71	-4.760	22.038	-18.313	1.00	11.91	AA	N
ATOM	538	CA	ASP	A	71	-4.718	20.645	-18.735	1.00	12.13	AA	C
ATOM	539	CB	ASP	A	71	-4.981	19.651	-17.540	1.00	14.93	AA	C
ATOM	540	CG	ASP	A	71	-3.704	19.204	-16.765	1.00	15.10	AA	C
ATOM	541	OD1	ASP	A	71	-2.663	19.715	-17.080	1.00	16.74	AA	O
ATOM	542	OD2	ASP	A	71	-3.965	18.328	-15.888	1.00	19.09	AA	O
ATOM	543	C	ASP	A	71	-5.705	20.361	-19.858	1.00	11.37	AA	C
ATOM	544	O	ASP	A	71	-6.224	19.242	-20.006	1.00	11.45	AA	O
ATOM	545	N	TYR	A	72	-5.922	21.350	-20.700	1.00	9.40	AA	N
ATOM	546	CA	TYR	A	72	-6.778	21.166	-21.882	1.00	8.92	AA	C
ATOM	547	CB	TYR	A	72	-6.141	20.224	-22.925	1.00	9.70	AA	C

ATOM	548	CG	TYR	A	72	-4.883	20.871	-23.507	1.00	8.62	AA	C
ATOM	549	CD1	TYR	A	72	-4.930	21.950	-24.361	1.00	8.59	AA	C
ATOM	550	CE1	TYR	A	72	-3.800	22.547	-24.897	1.00	9.77	AA	C
ATOM	551	CZ	TYR	A	72	-2.570	21.930	-24.682	1.00	11.55	AA	C
ATOM	552	OH	TYR	A	72	-1.412	22.463	-25.303	1.00	11.11	AA	O
ATOM	553	CE2	TYR	A	72	-2.453	20.840	-23.873	1.00	11.97	AA	C
ATOM	554	CD2	TYR	A	72	-3.572	20.274	-23.270	1.00	10.15	AA	C
ATOM	555	C	TYR	A	72	-8.180	20.751	-21.540	1.00	9.05	AA	C
ATOM	556	O	TYR	A	72	-8.855	20.042	-22.292	1.00	9.09	AA	O
ATOM	557	N	LEU	A	73	-8.711	21.275	-20.418	1.00	8.36	AA	N
ATOM	558	CA	LEU	A	73	-10.130	21.179	-20.103	1.00	9.84	AA	C
ATOM	559	CB	LEU	A	73	-10.382	20.204	-18.900	1.00	10.55	AA	C
ATOM	560	CG	LEU	A	73	-10.242	18.741	-19.304	1.00	13.67	AA	C
ATOM	561	CD1	LEU	A	73	-10.391	17.911	-17.997	1.00	16.02	AA	C
ATOM	562	CD2	LEU	A	73	-11.279	18.316	-20.314	1.00	12.63	AA	C
ATOM	563	C	LEU	A	73	-10.643	22.536	-19.726	1.00	9.37	AA	C
ATOM	564	O	LEU	A	73	-10.016	23.233	-18.871	1.00	11.06	AA	O
ATOM	565	N	LYS	A	74	-11.734	22.974	-20.322	1.00	9.85	AA	N
ATOM	566	CA	LYS	A	74	-12.367	24.243	-19.853	1.00	9.46	AA	C
ATOM	567	CB	LYS	A	74	-13.654	24.459	-20.666	1.00	12.26	AA	C
ATOM	568	CG	LYS	A	74	-14.304	25.837	-20.309	1.00	11.48	AA	C
ATOM	569	CD	LYS	A	74	-15.661	26.015	-20.998	1.00	17.75	AA	C
ATOM	570	CE	LYS	A	74	-16.675	25.040	-20.501	1.00	18.93	AA	C
ATOM	571	NZ	LYS	A	74	-18.161	25.576	-20.594	1.00	20.94	AA	N
ATOM	572	C	LYS	A	74	-12.681	24.161	-18.356	1.00	10.00	AA	C
ATOM	573	O	LYS	A	74	-13.236	23.172	-17.914	1.00	11.27	AA	O
ATOM	574	N	PRO	A	75	-12.311	25.208	-17.586	1.00	12.22	AA	N
ATOM	575	CA	PRO	A	75	-12.664	25.137	-16.163	1.00	14.07	AA	C
ATOM	576	CB	PRO	A	75	-12.115	26.435	-15.576	1.00	14.84	AA	C
ATOM	577	CG	PRO	A	75	-10.918	26.793	-16.546	1.00	15.79	AA	C
ATOM	578	CD	PRO	A	75	-11.457	26.317	-17.942	1.00	11.75	AA	C
ATOM	579	C	PRO	A	75	-14.188	25.137	-15.986	1.00	12.73	AA	C
ATOM	580	O	PRO	A	75	-14.954	25.686	-16.816	1.00	11.68	AA	O
ATOM	581	N	ASP	A	76	-14.582	24.452	-14.923	1.00	12.94	AA	N
ATOM	582	CA	ASP	A	76	-16.010	24.387	-14.567	1.00	16.53	AA	C
ATOM	583	CB	ASP	A	76	-16.583	25.782	-14.214	1.00	16.75	AA	C
ATOM	584	CG	ASP	A	76	-16.369	26.181	-12.799	1.00	27.06	AA	C
ATOM	585	OD1	ASP	A	76	-15.436	25.641	-12.149	1.00	31.54	AA	O
ATOM	586	OD2	ASP	A	76	-17.211	27.026	-12.356	1.00	31.94	AA	O
ATOM	587	C	ASP	A	76	-16.844	23.840	-15.711	1.00	14.23	AA	C
ATOM	588	O	ASP	A	76	-17.922	24.394	-16.087	1.00	15.77	AA	O
ATOM	589	N	ASP	A	77	-16.327	22.773	-16.333	1.00	13.08	AA	N
ATOM	590	CA	ASP	A	77	-17.059	22.162	-17.453	1.00	11.42	AA	C
ATOM	591	CB	ASP	A	77	-16.074	21.659	-18.502	1.00	12.05	AA	C
ATOM	592	CG	ASP	A	77	-16.780	21.318	-19.767	1.00	10.72	AA	C
ATOM	593	OD1	ASP	A	77	-18.018	21.125	-19.765	1.00	11.51	AA	O
ATOM	594	OD2	ASP	A	77	-16.145	21.253	-20.843	1.00	12.18	AA	O
ATOM	595	C	ASP	A	77	-17.919	21.041	-16.908	1.00	11.77	AA	C
ATOM	596	O	ASP	A	77	-17.436	19.922	-16.663	1.00	12.27	AA	O
ATOM	597	N	SER	A	78	-19.202	21.363	-16.714	1.00	11.61	AA	N
ATOM	598	CA	SER	A	78	-20.146	20.357	-16.166	1.00	13.11	AA	C
ATOM	599	CB	SER	A	78	-21.517	20.976	-15.876	1.00	14.96	AA	C
ATOM	600	OG	SER	A	78	-22.030	21.653	-17.034	1.00	19.53	AA	O
ATOM	601	C	SER	A	78	-20.352	19.146	-17.091	1.00	13.24	AA	C
ATOM	602	O	SER	A	78	-20.900	18.084	-16.654	1.00	14.89	AA	O

ATOM	603	N	ARG	A	79	-19.976	19.270	-18.363	1.00	9.94	AA	N
ATOM	604	CA	ARG	A	79	-20.141	18.148	-19.283	1.00	10.97	AA	C
ATOM	605	CB	ARG	A	79	-19.930	18.650	-20.710	1.00	10.07	AA	C
ATOM	606	CG	ARG	A	79	-20.904	19.726	-21.102	1.00	10.12	AA	C
ATOM	607	CD	ARG	A	79	-20.519	20.294	-22.463	1.00	12.52	AA	C
ATOM	608	NE	ARG	A	79	-19.216	20.980	-22.350	1.00	11.29	AA	N
ATOM	609	CZ	ARG	A	79	-18.741	21.811	-23.243	1.00	10.10	AA	C
ATOM	610	NH1	ARG	A	79	-19.402	22.016	-24.409	1.00	11.56	AA	N
ATOM	611	NH2	ARG	A	79	-17.554	22.356	-23.085	1.00	11.22	AA	N
ATOM	612	C	ARG	A	79	-19.174	17.004	-18.986	1.00	10.26	AA	C
ATOM	613	O	ARG	A	79	-19.336	15.878	-19.513	1.00	12.07	AA	O
ATOM	614	N	VAL	A	80	-18.073	17.332	-18.328	1.00	9.25	AA	N
ATOM	615	CA	VAL	A	80	-17.078	16.258	-18.013	1.00	10.37	AA	C
ATOM	616	CB	VAL	A	80	-15.679	16.892	-17.670	1.00	10.11	AA	C
ATOM	617	CG1	VAL	A	80	-14.712	15.800	-17.261	1.00	11.06	AA	C
ATOM	618	CG2	VAL	A	80	-15.116	17.672	-18.934	1.00	12.14	AA	C
ATOM	619	C	VAL	A	80	-17.586	15.395	-16.825	1.00	12.27	AA	C
ATOM	620	O	VAL	A	80	-17.744	15.883	-15.706	1.00	11.18	AA	O
ATOM	621	N	ILE	A	81	-17.860	14.135	-17.112	1.00	9.83	AA	N
ATOM	622	CA	ILE	A	81	-18.385	13.243	-16.095	1.00	10.15	AA	C
ATOM	623	CB	ILE	A	81	-19.019	12.064	-16.748	1.00	11.04	AA	C
ATOM	624	CG1	ILE	A	81	-20.219	12.530	-17.560	1.00	10.97	AA	C
ATOM	625	CD1	ILE	A	81	-20.813	11.372	-18.475	1.00	15.01	AA	C
ATOM	626	CG2	ILE	A	81	-19.448	11.020	-15.646	1.00	10.19	AA	C
ATOM	627	C	ILE	A	81	-17.242	12.795	-15.158	1.00	10.36	AA	C
ATOM	628	O	ILE	A	81	-17.402	12.726	-13.952	1.00	11.88	AA	O
ATOM	629	N	ALA	A	82	-16.057	12.496	-15.717	1.00	8.86	AA	N
ATOM	630	CA	ALA	A	82	-14.879	12.145	-14.931	1.00	9.74	AA	C
ATOM	631	CB	ALA	A	82	-14.946	10.652	-14.596	1.00	11.00	AA	C
ATOM	632	C	ALA	A	82	-13.685	12.391	-15.825	1.00	9.79	AA	C
ATOM	633	O	ALA	A	82	-13.765	12.356	-17.077	1.00	9.61	AA	O
ATOM	634	N	HIS	A	83	-12.501	12.544	-15.237	1.00	9.46	AA	N
ATOM	635	CA	HIS	A	83	-11.287	12.730	-16.034	1.00	8.72	AA	C
ATOM	636	CB	HIS	A	83	-11.188	14.159	-16.595	1.00	9.83	AA	C
ATOM	637	CG	HIS	A	83	-11.102	15.202	-15.538	1.00	9.67	AA	C
ATOM	638	ND1	HIS	A	83	-9.904	15.746	-15.107	1.00	11.00	AA	N
ATOM	639	CE1	HIS	A	83	-10.176	16.585	-14.107	1.00	11.89	AA	C
ATOM	640	NE2	HIS	A	83	-11.481	16.630	-13.894	1.00	12.53	AA	N
ATOM	641	CD2	HIS	A	83	-12.071	15.768	-14.798	1.00	11.77	AA	C
ATOM	642	C	HIS	A	83	-10.076	12.454	-15.154	1.00	9.24	AA	C
ATOM	643	O	HIS	A	83	-10.107	12.706	-13.915	1.00	10.06	AA	O
ATOM	644	N	THR	A	84	-8.971	12.064	-15.778	1.00	9.98	AA	N
ATOM	645	CA	THR	A	84	-7.661	12.066	-15.130	1.00	10.31	AA	C
ATOM	646	CB	THR	A	84	-6.693	11.008	-15.830	1.00	10.71	AA	C
ATOM	647	OG1	THR	A	84	-6.448	11.399	-17.172	1.00	10.05	AA	O
ATOM	648	CG2	THR	A	84	-7.268	9.611	-15.843	1.00	11.67	AA	C
ATOM	649	C	THR	A	84	-7.065	13.428	-15.326	1.00	10.82	AA	C
ATOM	650	O	THR	A	84	-7.581	14.277	-16.086	1.00	11.39	AA	O
ATOM	651	N	LYS	A	85	-5.903	13.639	-14.718	1.00	10.24	AA	N
ATOM	652	CA	LYS	A	85	-5.045	14.777	-15.105	1.00	11.70	AA	C
ATOM	653	CB	LYS	A	85	-4.008	15.108	-14.027	1.00	13.10	AA	C
ATOM	654	CG	LYS	A	85	-2.968	14.162	-13.977	1.00	14.24	AA	C
ATOM	655	CD	LYS	A	85	-1.983	14.543	-12.771	1.00	19.10	AA	C
ATOM	656	CE	LYS	A	85	-0.758	13.621	-12.783	1.00	21.05	AA	C
ATOM	657	NZ	LYS	A	85	0.175	13.697	-11.583	1.00	23.44	AA	N

ATOM	658	C	LYS	A	85	-4.370	14.469	-16.469	1.00	9.71	AA	C
ATOM	659	O	LYS	A	85	-4.351	13.309	-16.975	1.00	9.39	AA	O
ATOM	660	N	LEU	A	86	-3.716	15.499	-17.016	1.00	10.11	AA	N
ATOM	661	CA	LEU	A	86	-2.875	15.290	-18.210	1.00	9.24	AA	C
ATOM	662	CB	LEU	A	86	-2.732	16.672	-18.944	1.00	10.67	AA	C
ATOM	663	CG	LEU	A	86	-2.127	16.588	-20.357	1.00	9.93	AA	C
ATOM	664	CD1	LEU	A	86	-3.167	16.137	-21.310	1.00	10.60	AA	C
ATOM	665	CD2	LEU	A	86	-1.519	17.991	-20.744	1.00	12.37	AA	C
ATOM	666	C	LEU	A	86	-1.520	14.774	-17.817	1.00	10.79	AA	C
ATOM	667	O	LEU	A	86	-0.903	15.260	-16.870	1.00	10.50	AA	O
ATOM	668	N	ILE	A	87	-1.083	13.711	-18.475	1.00	9.60	AA	N
ATOM	669	CA	ILE	A	87	0.186	13.069	-18.138	1.00	10.26	AA	C
ATOM	670	CB	ILE	A	87	0.019	11.665	-17.491	1.00	10.10	AA	C
ATOM	671	CG1	ILE	A	87	-0.568	10.687	-18.495	1.00	10.51	AA	C
ATOM	672	CD1	ILE	A	87	-0.626	9.214	-17.958	1.00	12.63	AA	C
ATOM	673	CG2	ILE	A	87	-0.699	11.765	-16.111	1.00	11.84	AA	C
ATOM	674	C	ILE	A	87	1.137	13.022	-19.355	1.00	9.51	AA	C
ATOM	675	O	ILE	A	87	0.685	12.948	-20.514	1.00	10.07	AA	O
ATOM	676	N	GLY	A	88	2.443	13.035	-19.051	1.00	9.46	AA	N
ATOM	677	CA	GLY	A	88	3.509	12.815	-20.042	1.00	10.03	AA	C
ATOM	678	C	GLY	A	88	4.181	11.462	-19.808	1.00	10.23	AA	C
ATOM	679	O	GLY	A	88	3.790	10.660	-18.976	1.00	11.06	AA	O
ATOM	680	N	SER	A	89	5.169	11.212	-20.651	1.00	12.14	AA	N
ATOM	681	CA	SER	A	89	5.976	9.983	-20.602	1.00	15.13	AA	C
ATOM	682	CB	SER	A	89	7.289	10.267	-21.421	1.00	18.21	AA	C
ATOM	683	OG	SER	A	89	8.365	9.506	-20.966	1.00	21.43	AA	O
ATOM	684	C	SER	A	89	6.368	9.604	-19.171	1.00	12.17	AA	C
ATOM	685	O	SER	A	89	6.853	10.397	-18.399	1.00	13.10	AA	O
ATOM	686	N	GLY	A	90	6.062	8.351	-18.859	1.00	10.68	AA	N
ATOM	687	CA	GLY	A	90	6.522	7.812	-17.576	1.00	11.31	AA	C
ATOM	688	C	GLY	A	90	5.629	8.167	-16.399	1.00	11.22	AA	C
ATOM	689	O	GLY	A	90	5.827	7.640	-15.279	1.00	15.28	AA	O
ATOM	690	N	GLU	A	91	4.580	8.976	-16.612	1.00	11.30	AA	N
ATOM	691	CA	GLU	A	91	3.649	9.322	-15.532	1.00	9.23	AA	C
ATOM	692	CB	GLU	A	91	3.204	10.788	-15.602	1.00	10.09	AA	C
ATOM	693	CG	GLU	A	91	4.375	11.730	-15.435	1.00	12.53	AA	C
ATOM	694	CD	GLU	A	91	4.059	13.202	-15.575	1.00	18.23	AA	C
ATOM	695	OE1	GLU	A	91	3.229	13.581	-16.351	1.00	16.69	AA	O
ATOM	696	OE2	GLU	A	91	4.708	14.078	-14.910	1.00	22.86	AA	O
ATOM	697	C	GLU	A	91	2.443	8.400	-15.579	1.00	9.43	AA	C
ATOM	698	O	GLU	A	91	2.190	7.671	-16.545	1.00	10.71	AA	O
ATOM	699	N	LYS	A	92	1.725	8.409	-14.451	1.00	10.62	AA	N
ATOM	700	CA	LYS	A	92	0.434	7.642	-14.315	1.00	10.41	AA	C
ATOM	701	CB	LYS	A	92	0.708	6.414	-13.448	1.00	12.09	AA	C
ATOM	702	CG	LYS	A	92	1.640	5.456	-14.034	1.00	14.55	AA	C
ATOM	703	CD	LYS	A	92	1.748	4.289	-13.086	1.00	14.34	AA	C
ATOM	704	CE	LYS	A	92	2.853	3.286	-13.509	1.00	23.10	AA	C
ATOM	705	NZ	LYS	A	92	3.802	3.522	-14.660	1.00	25.05	AA	N
ATOM	706	C	LYS	A	92	-0.546	8.513	-13.577	1.00	10.67	AA	C
ATOM	707	O	LYS	A	92	-0.168	9.410	-12.812	1.00	12.25	AA	O
ATOM	708	N	ASP	A	93	-1.833	8.201	-13.763	1.00	10.12	AA	N
ATOM	709	CA	ASP	A	93	-2.867	8.818	-12.926	1.00	9.64	AA	C
ATOM	710	CB	ASP	A	93	-3.256	10.199	-13.486	1.00	10.24	AA	C
ATOM	711	CG	ASP	A	93	-4.056	11.031	-12.476	1.00	12.08	AA	C
ATOM	712	OD1	ASP	A	93	-3.672	11.077	-11.261	1.00	13.44	AA	O

ATOM	713	OD2	ASP	A	93	-5.056	11.616	-12.884	1.00	12.07	AA	O
ATOM	714	C	ASP	A	93	-4.090	7.935	-12.960	1.00	8.75	AA	C
ATOM	715	O	ASP	A	93	-4.414	7.348	-14.005	1.00	10.48	AA	O
ATOM	716	N	SER	A	94	-4.765	7.835	-11.840	1.00	9.30	AA	N
ATOM	717	CA	SER	A	94	-6.013	7.041	-11.745	1.00	8.68	AA	C
ATOM	718	CB	SER	A	94	-5.940	6.074	-10.589	1.00	8.91	AA	C
ATOM	719	OG	SER	A	94	-4.952	5.105	-10.868	1.00	10.20	AA	O
ATOM	720	C	SER	A	94	-7.155	8.007	-11.441	1.00	10.19	AA	C
ATOM	721	O	SER	A	94	-7.032	8.981	-10.680	1.00	12.13	AA	O
ATOM	722	N	VAL	A	95	-8.314	7.678	-11.974	1.00	9.34	AA	N
ATOM	723	CA	VAL	A	95	-9.559	8.376	-11.599	1.00	9.86	AA	C
ATOM	724	CB	VAL	A	95	-10.103	9.337	-12.764	1.00	10.08	AA	C
ATOM	725	CG1	VAL	A	95	-10.370	8.633	-14.050	1.00	12.87	AA	C
ATOM	726	CG2	VAL	A	95	-11.370	10.059	-12.281	1.00	12.58	AA	C
ATOM	727	C	VAL	A	95	-10.617	7.344	-11.199	1.00	9.00	AA	C
ATOM	728	O	VAL	A	95	-10.807	6.381	-11.924	1.00	10.38	AA	O
ATOM	729	N	THR	A	96	-11.315	7.614	-10.094	1.00	8.85	AA	N
ATOM	730	CA	THR	A	96	-12.405	6.786	-9.642	1.00	9.18	AA	C
ATOM	731	CB	THR	A	96	-12.163	6.291	-8.249	1.00	10.61	AA	C
ATOM	732	OG1	THR	A	96	-11.137	5.307	-8.370	1.00	9.34	AA	O
ATOM	733	CG2	THR	A	96	-13.410	5.598	-7.640	1.00	10.96	AA	C
ATOM	734	C	THR	A	96	-13.680	7.669	-9.691	1.00	10.07	AA	C
ATOM	735	O	THR	A	96	-13.674	8.859	-9.274	1.00	10.79	AA	O
ATOM	736	N	PHE	A	97	-14.760	7.095	-10.207	1.00	8.97	AA	N
ATOM	737	CA	PHE	A	97	-16.050	7.809	-10.214	1.00	10.08	AA	C
ATOM	738	CB	PHE	A	97	-16.305	8.520	-11.562	1.00	11.37	AA	C
ATOM	739	CG	PHE	A	97	-16.385	7.647	-12.710	1.00	10.20	AA	C
ATOM	740	CD1	PHE	A	97	-15.213	7.267	-13.473	1.00	11.31	AA	C
ATOM	741	CE1	PHE	A	97	-15.274	6.426	-14.593	1.00	11.76	AA	C
ATOM	742	CZ	PHE	A	97	-16.487	5.971	-14.999	1.00	12.68	AA	C
ATOM	743	CE2	PHE	A	97	-17.710	6.323	-14.219	1.00	12.99	AA	C
ATOM	744	CD2	PHE	A	97	-17.648	7.169	-13.139	1.00	11.53	AA	C
ATOM	745	C	PHE	A	97	-17.186	6.908	-9.828	1.00	9.84	AA	C
ATOM	746	O	PHE	A	97	-17.098	5.701	-9.984	1.00	10.65	AA	O
ATOM	747	N	ASP	A	98	-18.272	7.525	-9.385	1.00	9.32	AA	N
ATOM	748	CA	ASP	A	98	-19.489	6.786	-9.019	1.00	10.28	AA	C
ATOM	749	CB	ASP	A	98	-20.430	7.698	-8.226	1.00	12.16	AA	C
ATOM	750	CG	ASP	A	98	-19.912	8.093	-6.893	1.00	16.37	AA	C
ATOM	751	OD1	ASP	A	98	-19.120	7.399	-6.229	1.00	16.78	AA	O
ATOM	752	OD2	ASP	A	98	-20.478	9.090	-6.375	1.00	20.90	AA	O
ATOM	753	C	ASP	A	98	-20.227	6.301	-10.257	1.00	10.03	AA	C
ATOM	754	O	ASP	A	98	-20.510	7.118	-11.169	1.00	9.52	AA	O
ATOM	755	N	VAL	A	99	-20.568	4.997	-10.264	1.00	9.78	AA	N
ATOM	756	CA	VAL	A	99	-21.265	4.428	-11.374	1.00	11.32	AA	C
ATOM	757	CB	VAL	A	99	-21.229	2.901	-11.241	1.00	10.09	AA	C
ATOM	758	CG1	VAL	A	99	-22.130	2.192	-12.312	1.00	15.20	AA	C
ATOM	759	CG2	VAL	A	99	-19.785	2.402	-11.480	1.00	12.93	AA	C
ATOM	760	C	VAL	A	99	-22.690	4.994	-11.473	1.00	11.27	AA	C
ATOM	761	O	VAL	A	99	-23.268	5.006	-12.571	1.00	10.66	AA	O
ATOM	762	N	SER	A	100	-23.202	5.575	-10.355	1.00	9.97	AA	N
ATOM	763	CA	SER	A	100	-24.518	6.289	-10.413	1.00	10.77	AA	C
ATOM	764	CB	SER	A	100	-24.889	6.723	-9.005	1.00	12.56	AA	C
ATOM	765	OG	SER	A	100	-23.933	7.608	-8.459	1.00	14.49	AA	O
ATOM	766	C	SER	A	100	-24.451	7.470	-11.377	1.00	10.51	AA	C
ATOM	767	O	SER	A	100	-25.523	8.024	-11.755	1.00	10.96	AA	O

ATOM	768	N	LYS	A	101	-23.251	7.929	-11.759	1.00	10.79	AA	N
ATOM	769	CA	LYS	A	101	-23.201	8.981	-12.756	1.00	11.81	AA	C
ATOM	770	CB	LYS	A	101	-21.843	9.690	-12.790	1.00	14.35	AA	C
ATOM	771	CG	LYS	A	101	-21.615	10.538	-11.509	1.00	14.32	AA	C
ATOM	772	CD	LYS	A	101	-20.293	11.270	-11.527	1.00	21.28	AA	C
ATOM	773	CE	LYS	A	101	-20.282	12.363	-10.396	1.00	25.41	AA	C
ATOM	774	NZ	LYS	A	101	-20.548	11.709	-9.053	1.00	35.15	AA	N
ATOM	775	C	LYS	A	101	-23.638	8.554	-14.144	1.00	13.02	AA	C
ATOM	776	O	LYS	A	101	-23.810	9.465	-15.007	1.00	15.22	AA	O
ATOM	777	N	LEU	A	102	-23.703	7.245	-14.425	1.00	10.62	AA	N
ATOM	778	CA	LEU	A	102	-23.969	6.719	-15.745	1.00	8.80	AA	C
ATOM	779	CB	LEU	A	102	-23.040	5.555	-16.030	1.00	9.64	AA	C
ATOM	780	CG	LEU	A	102	-21.545	5.944	-15.946	1.00	10.40	AA	C
ATOM	781	CD1	LEU	A	102	-20.667	4.684	-16.242	1.00	13.20	AA	C
ATOM	782	CD2	LEU	A	102	-21.200	7.154	-16.865	1.00	13.14	AA	C
ATOM	783	C	LEU	A	102	-25.396	6.156	-15.741	1.00	11.28	AA	C
ATOM	784	O	LEU	A	102	-25.917	5.689	-14.699	1.00	12.32	AA	O
ATOM	785	N	LYS	A	103	-25.993	6.054	-16.945	1.00	10.61	AA	N
ATOM	786	CA	LYS	A	103	-27.312	5.428	-17.123	1.00	12.16	AA	C
ATOM	787	CB	LYS	A	103	-28.249	6.585	-17.590	1.00	13.79	AA	C
ATOM	788	CG	LYS	A	103	-29.744	6.393	-17.788	1.00	21.48	AA	C
ATOM	789	CD	LYS	A	103	-30.265	7.851	-18.133	1.00	26.31	AA	C
ATOM	790	CE	LYS	A	103	-31.753	7.944	-18.493	1.00	31.11	AA	C
ATOM	791	NZ	LYS	A	103	-32.687	7.642	-17.336	1.00	33.89	AA	N
ATOM	792	C	LYS	A	103	-27.185	4.364	-18.207	1.00	13.04	AA	C
ATOM	793	O	LYS	A	103	-26.477	4.566	-19.202	1.00	13.19	AA	O
ATOM	794	N	GLU	A	104	-27.907	3.234	-18.069	1.00	12.62	AA	N
ATOM	795	CA	GLU	A	104	-28.010	2.299	-19.211	1.00	15.57	AA	C
ATOM	796	CB	GLU	A	104	-28.888	1.120	-18.878	1.00	11.89	AA	C
ATOM	797	CG	GLU	A	104	-28.166	0.220	-17.950	1.00	16.05	AA	C
ATOM	798	CD	GLU	A	104	-28.816	-1.138	-17.879	1.00	18.04	AA	C
ATOM	799	OE1	GLU	A	104	-28.767	-1.922	-18.896	1.00	22.62	AA	O
ATOM	800	OE2	GLU	A	104	-29.336	-1.421	-16.803	1.00	18.45	AA	O
ATOM	801	C	GLU	A	104	-28.690	2.969	-20.380	1.00	16.29	AA	C
ATOM	802	O	GLU	A	104	-29.542	3.825	-20.174	1.00	17.17	AA	O
ATOM	803	N	GLY	A	105	-28.284	2.641	-21.597	1.00	18.05	AA	N
ATOM	804	CA	GLY	A	105	-28.944	3.187	-22.775	1.00	21.63	AA	C
ATOM	805	C	GLY	A	105	-28.375	4.526	-23.245	1.00	23.80	AA	C
ATOM	806	O	GLY	A	105	-28.546	4.868	-24.425	1.00	25.04	AA	O
ATOM	807	N	GLU	A	106	-27.701	5.271	-22.345	1.00	21.93	AA	N
ATOM	808	CA	GLU	A	106	-26.984	6.495	-22.730	1.00	21.33	AA	C
ATOM	809	CB	GLU	A	106	-26.809	7.471	-21.546	1.00	21.42	AA	C
ATOM	810	CG	GLU	A	106	-26.024	8.818	-21.848	1.00	21.58	AA	C
ATOM	811	CD	GLU	A	106	-26.669	9.776	-22.861	1.00	26.42	AA	C
ATOM	812	OE1	GLU	A	106	-27.367	10.723	-22.395	1.00	27.89	AA	O
ATOM	813	OE2	GLU	A	106	-26.488	9.596	-24.116	1.00	24.30	AA	O
ATOM	814	C	GLU	A	106	-25.705	6.085	-23.419	1.00	20.39	AA	C
ATOM	815	O	GLU	A	106	-25.105	4.991	-23.166	1.00	21.12	AA	O
ATOM	816	N	GLN	A	107	-25.277	6.970	-24.312	1.00	16.79	AA	N
ATOM	817	CA	GLN	A	107	-24.014	6.702	-25.035	1.00	16.63	AA	C
ATOM	818	CB	GLN	A	107	-24.144	6.792	-26.546	1.00	17.36	AA	C
ATOM	819	CG	GLN	A	107	-25.252	5.840	-27.157	1.00	22.40	AA	C
ATOM	820	CD	GLN	A	107	-25.582	6.249	-28.606	1.00	35.06	AA	C
ATOM	821	OE1	GLN	A	107	-25.672	7.456	-28.934	1.00	38.85	AA	O
ATOM	822	NE2	GLN	A	107	-25.758	5.254	-29.473	1.00	34.17	AA	N

ATOM	823	C	GLN	A 107	-22.924	7.648	-24.582	1.00	14.12	AA	C
ATOM	824	O	GLN	A 107	-23.202	8.887	-24.563	1.00	14.95	AA	O
ATOM	825	N	TYR	A 108	-21.813	7.043	-24.083	1.00	14.27	AA	N
ATOM	826	CA	TYR	A 108	-20.719	7.835	-23.628	1.00	9.78	AA	C
ATOM	827	CB	TYR	A 108	-20.345	7.451	-22.171	1.00	10.45	AA	C
ATOM	828	CG	TYR	A 108	-21.505	7.555	-21.221	1.00	9.47	AA	C
ATOM	829	CD1	TYR	A 108	-21.887	8.787	-20.761	1.00	9.42	AA	C
ATOM	830	CE1	TYR	A 108	-23.016	8.978	-19.942	1.00	10.93	AA	C
ATOM	831	CZ	TYR	A 108	-23.761	7.849	-19.619	1.00	9.33	AA	C
ATOM	832	OH	TYR	A 108	-24.889	7.995	-18.791	1.00	9.56	AA	O
ATOM	833	CE2	TYR	A 108	-23.397	6.603	-20.097	1.00	9.21	AA	C
ATOM	834	CD2	TYR	A 108	-22.294	6.446	-20.855	1.00	10.00	AA	C
ATOM	835	C	TYR	A 108	-19.489	7.598	-24.521	1.00	9.07	AA	C
ATOM	836	O	TYR	A 108	-19.304	6.571	-25.183	1.00	10.05	AA	O
ATOM	837	N	MET	A 109	-18.624	8.621	-24.512	1.00	8.19	AA	N
ATOM	838	CA	MET	A 109	-17.294	8.540	-25.195	1.00	8.63	AA	C
ATOM	839	CB	MET	A 109	-17.225	9.562	-26.334	1.00	10.30	AA	C
ATOM	840	CG	MET	A 109	-18.235	9.324	-27.508	1.00	11.08	AA	C
ATOM	841	SD	MET	A 109	-17.922	7.800	-28.427	1.00	11.83	AA	S
ATOM	842	CE	MET	A 109	-16.415	8.247	-29.207	1.00	13.65	AA	C
ATOM	843	C	MET	A 109	-16.254	8.884	-24.221	1.00	7.60	AA	C
ATOM	844	O	MET	A 109	-16.449	9.777	-23.380	1.00	9.86	AA	O
ATOM	845	N	PHE	A 110	-15.098	8.185	-24.314	1.00	7.73	AA	N
ATOM	846	CA	PHE	A 110	-13.918	8.634	-23.625	1.00	9.46	AA	C
ATOM	847	CB	PHE	A 110	-13.312	7.474	-22.746	1.00	8.44	AA	C
ATOM	848	CG	PHE	A 110	-12.741	6.319	-23.501	1.00	8.03	AA	C
ATOM	849	CD1	PHE	A 110	-11.425	6.326	-23.972	1.00	8.84	AA	C
ATOM	850	CE1	PHE	A 110	-10.827	5.251	-24.669	1.00	10.04	AA	C
ATOM	851	CZ	PHE	A 110	-11.625	4.151	-24.962	1.00	9.21	AA	C
ATOM	852	CE2	PHE	A 110	-12.951	4.107	-24.548	1.00	9.93	AA	C
ATOM	853	CD2	PHE	A 110	-13.493	5.210	-23.818	1.00	8.34	AA	C
ATOM	854	C	PHE	A 110	-12.895	9.095	-24.653	1.00	7.92	AA	C
ATOM	855	O	PHE	A 110	-12.923	8.634	-25.846	1.00	8.71	AA	O
ATOM	856	N	PHE	A 111	-12.040	10.004	-24.258	1.00	7.84	AA	N
ATOM	857	CA	PHE	A 111	-11.123	10.633	-25.248	1.00	8.68	AA	C
ATOM	858	CB	PHE	A 111	-11.912	11.564	-26.204	1.00	9.45	AA	C
ATOM	859	CG	PHE	A 111	-12.893	12.470	-25.492	1.00	8.44	AA	C
ATOM	860	CD1	PHE	A 111	-14.206	12.052	-25.215	1.00	8.62	AA	C
ATOM	861	CE1	PHE	A 111	-15.066	12.843	-24.534	1.00	8.54	AA	C
ATOM	862	CZ	PHE	A 111	-14.680	14.080	-24.057	1.00	8.13	AA	C
ATOM	863	CE2	PHE	A 111	-13.447	14.550	-24.305	1.00	8.74	AA	C
ATOM	864	CD2	PHE	A 111	-12.528	13.725	-25.024	1.00	8.18	AA	C
ATOM	865	C	PHE	A 111	-10.045	11.403	-24.515	1.00	8.32	AA	C
ATOM	866	O	PHE	A 111	-10.146	11.706	-23.321	1.00	8.58	AA	O
ATOM	867	N	CYS	A 112	-8.999	11.722	-25.247	1.00	8.37	AA	N
ATOM	868	CA	CYS	A 112	-7.935	12.667	-24.850	1.00	9.06	AA	C
ATOM	869	CB	CYS	A 112	-6.571	12.148	-25.377	1.00	9.23	AA	C
ATOM	870	SG	CYS	A 112	-5.221	13.275	-24.941	1.00	9.32	AA	S
ATOM	871	C	CYS	A 112	-8.252	14.059	-25.432	1.00	7.72	AA	C
ATOM	872	O	CYS	A 112	-8.580	14.134	-26.622	1.00	9.36	AA	O
ATOM	873	N	THR	A 113	-8.110	15.100	-24.609	1.00	7.83	AA	N
ATOM	874	CA	THR	A 113	-8.303	16.454	-25.107	1.00	8.70	AA	C
ATOM	875	CB	THR	A 113	-9.154	17.295	-24.118	1.00	7.73	AA	C
ATOM	876	OG1	THR	A 113	-8.430	17.432	-22.909	1.00	8.57	AA	O
ATOM	877	CG2	THR	A 113	-10.445	16.652	-23.818	1.00	8.83	AA	C

ATOM	878	C	THR	A 113	-7.064	17.228	-25.451	1.00	7.20	AA	C
ATOM	879	O	THR	A 113	-7.146	18.406	-25.793	1.00	9.30	AA	O
ATOM	880	N	PHE	A 114	-5.901	16.571	-25.452	1.00	8.23	AA	N
ATOM	881	CA	PHE	A 114	-4.694	17.292	-25.916	1.00	8.51	AA	C
ATOM	882	CB	PHE	A 114	-3.443	16.385	-25.792	1.00	8.02	AA	C
ATOM	883	CG	PHE	A 114	-2.162	17.117	-26.060	1.00	9.31	AA	C
ATOM	884	CD1	PHE	A 114	-1.750	17.359	-27.371	1.00	9.64	AA	C
ATOM	885	CE1	PHE	A 114	-0.568	18.060	-27.644	1.00	11.07	AA	C
ATOM	886	CZ	PHE	A 114	0.145	18.576	-26.543	1.00	11.47	AA	C
ATOM	887	CE2	PHE	A 114	-0.263	18.363	-25.228	1.00	9.06	AA	C
ATOM	888	CD2	PHE	A 114	-1.432	17.681	-25.004	1.00	9.36	AA	C
ATOM	889	C	PHE	A 114	-4.967	17.693	-27.413	1.00	9.12	AA	C
ATOM	890	O	PHE	A 114	-5.506	16.847	-28.134	1.00	8.70	AA	O
ATOM	891	N	PRO	A 115	-4.549	18.882	-27.898	1.00	7.88	AA	N
ATOM	892	CA	PRO	A 115	-4.902	19.297	-29.262	1.00	8.50	AA	C
ATOM	893	CB	PRO	A 115	-4.096	20.568	-29.435	1.00	8.46	AA	C
ATOM	894	CG	PRO	A 115	-3.970	21.141	-27.991	1.00	10.13	AA	C
ATOM	895	CD	PRO	A 115	-3.938	19.954	-27.083	1.00	8.41	AA	C
ATOM	896	C	PRO	A 115	-4.555	18.222	-30.290	1.00	7.60	AA	C
ATOM	897	O	PRO	A 115	-3.426	17.737	-30.353	1.00	8.02	AA	O
ATOM	898	N	GLY	A 116	-5.554	17.923	-31.126	1.00	7.94	AA	N
ATOM	899	CA	GLY	A 116	-5.373	16.935	-32.221	1.00	8.82	AA	C
ATOM	900	C	GLY	A 116	-5.604	15.509	-31.856	1.00	7.47	AA	C
ATOM	901	O	GLY	A 116	-5.870	14.716	-32.725	1.00	9.98	AA	O
ATOM	902	N	HIS	A 117	-5.484	15.167	-30.605	1.00	7.99	AA	N
ATOM	903	CA	HIS	A 117	-5.432	13.755	-30.241	1.00	8.60	AA	C
ATOM	904	CB	HIS	A 117	-4.874	13.517	-28.817	1.00	9.26	AA	C
ATOM	905	CG	HIS	A 117	-3.435	13.902	-28.685	1.00	8.77	AA	C
ATOM	906	ND1	HIS	A 117	-2.707	13.503	-27.592	1.00	7.21	AA	N
ATOM	907	CE1	HIS	A 117	-1.450	13.898	-27.800	1.00	7.59	AA	C
ATOM	908	NE2	HIS	A 117	-1.314	14.496	-28.976	1.00	8.66	AA	N
ATOM	909	CD2	HIS	A 117	-2.577	14.503	-29.549	1.00	8.80	AA	C
ATOM	910	C	HIS	A 117	-6.712	13.007	-30.379	1.00	8.21	AA	C
ATOM	911	O	HIS	A 117	-6.680	11.791	-30.706	1.00	10.15	AA	O
ATOM	912	N	SER	A 118	-7.840	13.669	-30.193	1.00	8.17	AA	N
ATOM	913	CA	SER	A 118	-9.125	12.922	-30.220	1.00	8.77	AA	C
ATOM	914	CB	SER	A 118	-10.273	13.851	-29.780	1.00	10.59	AA	C
ATOM	915	OG	SER	A 118	-10.546	14.871	-30.704	1.00	12.27	AA	O
ATOM	916	C	SER	A 118	-9.415	12.409	-31.648	1.00	10.16	AA	C
ATOM	917	O	SER	A 118	-10.354	11.599	-31.832	1.00	11.22	AA	O
ATOM	918	N	ALA	A 119	-8.714	12.825	-32.698	1.00	8.35	AA	N
ATOM	919	CA	ALA	A 119	-8.863	12.207	-34.013	1.00	10.31	AA	C
ATOM	920	CB	ALA	A 119	-7.821	12.792	-35.071	1.00	11.07	AA	C
ATOM	921	C	ALA	A 119	-8.726	10.726	-33.933	1.00	10.44	AA	C
ATOM	922	O	ALA	A 119	-9.389	10.006	-34.710	1.00	11.79	AA	O
ATOM	923	N	LEU	A 120	-7.825	10.234	-33.073	1.00	8.71	AA	N
ATOM	924	CA	LEU	A 120	-7.605	8.808	-32.847	1.00	9.94	AA	C
ATOM	925	CB	LEU	A 120	-6.187	8.318	-33.285	1.00	13.06	AA	C
ATOM	926	CG	LEU	A 120	-6.018	8.297	-34.820	1.00	17.49	AA	C
ATOM	927	CD1	LEU	A 120	-4.616	7.907	-35.150	1.00	19.16	AA	C
ATOM	928	CD2	LEU	A 120	-7.021	7.328	-35.502	1.00	16.93	AA	C
ATOM	929	C	LEU	A 120	-7.857	8.356	-31.453	1.00	9.27	AA	C
ATOM	930	O	LEU	A 120	-8.280	7.242	-31.278	1.00	9.45	AA	O
ATOM	931	N	MET	A 121	-7.644	9.213	-30.490	1.00	8.26	AA	N
ATOM	932	CA	MET	A 121	-7.654	8.783	-29.055	1.00	8.56	AA	C

ATOM	933	CB	MET	A	121	-6.637	9.598	-28.286	1.00	8.37	AA	C
ATOM	934	CG	MET	A	121	-5.228	9.261	-28.776	1.00	10.95	AA	C
ATOM	935	SD	MET	A	121	-3.921	9.991	-27.741	1.00	9.56	AA	S
ATOM	936	CE	MET	A	121	-3.871	8.772	-26.423	1.00	9.54	AA	C
ATOM	937	C	MET	A	121	-9.071	8.970	-28.524	1.00	9.21	AA	C
ATOM	938	O	MET	A	121	-9.357	9.957	-27.840	1.00	9.45	AA	O
ATOM	939	N	LYS	A	122	-9.956	8.042	-28.883	1.00	8.44	AA	N
ATOM	940	CA	ALYS	A	122	-11.331	8.090	-28.394	0.50	8.33	AA	C
ATOM	941	CA	BLYS	A	122	-11.355	8.091	-28.422	0.50	8.40	AA	C
ATOM	942	CB	ALYS	A	122	-12.168	9.089	-29.233	0.50	9.70	AA	C
ATOM	943	CB	BLYS	A	122	-12.240	9.025	-29.304	0.50	9.91	AA	C
ATOM	944	CG	ALYS	A	122	-12.593	8.575	-30.618	0.50	9.83	AA	C
ATOM	945	CG	BLYS	A	122	-12.387	8.598	-30.774	0.50	10.30	AA	C
ATOM	946	CD	ALYS	A	122	-11.459	8.468	-31.645	0.50	8.54	AA	C
ATOM	947	CD	BLYS	A	122	-13.204	9.608	-31.518	0.50	14.15	AA	C
ATOM	948	CE	ALYS	A	122	-12.073	8.228	-33.052	0.50	13.27	AA	C
ATOM	949	CE	BLYS	A	122	-12.707	9.853	-32.977	0.50	15.12	AA	C
ATOM	950	NZ	ALYS	A	122	-10.967	7.687	-33.945	0.50	10.86	AA	N
ATOM	951	NZ	BLYS	A	122	-12.790	11.380	-33.152	0.50	19.31	AA	N
ATOM	952	C	LYS	A	122	-11.899	6.679	-28.481	1.00	8.74	AA	C
ATOM	953	O	LYS	A	122	-11.398	5.826	-29.241	1.00	9.21	AA	O
ATOM	954	N	GLY	A	123	-12.988	6.447	-27.756	1.00	8.61	AA	N
ATOM	955	CA	GLY	A	123	-13.700	5.147	-27.836	1.00	7.60	AA	C
ATOM	956	C	GLY	A	123	-14.950	5.246	-27.047	1.00	8.04	AA	C
ATOM	957	O	GLY	A	123	-15.299	6.325	-26.546	1.00	8.70	AA	O
ATOM	958	N	THR	A	124	-15.733	4.153	-27.103	1.00	8.39	AA	N
ATOM	959	CA	THR	A	124	-17.059	4.189	-26.450	1.00	8.66	AA	C
ATOM	960	CB	THR	A	124	-18.072	3.300	-27.227	1.00	7.93	AA	C
ATOM	961	OG1	THR	A	124	-17.533	1.964	-27.285	1.00	7.96	AA	O
ATOM	962	CG2	THR	A	124	-18.162	3.828	-28.625	1.00	7.31	AA	C
ATOM	963	C	THR	A	124	-16.943	3.657	-25.012	1.00	8.22	AA	C
ATOM	964	O	THR	A	124	-16.070	2.866	-24.656	1.00	8.84	AA	O
ATOM	965	N	LEU	A	125	-17.929	4.125	-24.209	1.00	9.24	AA	N
ATOM	966	CA	LEU	A	125	-18.058	3.560	-22.850	1.00	10.34	AA	C
ATOM	967	CB	LEU	A	125	-17.486	4.474	-21.751	1.00	9.09	AA	C
ATOM	968	CG	LEU	A	125	-17.573	3.863	-20.317	1.00	8.31	AA	C
ATOM	969	CD1	LEU	A	125	-16.399	4.330	-19.484	1.00	10.53	AA	C
ATOM	970	CD2	LEU	A	125	-18.926	4.393	-19.638	1.00	9.06	AA	C
ATOM	971	C	LEU	A	125	-19.575	3.311	-22.619	1.00	9.13	AA	C
ATOM	972	O	LEU	A	125	-20.399	4.142	-22.969	1.00	10.51	AA	O
ATOM	973	N	THR	A	126	-19.896	2.152	-22.101	1.00	10.04	AA	N
ATOM	974	CA	ATHR	A	126	-21.290	1.789	-21.875	0.50	10.57	AA	C
ATOM	975	CA	BTHR	A	126	-21.296	1.831	-21.855	0.50	10.65	AA	C
ATOM	976	CB	ATHR	A	126	-21.823	0.817	-22.987	0.50	11.84	AA	C
ATOM	977	CB	BTHR	A	126	-21.844	0.943	-23.012	0.50	12.54	AA	C
ATOM	978	OG1	ATHR	A	126	-21.601	-0.550	-22.636	0.50	17.39	AA	O
ATOM	979	OG1	BTHR	A	126	-23.239	0.606	-22.834	0.50	14.19	AA	O
ATOM	980	CG2	ATHR	A	126	-21.170	1.137	-24.367	0.50	10.02	AA	C
ATOM	981	CG2	BTHR	A	126	-20.980	-0.270	-23.189	0.50	13.03	AA	C
ATOM	982	C	THR	A	126	-21.459	1.138	-20.526	1.00	13.09	AA	C
ATOM	983	O	THR	A	126	-20.561	0.363	-20.110	1.00	12.75	AA	O
ATOM	984	N	LEU	A	127	-22.616	1.405	-19.886	1.00	12.72	AA	N
ATOM	985	CA	LEU	A	127	-22.949	0.754	-18.620	1.00	14.49	AA	C
ATOM	986	CB	LEU	A	127	-23.769	1.744	-17.796	1.00	15.69	AA	C
ATOM	987	CG	LEU	A	127	-24.284	1.281	-16.442	1.00	13.62	AA	C

ATOM	988	CD1 LEU A 127	-23.053	0.974	-15.606	1.00	15.17	AA	C
ATOM	989	CD2 LEU A 127	-25.209	2.306	-15.824	1.00	14.24	AA	C
ATOM	990	C LEU A 127	-23.850	-0.423	-18.973	1.00	15.12	AA	C
ATOM	991	O LEU A 127	-24.858	-0.205	-19.699	1.00	15.50	AA	O
ATOM	992	N LYS A 128	-23.434	-1.620	-18.566	1.00	14.46	AA	N
ATOM	993	CA LYS A 128	-24.309	-2.774	-18.771	1.00	17.12	AA	C
ATOM	994	CB LYS A 128	-23.879	-3.522	-20.022	1.00	18.04	AA	C
ATOM	995	CG LYS A 128	-22.755	-4.463	-19.843	1.00	23.54	AA	C
ATOM	996	CD LYS A 128	-23.104	-5.761	-20.637	1.00	28.32	AA	C
ATOM	997	CE LYS A 128	-22.045	-6.899	-20.574	1.00	29.21	AA	C
ATOM	998	NZ LYS A 128	-22.475	-8.053	-21.465	1.00	36.07	AA	N
ATOM	999	C LYS A 128	-24.364	-3.740	-17.597	1.00	17.14	AA	C
ATOM	1000	O LYS A 128	-23.490	-3.717	-16.741	1.00	18.73	AA	O
ATOM	1001	OXT LYS A 128	-25.305	-4.608	-17.550	1.00	17.50	AA	O
ATOM	1002	N ALA B 1	-47.066	24.741	-13.022	1.00	13.47	BA	N
ATOM	1003	CA ALA B 1	-45.902	24.219	-12.243	1.00	11.73	BA	C
ATOM	1004	CB ALA B 1	-44.820	23.692	-13.204	1.00	15.51	BA	C
ATOM	1005	C ALA B 1	-45.340	25.235	-11.304	1.00	12.42	BA	C
ATOM	1006	O ALA B 1	-45.667	26.430	-11.437	1.00	13.52	BA	O
ATOM	1007	N GLU B 2	-44.646	24.838	-10.286	1.00	12.12	BA	N
ATOM	1008	CA GLU B 2	-44.004	25.767	-9.353	1.00	12.95	BA	C
ATOM	1009	CB GLU B 2	-44.799	25.880	-8.046	1.00	16.05	BA	C
ATOM	1010	CG GLU B 2	-46.127	26.719	-8.097	1.00	21.36	BA	C
ATOM	1011	CD GLU B 2	-45.955	28.257	-8.399	1.00	27.46	BA	C
ATOM	1012	OE1 GLU B 2	-44.812	28.800	-8.565	1.00	26.74	BA	O
ATOM	1013	OE2 GLU B 2	-47.026	28.914	-8.476	1.00	33.29	BA	O
ATOM	1014	C GLU B 2	-42.643	25.148	-9.066	1.00	13.18	BA	C
ATOM	1015	O GLU B 2	-42.557	24.176	-8.288	1.00	15.26	BA	O
ATOM	1016	N CYS B 3	-41.621	25.628	-9.747	1.00	12.02	BA	N
ATOM	1017	CA CYS B 3	-40.290	25.084	-9.758	1.00	8.85	BA	C
ATOM	1018	CB CYS B 3	-40.034	24.468	-11.132	1.00	8.99	BA	C
ATOM	1019	SG CYS B 3	-41.080	23.027	-11.407	1.00	11.35	BA	S
ATOM	1020	C CYS B 3	-39.283	26.180	-9.448	1.00	11.50	BA	C
ATOM	1021	O CYS B 3	-38.100	25.984	-9.817	1.00	10.46	BA	O
ATOM	1022	N SER B 4	-39.666	27.200	-8.648	1.00	11.47	BA	N
ATOM	1023	CA ASER B 4	-38.651	28.151	-8.178	0.50	11.27	BA	C
ATOM	1024	CA BSER B 4	-38.776	28.332	-8.267	0.50	9.68	BA	C
ATOM	1025	CB ASER B 4	-38.618	29.276	-9.163	0.50	11.38	BA	C
ATOM	1026	CB BSER B 4	-39.094	29.593	-9.100	0.50	7.57	BA	C
ATOM	1027	OG ASER B 4	-39.820	29.946	-9.054	0.50	15.63	BA	O
ATOM	1028	OG BSER B 4	-38.941	29.385	-10.521	0.50	7.47	BA	O
ATOM	1029	C SER B 4	-38.996	28.670	-6.796	1.00	11.99	BA	C
ATOM	1030	O SER B 4	-40.089	28.373	-6.263	1.00	14.09	BA	O
ATOM	1031	N VAL B 5	-38.052	29.344	-6.159	1.00	11.12	BA	N
ATOM	1032	CA VAL B 5	-38.220	29.974	-4.859	1.00	10.66	BA	C
ATOM	1033	CB VAL B 5	-37.804	29.019	-3.777	1.00	13.58	BA	C
ATOM	1034	CG1 VAL B 5	-36.282	28.733	-3.728	1.00	14.71	BA	C
ATOM	1035	CG2 VAL B 5	-38.301	29.594	-2.468	1.00	15.81	BA	C
ATOM	1036	C VAL B 5	-37.424	31.281	-4.880	1.00	12.49	BA	C
ATOM	1037	O VAL B 5	-36.391	31.358	-5.527	1.00	13.18	BA	O
ATOM	1038	N ASP B 6	-37.948	32.319	-4.228	1.00	12.87	BA	N
ATOM	1039	CA ASP B 6	-37.256	33.585	-3.945	1.00	13.04	BA	C
ATOM	1040	CB ASP B 6	-38.189	34.744	-4.143	1.00	15.20	BA	C
ATOM	1041	CG ASP B 6	-38.758	34.731	-5.573	1.00	20.37	BA	C
ATOM	1042	OD1 ASP B 6	-38.015	34.875	-6.552	1.00	21.50	BA	O

ATOM	1043	OD2 ASP B	6	-39.956	34.447	-5.686	1.00	22.74	BA	O
ATOM	1044	C ASP B	6	-36.683	33.535	-2.560	1.00	15.23	BA	C
ATOM	1045	O ASP B	6	-37.421	33.250	-1.587	1.00	16.65	BA	O
ATOM	1046	N ILE B	7	-35.376	33.812	-2.497	1.00	12.76	BA	N
ATOM	1047	CA ILE B	7	-34.625	33.860	-1.217	1.00	13.48	BA	C
ATOM	1048	CB ILE B	7	-33.503	32.870	-1.258	1.00	13.45	BA	C
ATOM	1049	CG1 ILE B	7	-33.986	31.417	-1.402	1.00	15.65	BA	C
ATOM	1050	CD1 ILE B	7	-34.858	30.876	-0.233	1.00	18.97	BA	C
ATOM	1051	CG2 ILE B	7	-32.726	32.951	0.132	1.00	16.52	BA	C
ATOM	1052	C ILE B	7	-34.047	35.277	-1.148	1.00	12.83	BA	C
ATOM	1053	O ILE B	7	-33.432	35.790	-2.087	1.00	13.06	BA	O
ATOM	1054	N MET B	8	-34.190	35.891	0.036	1.00	13.40	BA	N
ATOM	1055	CA AMET B	8	-33.590	37.213	0.252	0.50	12.71	BA	C
ATOM	1056	CA BMET B	8	-33.677	37.264	0.308	0.50	13.31	BA	C
ATOM	1057	CB AMET B	8	-34.675	38.292	0.333	0.50	13.65	BA	C
ATOM	1058	CB BMET B	8	-34.816	38.273	0.664	0.50	14.60	BA	C
ATOM	1059	CG AMET B	8	-35.494	38.375	-0.964	0.50	15.17	BA	C
ATOM	1060	CG BMET B	8	-35.710	38.745	-0.476	0.50	16.74	BA	C
ATOM	1061	SD AMET B	8	-37.249	38.891	-0.844	0.50	19.13	BA	S
ATOM	1062	SD BMET B	8	-36.699	37.358	-1.133	0.50	15.35	BA	S
ATOM	1063	CE AMET B	8	-37.666	38.843	-2.601	0.50	19.36	BA	C
ATOM	1064	CE BMET B	8	-37.961	38.114	-2.137	0.50	13.60	BA	C
ATOM	1065	C MET B	8	-32.777	37.197	1.541	1.00	11.47	BA	C
ATOM	1066	O MET B	8	-33.270	36.716	2.573	1.00	14.29	BA	O
ATOM	1067	N GLY B	9	-31.560	37.722	1.407	1.00	12.39	BA	N
ATOM	1068	CA GLY B	9	-30.633	37.878	2.567	1.00	12.60	BA	C
ATOM	1069	C GLY B	9	-30.426	39.349	2.771	1.00	12.96	BA	C
ATOM	1070	O GLY B	9	-30.076	40.046	1.825	1.00	12.55	BA	O
ATOM	1071	N ASN B	10	-30.608	39.782	4.031	1.00	11.75	BA	N
ATOM	1072	CA ASN B	10	-30.589	41.253	4.290	1.00	12.30	BA	C
ATOM	1073	CB ASN B	10	-31.876	41.677	5.049	1.00	12.40	BA	C
ATOM	1074	CG ASN B	10	-31.983	41.064	6.482	1.00	12.93	BA	C
ATOM	1075	OD1 ASN B	10	-31.036	40.545	7.052	1.00	11.25	BA	O
ATOM	1076	ND2 ASN B	10	-33.187	41.091	7.051	1.00	14.57	BA	N
ATOM	1077	C ASN B	10	-29.340	41.671	5.068	1.00	11.37	BA	C
ATOM	1078	O ASN B	10	-28.404	40.799	5.282	1.00	11.61	BA	O
ATOM	1079	N ASP B	11	-29.214	42.917	5.555	1.00	9.81	BA	N
ATOM	1080	CA ASP B	11	-28.001	43.387	6.149	1.00	10.03	BA	C
ATOM	1081	CB ASP B	11	-28.004	44.912	6.245	1.00	11.30	BA	C
ATOM	1082	CG ASP B	11	-27.688	45.630	4.954	1.00	11.56	BA	C
ATOM	1083	OD1 ASP B	11	-27.233	45.057	3.949	1.00	14.08	BA	O
ATOM	1084	OD2 ASP B	11	-28.020	46.843	4.977	1.00	13.78	BA	O
ATOM	1085	C ASP B	11	-27.857	42.864	7.553	1.00	10.69	BA	C
ATOM	1086	O ASP B	11	-26.830	43.119	8.212	1.00	13.75	BA	O
ATOM	1087	N GLN B	12	-28.890	42.257	8.086	1.00	10.05	BA	N
ATOM	1088	CA GLN B	12	-28.898	41.650	9.439	1.00	11.41	BA	C
ATOM	1089	CB GLN B	12	-30.273	41.818	10.139	1.00	12.78	BA	C
ATOM	1090	CG GLN B	12	-30.597	43.299	10.612	1.00	11.20	BA	C
ATOM	1091	CD GLN B	12	-30.774	44.248	9.480	1.00	14.02	BA	C
ATOM	1092	OE1 GLN B	12	-31.530	43.982	8.538	1.00	15.38	BA	O
ATOM	1093	NE2 GLN B	12	-30.143	45.376	9.578	1.00	14.98	BA	N
ATOM	1094	C GLN B	12	-28.536	40.151	9.396	1.00	11.15	BA	C
ATOM	1095	O GLN B	12	-28.657	39.415	10.379	1.00	13.29	BA	O
ATOM	1096	N MET B	13	-28.052	39.678	8.268	1.00	9.80	BA	N
ATOM	1097	CA MET B	13	-27.594	38.281	8.149	1.00	10.96	BA	C

ATOM	1098	CB	MET	B	13	-26.328	37.954	8.922	1.00	14.82	BA	C
ATOM	1099	CG	MET	B	13	-25.127	38.630	8.287	1.00	15.75	BA	C
ATOM	1100	SD	MET	B	13	-23.980	39.247	9.499	1.00	15.28	BA	S
ATOM	1101	CE	MET	B	13	-24.970	40.531	10.261	1.00	16.47	BA	C
ATOM	1102	C	MET	B	13	-28.725	37.336	8.396	1.00	12.78	BA	C
ATOM	1103	O	MET	B	13	-28.576	36.293	9.125	1.00	12.40	BA	O
ATOM	1104	N	HIS	B	14	-29.842	37.625	7.758	1.00	11.08	BA	N
ATOM	1105	CA	HIS	B	14	-31.069	36.818	7.921	1.00	13.01	BA	C
ATOM	1106	CB	HIS	B	14	-32.114	37.646	8.724	1.00	13.49	BA	C
ATOM	1107	CG	HIS	B	14	-33.458	37.031	8.866	1.00	13.46	BA	C
ATOM	1108	ND1	HIS	B	14	-33.722	36.014	9.773	1.00	15.76	BA	N
ATOM	1109	CE1	HIS	B	14	-34.998	35.671	9.697	1.00	14.92	BA	C
ATOM	1110	NE2	HIS	B	14	-35.604	36.471	8.829	1.00	14.74	BA	N
ATOM	1111	CD2	HIS	B	14	-34.661	37.339	8.308	1.00	12.46	BA	C
ATOM	1112	C	HIS	B	14	-31.620	36.466	6.577	1.00	12.33	BA	C
ATOM	1113	O	HIS	B	14	-31.725	37.331	5.690	1.00	12.36	BA	O
ATOM	1114	N	PHE	B	15	-31.863	35.175	6.366	1.00	11.74	BA	N
ATOM	1115	CA	PHE	B	15	-32.765	34.755	5.305	1.00	12.67	BA	C
ATOM	1116	CB	PHE	B	15	-32.395	33.327	4.778	1.00	11.86	BA	C
ATOM	1117	CG	PHE	B	15	-31.100	33.242	3.936	1.00	11.84	BA	C
ATOM	1118	CD1	PHE	B	15	-30.868	34.074	2.821	1.00	12.45	BA	C
ATOM	1119	CE1	PHE	B	15	-29.742	33.863	2.032	1.00	11.94	BA	C
ATOM	1120	CZ	PHE	B	15	-28.823	32.808	2.362	1.00	11.38	BA	C
ATOM	1121	CE2	PHE	B	15	-29.030	31.966	3.386	1.00	10.71	BA	C
ATOM	1122	CD2	PHE	B	15	-30.206	32.188	4.186	1.00	12.35	BA	C
ATOM	1123	C	PHE	B	15	-34.182	34.637	5.909	1.00	10.92	BA	C
ATOM	1124	O	PHE	B	15	-34.351	34.048	6.993	1.00	13.33	BA	O
ATOM	1125	N	HIS	B	16	-35.231	35.082	5.231	1.00	11.62	BA	N
ATOM	1126	CA	HIS	B	16	-36.586	34.903	5.774	1.00	13.03	BA	C
ATOM	1127	CB	HIS	B	16	-37.594	35.874	5.117	1.00	13.84	BA	C
ATOM	1128	CG	HIS	B	16	-37.378	37.301	5.466	1.00	12.99	BA	C
ATOM	1129	ND1	HIS	B	16	-37.418	37.754	6.746	1.00	12.45	BA	N
ATOM	1130	CE1	HIS	B	16	-37.216	39.074	6.756	1.00	15.42	BA	C
ATOM	1131	NE2	HIS	B	16	-37.054	39.457	5.502	1.00	16.20	BA	N
ATOM	1132	CD2	HIS	B	16	-37.151	38.379	4.683	1.00	17.91	BA	C
ATOM	1133	C	HIS	B	16	-37.044	33.429	5.663	1.00	14.67	BA	C
ATOM	1134	O	HIS	B	16	-37.935	32.944	6.459	1.00	16.84	BA	O
ATOM	1135	N	THR	B	17	-36.420	32.689	4.761	1.00	13.54	BA	N
ATOM	1136	CA	THR	B	17	-36.778	31.267	4.483	1.00	14.97	BA	C
ATOM	1137	CB	THR	B	17	-36.515	30.976	3.021	1.00	14.07	BA	C
ATOM	1138	OG1	THR	B	17	-37.362	31.801	2.298	1.00	18.78	BA	O
ATOM	1139	CG2	THR	B	17	-36.741	29.518	2.594	1.00	16.93	BA	C
ATOM	1140	C	THR	B	17	-35.903	30.323	5.250	1.00	13.39	BA	C
ATOM	1141	O	THR	B	17	-34.675	30.400	5.192	1.00	14.30	BA	O
ATOM	1142	N	ASN	B	18	-36.544	29.409	6.009	1.00	13.68	BA	N
ATOM	1143	CA	ASN	B	18	-35.832	28.423	6.783	1.00	14.78	BA	C
ATOM	1144	CB	ASN	B	18	-36.613	28.204	8.138	1.00	14.20	BA	C
ATOM	1145	CG	ASN	B	18	-38.076	27.598	7.950	1.00	17.33	BA	C
ATOM	1146	OD1	ASN	B	18	-38.488	26.538	8.582	1.00	16.28	BA	O
ATOM	1147	ND2	ASN	B	18	-38.935	28.384	7.229	1.00	14.51	BA	N
ATOM	1148	C	ASN	B	18	-35.636	27.079	6.030	1.00	12.84	BA	C
ATOM	1149	O	ASN	B	18	-34.753	26.242	6.406	1.00	12.30	BA	O
ATOM	1150	N	ALA	B	19	-36.533	26.792	5.101	1.00	11.18	BA	N
ATOM	1151	CA	ALA	B	19	-36.674	25.474	4.560	1.00	11.39	BA	C
ATOM	1152	CB	ALA	B	19	-37.596	24.587	5.454	1.00	13.31	BA	C

ATOM	1153	C	ALA	B	19	-37.244	25.634	3.154	1.00	10.60	BA	C
ATOM	1154	O	ALA	B	19	-38.195	26.346	2.946	1.00	13.04	BA	O
ATOM	1155	N	ILE	B	20	-36.714	24.855	2.234	1.00	11.94	BA	N
ATOM	1156	CA	ILE	B	20	-37.170	24.845	0.819	1.00	10.62	BA	C
ATOM	1157	CB	ILE	B	20	-35.977	25.260	-0.140	1.00	11.47	BA	C
ATOM	1158	CG1	ILE	B	20	-35.564	26.709	0.143	1.00	11.80	BA	C
ATOM	1159	CD1	ILE	B	20	-34.207	27.100	-0.450	1.00	13.13	BA	C
ATOM	1160	CG2	ILE	B	20	-36.316	25.027	-1.605	1.00	13.29	BA	C
ATOM	1161	C	ILE	B	20	-37.575	23.429	0.482	1.00	10.50	BA	C
ATOM	1162	O	ILE	B	20	-36.833	22.462	0.686	1.00	12.90	BA	O
ATOM	1163	N	THR	B	21	-38.755	23.319	-0.117	1.00	10.86	BA	N
ATOM	1164	CA	THR	B	21	-39.186	21.997	-0.627	1.00	12.54	BA	C
ATOM	1165	CB	THR	B	21	-40.624	21.724	-0.147	1.00	14.08	BA	C
ATOM	1166	OG1	THR	B	21	-40.632	21.725	1.287	1.00	15.54	BA	O
ATOM	1167	CG2	THR	B	21	-41.112	20.358	-0.754	1.00	14.86	BA	C
ATOM	1168	C	THR	B	21	-39.180	22.068	-2.118	1.00	12.93	BA	C
ATOM	1169	O	THR	B	21	-39.861	22.946	-2.707	1.00	16.96	BA	O
ATOM	1170	N	VAL	B	22	-38.539	21.142	-2.763	1.00	11.09	BA	N
ATOM	1171	CA	VAL	B	22	-38.487	21.069	-4.220	1.00	11.84	BA	C
ATOM	1172	CB	VAL	B	22	-37.051	20.700	-4.642	1.00	11.47	BA	C
ATOM	1173	CG1	VAL	B	22	-36.979	20.467	-6.163	1.00	12.80	BA	C
ATOM	1174	CG2	VAL	B	22	-36.049	21.787	-4.142	1.00	12.72	BA	C
ATOM	1175	C	VAL	B	22	-39.468	20.012	-4.684	1.00	12.38	BA	C
ATOM	1176	O	VAL	B	22	-39.362	18.840	-4.239	1.00	13.38	BA	O
ATOM	1177	N	ASP	B	23	-40.393	20.381	-5.543	1.00	12.05	BA	N
ATOM	1178	CA	ASP	B	23	-41.354	19.442	-6.079	1.00	10.10	BA	C
ATOM	1179	CB	ASP	B	23	-42.493	20.295	-6.780	1.00	14.03	BA	C
ATOM	1180	CG	ASP	B	23	-43.488	19.482	-7.551	1.00	14.81	BA	C
ATOM	1181	OD1	ASP	B	23	-43.324	18.278	-7.765	1.00	15.42	BA	O
ATOM	1182	OD2	ASP	B	23	-44.539	20.099	-7.863	1.00	18.09	BA	O
ATOM	1183	C	ASP	B	23	-40.559	18.511	-6.979	1.00	12.26	BA	C
ATOM	1184	O	ASP	B	23	-39.985	18.944	-8.026	1.00	12.77	BA	O
ATOM	1185	N	LYS	B	24	-40.683	17.200	-6.667	1.00	12.71	BA	N
ATOM	1186	CA	LYS	B	24	-39.866	16.251	-7.407	1.00	15.03	BA	C
ATOM	1187	CB	LYS	B	24	-39.801	14.887	-6.679	1.00	14.87	BA	C
ATOM	1188	CG	LYS	B	24	-41.176	14.194	-6.756	1.00	14.68	BA	C
ATOM	1189	CD	LYS	B	24	-41.181	12.930	-5.820	1.00	14.75	BA	C
ATOM	1190	CE	LYS	B	24	-42.515	12.214	-5.753	1.00	12.15	BA	C
ATOM	1191	NZ	LYS	B	24	-43.029	11.871	-7.128	1.00	13.60	BA	N
ATOM	1192	C	LYS	B	24	-40.325	16.083	-8.866	1.00	14.27	BA	C
ATOM	1193	O	LYS	B	24	-39.650	15.420	-9.645	1.00	15.60	BA	O
ATOM	1194	N	SER	B	25	-41.506	16.601	-9.248	1.00	13.17	BA	N
ATOM	1195	CA	SER	B	25	-41.880	16.582	-10.679	1.00	12.14	BA	C
ATOM	1196	CB	SER	B	25	-43.371	16.724	-10.838	1.00	14.12	BA	C
ATOM	1197	OG	SER	B	25	-43.814	17.967	-10.483	1.00	15.12	BA	O
ATOM	1198	C	SER	B	25	-41.156	17.635	-11.537	1.00	12.40	BA	C
ATOM	1199	O	SER	B	25	-41.314	17.647	-12.764	1.00	15.21	BA	O
ATOM	1200	N	CYS	B	26	-40.525	18.603	-10.874	1.00	11.03	BA	N
ATOM	1201	CA	CYS	B	26	-39.754	19.632	-11.603	1.00	13.09	BA	C
ATOM	1202	CB	CYS	B	26	-39.343	20.783	-10.608	1.00	12.47	BA	C
ATOM	1203	SG	CYS	B	26	-40.633	21.707	-9.935	1.00	11.92	BA	S
ATOM	1204	C	CYS	B	26	-38.511	18.996	-12.236	1.00	11.21	BA	C
ATOM	1205	O	CYS	B	26	-37.803	18.280	-11.592	1.00	15.18	BA	O
ATOM	1206	N	LYS	B	27	-38.283	19.285	-13.492	1.00	11.00	BA	N
ATOM	1207	CA	ALYS	B	27	-37.036	18.821	-14.123	0.50	10.76	BA	C

ATOM	1208	CA	BLYS	B	27	-37.038	18.822	-14.112	0.50	10.58	BA	C
ATOM	1209	CB	ALYS	B	27	-37.120	18.927	-15.637	0.50	12.25	BA	C
ATOM	1210	CB	BLYS	B	27	-37.155	18.922	-15.615	0.50	12.02	BA	C
ATOM	1211	CG	ALYS	B	27	-38.215	18.104	-16.322	0.50	13.24	BA	C
ATOM	1212	CG	BLYS	B	27	-35.895	18.538	-16.361	0.50	11.94	BA	C
ATOM	1213	CD	ALYS	B	27	-38.328	16.671	-15.833	0.50	17.25	BA	C
ATOM	1214	CD	BLYS	B	27	-36.112	18.729	-17.815	0.50	18.60	BA	C
ATOM	1215	CE	ALYS	B	27	-39.109	15.806	-16.856	0.50	19.87	BA	C
ATOM	1216	CE	BLYS	B	27	-34.939	19.390	-18.441	0.50	18.78	BA	C
ATOM	1217	NZ	ALYS	B	27	-40.251	16.552	-17.591	0.50	20.27	BA	N
ATOM	1218	NZ	BLYS	B	27	-35.383	19.829	-19.752	0.50	21.39	BA	N
ATOM	1219	C	LYS	B	27	-35.838	19.640	-13.595	1.00	11.40	BA	C
ATOM	1220	O	LYS	B	27	-34.747	19.097	-13.369	1.00	11.14	BA	O
ATOM	1221	N	GLN	B	28	-36.073	20.934	-13.438	1.00	10.25	BA	N
ATOM	1222	CA	GLN	B	28	-35.060	21.836	-12.913	1.00	9.52	BA	C
ATOM	1223	CB	GLN	B	28	-34.395	22.704	-14.007	1.00	10.40	BA	C
ATOM	1224	CG	GLN	B	28	-33.761	21.920	-15.229	1.00	13.06	BA	C
ATOM	1225	CD	GLN	B	28	-33.087	22.831	-16.155	1.00	17.00	BA	C
ATOM	1226	OE1	GLN	B	28	-31.846	22.914	-16.176	1.00	24.03	BA	O
ATOM	1227	NE2	GLN	B	28	-33.854	23.525	-16.924	1.00	21.87	BA	N
ATOM	1228	C	GLN	B	28	-35.677	22.707	-11.863	1.00	9.89	BA	C
ATOM	1229	O	GLN	B	28	-36.917	22.968	-11.931	1.00	12.18	BA	O
ATOM	1230	N	PHE	B	29	-34.859	23.238	-10.943	1.00	10.40	BA	N
ATOM	1231	CA	PHE	B	29	-35.347	24.110	-9.844	1.00	9.61	BA	C
ATOM	1232	CB	PHE	B	29	-35.231	23.376	-8.502	1.00	12.00	BA	C
ATOM	1233	CG	PHE	B	29	-35.823	24.136	-7.408	1.00	11.97	BA	C
ATOM	1234	CD1	PHE	B	29	-37.201	24.104	-7.184	1.00	12.56	BA	C
ATOM	1235	CE1	PHE	B	29	-37.779	24.859	-6.100	1.00	12.77	BA	C
ATOM	1236	CZ	PHE	B	29	-36.999	25.552	-5.275	1.00	13.23	BA	C
ATOM	1237	CE2	PHE	B	29	-35.633	25.566	-5.450	1.00	14.44	BA	C
ATOM	1238	CD2	PHE	B	29	-35.017	24.848	-6.511	1.00	13.37	BA	C
ATOM	1239	C	PHE	B	29	-34.523	25.363	-9.880	1.00	9.40	BA	C
ATOM	1240	O	PHE	B	29	-33.305	25.314	-10.118	1.00	9.79	BA	O
ATOM	1241	N	THR	B	30	-35.167	26.483	-9.673	1.00	9.49	BA	N
ATOM	1242	CA	THR	B	30	-34.502	27.817	-9.763	1.00	9.74	BA	C
ATOM	1243	CB	THR	B	30	-35.208	28.653	-10.891	1.00	8.97	BA	C
ATOM	1244	OG1	THR	B	30	-35.084	27.942	-12.126	1.00	10.85	BA	O
ATOM	1245	CG2	THR	B	30	-34.509	30.025	-11.046	1.00	11.21	BA	C
ATOM	1246	C	THR	B	30	-34.642	28.562	-8.446	1.00	10.21	BA	C
ATOM	1247	O	THR	B	30	-35.742	28.712	-7.908	1.00	11.19	BA	O
ATOM	1248	N	VAL	B	31	-33.489	29.020	-7.942	1.00	10.10	BA	N
ATOM	1249	CA	VAL	B	31	-33.438	29.864	-6.765	1.00	11.34	BA	C
ATOM	1250	CB	VAL	B	31	-32.359	29.333	-5.802	1.00	11.68	BA	C
ATOM	1251	CG1	VAL	B	31	-32.166	30.353	-4.629	1.00	14.55	BA	C
ATOM	1252	CG2	VAL	B	31	-32.727	27.958	-5.246	1.00	14.60	BA	C
ATOM	1253	C	VAL	B	31	-33.066	31.257	-7.212	1.00	10.79	BA	C
ATOM	1254	O	VAL	B	31	-32.071	31.490	-7.904	1.00	11.57	BA	O
ATOM	1255	N	ASN	B	32	-33.948	32.215	-6.826	1.00	11.33	BA	N
ATOM	1256	CA	ASN	B	32	-33.785	33.656	-7.174	1.00	11.05	BA	C
ATOM	1257	CB	ASN	B	32	-35.129	34.227	-7.747	1.00	12.26	BA	C
ATOM	1258	CG	ASN	B	32	-35.593	33.428	-8.915	1.00	13.41	BA	C
ATOM	1259	OD1	ASN	B	32	-34.859	33.345	-9.906	1.00	12.11	BA	O
ATOM	1260	ND2	ASN	B	32	-36.786	32.878	-8.850	1.00	15.08	BA	N
ATOM	1261	C	ASN	B	32	-33.345	34.367	-5.886	1.00	10.80	BA	C
ATOM	1262	O	ASN	B	32	-34.184	34.505	-4.999	1.00	11.62	BA	O

ATOM	1263	N	LEU	B	33	-32.044	34.732	-5.820	1.00	11.41	BA	N
ATOM	1264	CA	LEU	B	33	-31.487	35.397	-4.577	1.00	10.97	BA	C
ATOM	1265	CB	LEU	B	33	-30.047	34.913	-4.386	1.00	11.56	BA	C
ATOM	1266	CG	LEU	B	33	-29.419	35.503	-3.105	1.00	11.79	BA	C
ATOM	1267	CD1	LEU	B	33	-30.169	35.065	-1.806	1.00	13.41	BA	C
ATOM	1268	CD2	LEU	B	33	-27.980	35.115	-3.190	1.00	12.76	BA	C
ATOM	1269	C	LEU	B	33	-31.524	36.902	-4.800	1.00	12.61	BA	C
ATOM	1270	O	LEU	B	33	-31.099	37.408	-5.873	1.00	13.48	BA	O
ATOM	1271	N	SER	B	34	-31.913	37.604	-3.749	1.00	13.27	BA	N
ATOM	1272	CA	ASER	B	34	-31.670	39.043	-3.771	0.50	13.45	BA	C
ATOM	1273	CA	BSER	B	34	-31.953	39.072	-3.688	0.50	14.34	BA	C
ATOM	1274	CB	ASER	B	34	-32.926	39.808	-4.140	0.50	13.19	BA	C
ATOM	1275	CB	BSER	B	34	-33.417	39.583	-3.594	0.50	14.87	BA	C
ATOM	1276	OG	ASER	B	34	-33.946	39.480	-3.201	0.50	11.42	BA	O
ATOM	1277	OG	BSER	B	34	-34.222	39.224	-4.735	0.50	15.85	BA	O
ATOM	1278	C	SER	B	34	-31.179	39.513	-2.415	1.00	14.07	BA	C
ATOM	1279	O	SER	B	34	-31.146	38.755	-1.422	1.00	13.33	BA	O
ATOM	1280	N	HIS	B	35	-30.708	40.772	-2.382	1.00	14.49	BA	N
ATOM	1281	CA	HIS	B	35	-30.017	41.378	-1.200	1.00	12.67	BA	C
ATOM	1282	CB	HIS	B	35	-28.537	41.469	-1.488	1.00	14.34	BA	C
ATOM	1283	CG	HIS	B	35	-27.692	41.818	-0.294	1.00	13.00	BA	C
ATOM	1284	ND1	HIS	B	35	-28.068	42.706	0.729	1.00	14.61	BA	N
ATOM	1285	CE1	HIS	B	35	-27.073	42.763	1.608	1.00	13.51	BA	C
ATOM	1286	NE2	HIS	B	35	-26.073	42.021	1.139	1.00	11.88	BA	N
ATOM	1287	CD2	HIS	B	35	-26.487	41.326	0.032	1.00	12.52	BA	C
ATOM	1288	C	HIS	B	35	-30.630	42.789	-1.028	1.00	15.14	BA	C
ATOM	1289	O	HIS	B	35	-30.054	43.765	-1.477	1.00	13.57	BA	O
ATOM	1290	N	PRO	B	36	-31.689	42.870	-0.215	1.00	13.91	BA	N
ATOM	1291	CA	PRO	B	36	-32.199	44.146	0.328	1.00	17.00	BA	C
ATOM	1292	CB	PRO	B	36	-33.311	43.667	1.245	1.00	16.47	BA	C
ATOM	1293	CG	PRO	B	36	-33.676	42.445	0.782	1.00	18.82	BA	C
ATOM	1294	CD	PRO	B	36	-32.514	41.788	0.272	1.00	16.69	BA	C
ATOM	1295	C	PRO	B	36	-31.229	44.686	1.333	1.00	17.64	BA	C
ATOM	1296	O	PRO	B	36	-30.490	43.901	1.953	1.00	18.41	BA	O
ATOM	1297	N	GLY	B	37	-31.425	45.967	1.577	1.00	19.46	BA	N
ATOM	1298	CA	GLY	B	37	-30.542	46.691	2.550	1.00	18.53	BA	C
ATOM	1299	C	GLY	B	37	-29.570	47.505	1.727	1.00	18.40	BA	C
ATOM	1300	O	GLY	B	37	-29.823	47.867	0.550	1.00	19.49	BA	O
ATOM	1301	N	ASN	B	38	-28.458	47.847	2.363	1.00	15.13	BA	N
ATOM	1302	CA	ASN	B	38	-27.532	48.890	1.887	1.00	15.72	BA	C
ATOM	1303	CB	ASN	B	38	-27.565	50.157	2.782	1.00	17.11	BA	C
ATOM	1304	CG	ASN	B	38	-28.920	50.544	3.209	1.00	24.71	BA	C
ATOM	1305	OD1	ASN	B	38	-29.224	50.590	4.424	1.00	30.91	BA	O
ATOM	1306	ND2	ASN	B	38	-29.744	50.881	2.236	1.00	25.87	BA	N
ATOM	1307	C	ASN	B	38	-26.052	48.575	1.846	1.00	14.91	BA	C
ATOM	1308	O	ASN	B	38	-25.279	49.402	1.333	1.00	14.42	BA	O
ATOM	1309	N	LEU	B	39	-25.638	47.414	2.372	1.00	12.29	BA	N
ATOM	1310	CA	LEU	B	39	-24.229	47.168	2.504	1.00	13.02	BA	C
ATOM	1311	CB	LEU	B	39	-23.967	46.436	3.817	1.00	12.31	BA	C
ATOM	1312	CG	LEU	B	39	-24.202	47.184	5.129	1.00	11.33	BA	C
ATOM	1313	CD1	LEU	B	39	-23.945	46.306	6.352	1.00	12.77	BA	C
ATOM	1314	CD2	LEU	B	39	-23.230	48.407	5.230	1.00	11.44	BA	C
ATOM	1315	C	LEU	B	39	-23.708	46.386	1.314	1.00	12.73	BA	C
ATOM	1316	O	LEU	B	39	-24.410	45.511	0.800	1.00	13.44	BA	O
ATOM	1317	N	PRO	B	40	-22.460	46.697	0.880	1.00	13.62	BA	N

ATOM	1318	CA	PRO	B	40	-21.926	46.088	-0.325	1.00	14.12	BA	C
ATOM	1319	CB	PRO	B	40	-20.850	47.101	-0.798	1.00	15.00	BA	C
ATOM	1320	CG	PRO	B	40	-20.338	47.589	0.381	1.00	13.78	BA	C
ATOM	1321	CD	PRO	B	40	-21.583	47.806	1.297	1.00	13.12	BA	C
ATOM	1322	C	PRO	B	40	-21.317	44.703	-0.061	1.00	13.58	BA	C
ATOM	1323	O	PRO	B	40	-21.216	44.313	1.146	1.00	13.16	BA	O
ATOM	1324	N	LYS	B	41	-20.916	44.019	-1.133	1.00	13.32	BA	N
ATOM	1325	CA	LYS	B	41	-20.496	42.619	-1.049	1.00	13.04	BA	C
ATOM	1326	CB	LYS	B	41	-20.515	41.854	-2.395	1.00	17.15	BA	C
ATOM	1327	CG	LYS	B	41	-19.366	42.160	-3.275	1.00	19.69	BA	C
ATOM	1328	CD	LYS	B	41	-19.553	41.307	-4.532	1.00	19.95	BA	C
ATOM	1329	CE	LYS	B	41	-18.283	41.470	-5.312	1.00	22.00	BA	C
ATOM	1330	NZ	LYS	B	41	-18.356	40.905	-6.701	1.00	22.08	BA	N
ATOM	1331	C	LYS	B	41	-19.222	42.398	-0.231	1.00	12.78	BA	C
ATOM	1332	O	LYS	B	41	-19.074	41.304	0.319	1.00	12.66	BA	O
ATOM	1333	N	ASN	B	42	-18.322	43.405	-0.117	1.00	10.48	BA	N
ATOM	1334	CA	ASN	B	42	-17.074	43.119	0.614	1.00	12.08	BA	C
ATOM	1335	CB	ASN	B	42	-15.975	44.105	0.214	1.00	12.03	BA	C
ATOM	1336	CG	ASN	B	42	-16.246	45.547	0.656	1.00	15.06	BA	C
ATOM	1337	OD1	ASN	B	42	-17.334	45.907	1.116	1.00	15.57	BA	O
ATOM	1338	ND2	ASN	B	42	-15.223	46.420	0.451	1.00	16.51	BA	N
ATOM	1339	C	ASN	B	42	-17.236	42.958	2.108	1.00	10.98	BA	C
ATOM	1340	O	ASN	B	42	-16.374	42.355	2.769	1.00	13.31	BA	O
ATOM	1341	N	VAL	B	43	-18.367	43.451	2.639	1.00	11.97	BA	N
ATOM	1342	CA	VAL	B	43	-18.656	43.383	4.128	1.00	11.16	BA	C
ATOM	1343	CB	VAL	B	43	-18.643	44.759	4.858	1.00	11.03	BA	C
ATOM	1344	CG1	VAL	B	43	-17.252	45.377	4.717	1.00	13.16	BA	C
ATOM	1345	CG2	VAL	B	43	-19.583	45.762	4.153	1.00	12.68	BA	C
ATOM	1346	C	VAL	B	43	-19.969	42.667	4.424	1.00	11.05	BA	C
ATOM	1347	O	VAL	B	43	-20.165	42.233	5.550	1.00	11.64	BA	O
ATOM	1348	N	MET	B	44	-20.841	42.510	3.403	1.00	9.87	BA	N
ATOM	1349	CA	MET	B	44	-22.176	41.917	3.603	1.00	11.02	BA	C
ATOM	1350	CB	MET	B	44	-23.201	42.986	3.998	1.00	12.54	BA	C
ATOM	1351	CG	MET	B	44	-24.499	42.450	4.609	1.00	11.34	BA	C
ATOM	1352	SD	MET	B	44	-24.391	41.310	5.941	1.00	12.97	BA	S
ATOM	1353	CE	MET	B	44	-23.562	42.345	7.180	1.00	14.07	BA	C
ATOM	1354	C	MET	B	44	-22.632	41.110	2.404	1.00	11.11	BA	C
ATOM	1355	O	MET	B	44	-23.830	41.056	2.064	1.00	12.64	BA	O
ATOM	1356	N	GLY	B	45	-21.646	40.431	1.801	1.00	10.28	BA	N
ATOM	1357	CA	GLY	B	45	-21.984	39.611	0.601	1.00	10.39	BA	C
ATOM	1358	C	GLY	B	45	-22.810	38.416	1.034	1.00	10.71	BA	C
ATOM	1359	O	GLY	B	45	-22.581	37.828	2.153	1.00	10.83	BA	O
ATOM	1360	N	HIS	B	46	-23.785	38.053	0.196	1.00	11.01	BA	N
ATOM	1361	CA	HIS	B	46	-24.494	36.789	0.367	1.00	10.19	BA	C
ATOM	1362	CB	HIS	B	46	-25.968	36.994	0.759	1.00	10.51	BA	C
ATOM	1363	CG	HIS	B	46	-26.145	37.805	2.015	1.00	9.67	BA	C
ATOM	1364	ND1	HIS	B	46	-25.429	37.619	3.167	1.00	9.13	BA	N
ATOM	1365	CE1	HIS	B	46	-25.868	38.466	4.109	1.00	8.76	BA	C
ATOM	1366	NE2	HIS	B	46	-26.837	39.199	3.588	1.00	11.40	BA	N
ATOM	1367	CD2	HIS	B	46	-27.029	38.811	2.281	1.00	12.82	BA	C
ATOM	1368	C	HIS	B	46	-24.457	35.959	-0.875	1.00	10.92	BA	C
ATOM	1369	O	HIS	B	46	-24.604	36.511	-1.982	1.00	11.66	BA	O
ATOM	1370	N	ASN	B	47	-24.423	34.661	-0.678	1.00	10.26	BA	N
ATOM	1371	CA	ASN	B	47	-24.663	33.690	-1.754	1.00	10.35	BA	C
ATOM	1372	CB	ASN	B	47	-23.381	33.044	-2.325	1.00	10.91	BA	C

ATOM	1373	CG	ASN	B	47	-22.507	32.459	-1.317	1.00	11.47	BA	C
ATOM	1374	OD1	ASN	B	47	-22.914	32.207	-0.202	1.00	10.34	BA	O
ATOM	1375	ND2	ASN	B	47	-21.257	32.226	-1.704	1.00	11.47	BA	N
ATOM	1376	C	ASN	B	47	-25.655	32.683	-1.193	1.00	9.26	BA	C
ATOM	1377	O	ASN	B	47	-26.012	32.729	-0.001	1.00	11.03	BA	O
ATOM	1378	N	TRP	B	48	-26.193	31.822	-2.064	1.00	9.66	BA	N
ATOM	1379	CA	TRP	B	48	-26.960	30.643	-1.705	1.00	10.05	BA	C
ATOM	1380	CB	TRP	B	48	-28.389	30.800	-2.310	1.00	11.35	BA	C
ATOM	1381	CG	TRP	B	48	-29.316	29.688	-1.978	1.00	10.37	BA	C
ATOM	1382	CD1	TRP	B	48	-30.181	29.672	-0.896	1.00	11.86	BA	C
ATOM	1383	NE1	TRP	B	48	-30.931	28.518	-0.941	1.00	11.44	BA	N
ATOM	1384	CE2	TRP	B	48	-30.489	27.748	-1.934	1.00	10.39	BA	C
ATOM	1385	CD2	TRP	B	48	-29.494	28.447	-2.663	1.00	10.44	BA	C
ATOM	1386	CE3	TRP	B	48	-28.939	27.861	-3.804	1.00	11.47	BA	C
ATOM	1387	CZ3	TRP	B	48	-29.391	26.542	-4.164	1.00	12.22	BA	C
ATOM	1388	CH2	TRP	B	48	-30.394	25.920	-3.371	1.00	11.23	BA	C
ATOM	1389	CZ2	TRP	B	48	-30.944	26.492	-2.315	1.00	10.53	BA	C
ATOM	1390	C	TRP	B	48	-26.224	29.474	-2.251	1.00	10.22	BA	C
ATOM	1391	O	TRP	B	48	-26.000	29.373	-3.463	1.00	10.69	BA	O
ATOM	1392	N	VAL	B	49	-25.831	28.570	-1.350	1.00	9.51	BA	N
ATOM	1393	CA	VAL	B	49	-25.012	27.419	-1.692	1.00	9.89	BA	C
ATOM	1394	CB	VAL	B	49	-23.595	27.609	-1.042	1.00	9.77	BA	C
ATOM	1395	CG1	VAL	B	49	-22.663	26.441	-1.429	1.00	12.01	BA	C
ATOM	1396	CG2	VAL	B	49	-22.975	28.975	-1.475	1.00	11.72	BA	C
ATOM	1397	C	VAL	B	49	-25.655	26.180	-1.102	1.00	9.53	BA	C
ATOM	1398	O	VAL	B	49	-26.129	26.222	0.008	1.00	10.35	BA	O
ATOM	1399	N	LEU	B	50	-25.705	25.103	-1.889	1.00	9.79	BA	N
ATOM	1400	CA	LEU	B	50	-26.412	23.868	-1.477	1.00	9.65	BA	C
ATOM	1401	CB	LEU	B	50	-27.437	23.543	-2.585	1.00	10.45	BA	C
ATOM	1402	CG	LEU	B	50	-28.372	22.364	-2.275	1.00	10.50	BA	C
ATOM	1403	CD1	LEU	B	50	-29.269	22.588	-1.064	1.00	11.79	BA	C
ATOM	1404	CD2	LEU	B	50	-29.178	21.940	-3.572	1.00	11.53	BA	C
ATOM	1405	C	LEU	B	50	-25.398	22.738	-1.408	1.00	9.16	BA	C
ATOM	1406	O	LEU	B	50	-24.579	22.515	-2.341	1.00	10.70	BA	O
ATOM	1407	N	SER	B	51	-25.420	21.981	-0.324	1.00	9.31	BA	N
ATOM	1408	CA	SER	B	51	-24.604	20.728	-0.185	1.00	9.52	BA	C
ATOM	1409	CB	SER	B	51	-23.237	21.094	0.403	1.00	9.99	BA	C
ATOM	1410	OG	SER	B	51	-23.320	21.395	1.819	1.00	10.73	BA	O
ATOM	1411	C	SER	B	51	-25.322	19.733	0.693	1.00	10.09	BA	C
ATOM	1412	O	SER	B	51	-26.376	20.028	1.300	1.00	10.20	BA	O
ATOM	1413	N	THR	B	52	-24.731	18.542	0.804	1.00	10.24	BA	N
ATOM	1414	CA	THR	B	52	-25.212	17.663	1.855	1.00	9.45	BA	C
ATOM	1415	CB	THR	B	52	-24.597	16.281	1.745	1.00	8.98	BA	C
ATOM	1416	OG1	THR	B	52	-23.204	16.454	1.998	1.00	11.09	BA	O
ATOM	1417	CG2	THR	B	52	-24.878	15.651	0.391	1.00	10.55	BA	C
ATOM	1418	C	THR	B	52	-24.876	18.290	3.226	1.00	8.95	BA	C
ATOM	1419	O	THR	B	52	-24.008	19.181	3.354	1.00	9.41	BA	O
ATOM	1420	N	ALA	B	53	-25.588	17.857	4.270	1.00	10.92	BA	N
ATOM	1421	CA	ALA	B	53	-25.267	18.293	5.674	1.00	12.94	BA	C
ATOM	1422	CB	ALA	B	53	-26.180	17.604	6.666	1.00	13.65	BA	C
ATOM	1423	C	ALA	B	53	-23.822	17.983	6.031	1.00	11.99	BA	C
ATOM	1424	O	ALA	B	53	-23.170	18.801	6.629	1.00	11.95	BA	O
ATOM	1425	N	ALA	B	54	-23.336	16.789	5.601	1.00	11.72	BA	N
ATOM	1426	CA	ALA	B	54	-21.996	16.355	5.967	1.00	12.75	BA	C
ATOM	1427	CB	ALA	B	54	-21.737	14.908	5.701	1.00	12.97	BA	C

ATOM	1428	C	ALA	B	54	-20.907	17.198	5.337	1.00	10.81	BA	C
ATOM	1429	O	ALA	B	54	-19.819	17.374	5.887	1.00	11.17	BA	O
ATOM	1430	N	ASP	B	55	-21.271	17.789	4.181	1.00	10.07	BA	N
ATOM	1431	CA	ASP	B	55	-20.285	18.580	3.464	1.00	11.16	BA	C
ATOM	1432	CB	ASP	B	55	-20.499	18.465	1.947	1.00	10.36	BA	C
ATOM	1433	CG	ASP	B	55	-19.921	17.185	1.342	1.00	11.95	BA	C
ATOM	1434	OD1	ASP	B	55	-18.999	16.571	1.977	1.00	12.95	BA	O
ATOM	1435	OD2	ASP	B	55	-20.337	16.728	0.312	1.00	10.74	BA	O
ATOM	1436	C	ASP	B	55	-20.338	20.087	3.757	1.00	10.52	BA	C
ATOM	1437	O	ASP	B	55	-19.430	20.836	3.353	1.00	11.22	BA	O
ATOM	1438	N	MET	B	56	-21.367	20.563	4.459	1.00	10.69	BA	N
ATOM	1439	CA	MET	B	56	-21.540	22.024	4.636	1.00	9.66	BA	C
ATOM	1440	CB	MET	B	56	-22.778	22.306	5.488	1.00	9.07	BA	C
ATOM	1441	CG	MET	B	56	-22.878	23.758	5.877	1.00	9.77	BA	C
ATOM	1442	SD	MET	B	56	-24.455	24.082	6.724	1.00	11.20	BA	S
ATOM	1443	CE	MET	B	56	-24.117	25.764	7.260	1.00	13.34	BA	C
ATOM	1444	C	MET	B	56	-20.297	22.651	5.284	1.00	9.83	BA	C
ATOM	1445	O	MET	B	56	-19.785	23.665	4.855	1.00	10.20	BA	O
ATOM	1446	N	GLN	B	57	-19.817	22.054	6.379	1.00	10.02	BA	N
ATOM	1447	CA	GLN	B	57	-18.762	22.719	7.141	1.00	11.12	BA	C
ATOM	1448	CB	GLN	B	57	-18.372	21.961	8.429	1.00	11.01	BA	C
ATOM	1449	CG	GLN	B	57	-17.238	22.706	9.194	1.00	14.30	BA	C
ATOM	1450	CD	GLN	B	57	-17.717	23.989	9.742	1.00	14.19	BA	C
ATOM	1451	OE1	GLN	B	57	-18.726	23.995	10.528	1.00	16.83	BA	O
ATOM	1452	NE2	GLN	B	57	-17.212	25.111	9.236	1.00	17.50	BA	N
ATOM	1453	C	GLN	B	57	-17.491	22.888	6.298	1.00	10.51	BA	C
ATOM	1454	O	GLN	B	57	-16.840	23.979	6.341	1.00	10.59	BA	O
ATOM	1455	N	GLY	B	58	-17.142	21.842	5.520	1.00	10.53	BA	N
ATOM	1456	CA	GLY	B	58	-15.969	21.929	4.642	1.00	11.19	BA	C
ATOM	1457	C	GLY	B	58	-16.112	22.950	3.504	1.00	9.56	BA	C
ATOM	1458	O	GLY	B	58	-15.185	23.686	3.192	1.00	10.93	BA	O
ATOM	1459	N	VAL	B	59	-17.324	23.004	2.894	1.00	9.48	BA	N
ATOM	1460	CA	VAL	B	59	-17.618	24.014	1.869	1.00	10.51	BA	C
ATOM	1461	CB	VAL	B	59	-19.039	23.834	1.277	1.00	10.31	BA	C
ATOM	1462	CG1	VAL	B	59	-19.439	24.945	0.346	1.00	11.12	BA	C
ATOM	1463	CG2	VAL	B	59	-19.092	22.507	0.470	1.00	10.01	BA	C
ATOM	1464	C	VAL	B	59	-17.436	25.418	2.485	1.00	11.29	BA	C
ATOM	1465	O	VAL	B	59	-16.840	26.270	1.860	1.00	11.19	BA	O
ATOM	1466	N	VAL	B	60	-17.973	25.633	3.691	1.00	10.19	BA	N
ATOM	1467	CA	VAL	B	60	-17.917	26.924	4.373	1.00	10.64	BA	C
ATOM	1468	CB	VAL	B	60	-18.791	26.903	5.649	1.00	10.08	BA	C
ATOM	1469	CG1	VAL	B	60	-18.494	28.156	6.477	1.00	11.22	BA	C
ATOM	1470	CG2	VAL	B	60	-20.269	26.894	5.194	1.00	11.45	BA	C
ATOM	1471	C	VAL	B	60	-16.468	27.329	4.684	1.00	9.90	BA	C
ATOM	1472	O	VAL	B	60	-16.042	28.434	4.390	1.00	10.49	BA	O
ATOM	1473	N	THR	B	61	-15.743	26.405	5.286	1.00	11.02	BA	N
ATOM	1474	CA	THR	B	61	-14.381	26.717	5.724	1.00	12.03	BA	C
ATOM	1475	CB	THR	B	61	-13.885	25.557	6.534	1.00	13.72	BA	C
ATOM	1476	OG1	THR	B	61	-14.542	25.601	7.773	1.00	15.66	BA	O
ATOM	1477	CG2	THR	B	61	-12.340	25.693	6.863	1.00	15.08	BA	C
ATOM	1478	C	THR	B	61	-13.486	26.924	4.504	1.00	11.64	BA	C
ATOM	1479	O	THR	B	61	-12.728	27.907	4.415	1.00	13.52	BA	O
ATOM	1480	N	ASP	B	62	-13.543	26.025	3.522	1.00	11.33	BA	N
ATOM	1481	CA	ASP	B	62	-12.724	26.188	2.285	1.00	11.64	BA	C
ATOM	1482	CB	ASP	B	62	-12.679	24.927	1.443	1.00	10.98	BA	C

ATOM	1483	CG	ASP	B	62	-12.048	23.766	2.160	1.00	12.85	BA	C
ATOM	1484	OD1	ASP	B	62	-11.354	23.956	3.229	1.00	16.62	BA	O
ATOM	1485	OD2	ASP	B	62	-12.274	22.663	1.649	1.00	13.63	BA	O
ATOM	1486	C	ASP	B	62	-13.113	27.422	1.520	1.00	11.38	BA	C
ATOM	1487	O	ASP	B	62	-12.240	28.124	0.947	1.00	13.17	BA	O
ATOM	1488	N	GLY	B	63	-14.423	27.751	1.525	1.00	11.62	BA	N
ATOM	1489	CA	GLY	B	63	-14.891	28.933	0.834	1.00	13.29	BA	C
ATOM	1490	C	GLY	B	63	-14.357	30.197	1.478	1.00	14.67	BA	C
ATOM	1491	O	GLY	B	63	-13.929	31.085	0.749	1.00	13.69	BA	O
ATOM	1492	N	MET	B	64	-14.329	30.251	2.818	1.00	14.73	BA	N
ATOM	1493	CA	MET	B	64	-13.737	31.406	3.528	1.00	16.72	BA	C
ATOM	1494	CB	MET	B	64	-13.780	31.201	5.041	1.00	18.24	BA	C
ATOM	1495	CG	MET	B	64	-12.863	32.234	5.743	1.00	18.22	BA	C
ATOM	1496	SD	MET	B	64	-13.040	32.168	7.569	1.00	27.19	BA	S
ATOM	1497	CE	MET	B	64	-14.773	32.215	7.937	1.00	28.58	BA	C
ATOM	1498	C	MET	B	64	-12.325	31.642	3.033	1.00	17.17	BA	C
ATOM	1499	O	MET	B	64	-11.936	32.802	2.650	1.00	17.17	BA	O
ATOM	1500	N	ALA	B	65	-11.537	30.574	2.981	1.00	16.03	BA	N
ATOM	1501	CA	ALA	B	65	-10.139	30.677	2.560	1.00	17.31	BA	C
ATOM	1502	CB	ALA	B	65	-9.490	29.307	2.710	1.00	16.77	BA	C
ATOM	1503	C	ALA	B	65	-10.023	31.191	1.111	1.00	16.05	BA	C
ATOM	1504	O	ALA	B	65	-9.023	31.822	0.743	1.00	17.31	BA	O
ATOM	1505	N	SER	B	66	-10.978	30.932	0.241	1.00	15.57	BA	N
ATOM	1506	CA	SER	B	66	-10.884	31.296	-1.144	1.00	16.13	BA	C
ATOM	1507	CB	SER	B	66	-11.907	30.443	-1.898	1.00	16.36	BA	C
ATOM	1508	OG	SER	B	66	-11.554	29.111	-1.668	1.00	24.38	BA	O
ATOM	1509	C	SER	B	66	-11.066	32.782	-1.469	1.00	16.19	BA	C
ATOM	1510	O	SER	B	66	-10.541	33.299	-2.461	1.00	16.13	BA	O
ATOM	1511	N	GLY	B	67	-11.796	33.493	-0.625	1.00	14.55	BA	N
ATOM	1512	CA	GLY	B	67	-11.912	34.948	-0.753	1.00	13.59	BA	C
ATOM	1513	C	GLY	B	67	-13.027	35.461	-1.579	1.00	14.94	BA	C
ATOM	1514	O	GLY	B	67	-13.651	34.784	-2.364	1.00	13.47	BA	O
ATOM	1515	N	LEU	B	68	-13.169	36.827	-1.421	1.00	16.02	BA	N
ATOM	1516	CA	LEU	B	68	-14.248	37.560	-2.091	1.00	15.39	BA	C
ATOM	1517	CB	LEU	B	68	-14.148	39.083	-1.704	1.00	18.25	BA	C
ATOM	1518	CG	LEU	B	68	-15.012	40.058	-2.447	1.00	20.47	BA	C
ATOM	1519	CD1	LEU	B	68	-16.414	40.069	-1.967	1.00	25.72	BA	C
ATOM	1520	CD2	LEU	B	68	-14.277	41.332	-1.968	1.00	19.64	BA	C
ATOM	1521	C	LEU	B	68	-14.263	37.330	-3.642	1.00	12.87	BA	C
ATOM	1522	O	LEU	B	68	-15.276	37.274	-4.301	1.00	13.59	BA	O
ATOM	1523	N	ASP	B	69	-13.043	37.372	-4.194	1.00	14.84	BA	N
ATOM	1524	CA	ASP	B	69	-12.927	37.347	-5.677	1.00	13.35	BA	C
ATOM	1525	CB	ASP	B	69	-11.504	37.741	-6.163	1.00	11.58	BA	C
ATOM	1526	CG	ASP	B	69	-11.125	39.168	-5.794	1.00	15.72	BA	C
ATOM	1527	OD1	ASP	B	69	-11.993	40.009	-5.556	1.00	14.71	BA	O
ATOM	1528	OD2	ASP	B	69	-9.938	39.406	-5.712	1.00	19.22	BA	O
ATOM	1529	C	ASP	B	69	-13.306	35.997	-6.253	1.00	13.15	BA	C
ATOM	1530	O	ASP	B	69	-13.531	35.924	-7.464	1.00	17.70	BA	O
ATOM	1531	N	LYS	B	70	-13.310	34.961	-5.392	1.00	14.21	BA	N
ATOM	1532	CA	LYS	B	70	-13.809	33.630	-5.794	1.00	14.36	BA	C
ATOM	1533	CB	LYS	B	70	-12.866	32.514	-5.316	1.00	14.08	BA	C
ATOM	1534	CG	LYS	B	70	-11.374	32.637	-5.767	1.00	19.70	BA	C
ATOM	1535	CD	LYS	B	70	-11.252	32.290	-7.152	1.00	22.71	BA	C
ATOM	1536	CE	LYS	B	70	-9.767	32.011	-7.523	1.00	20.00	BA	C
ATOM	1537	NZ	LYS	B	70	-8.740	32.757	-6.755	1.00	28.09	BA	N

ATOM	1538	C	LYS	B	70	-15.170	33.408	-5.182	1.00	14.12	BA	C
ATOM	1539	O	LYS	B	70	-15.578	32.242	-5.077	1.00	13.08	BA	O
ATOM	1540	N	ASP	B	71	-15.875	34.514	-4.821	1.00	14.49	BA	N
ATOM	1541	CA	ASP	B	71	-17.206	34.421	-4.253	1.00	14.41	BA	C
ATOM	1542	CB	ASP	B	71	-18.187	33.923	-5.299	1.00	16.35	BA	C
ATOM	1543	CG	ASP	B	71	-18.696	35.041	-6.250	1.00	14.65	BA	C
ATOM	1544	OD1	ASP	B	71	-18.034	36.105	-6.227	1.00	23.21	BA	O
ATOM	1545	OD2	ASP	B	71	-19.537	34.715	-7.054	1.00	23.60	BA	O
ATOM	1546	C	ASP	B	71	-17.291	33.533	-2.964	1.00	12.69	BA	C
ATOM	1547	O	ASP	B	71	-18.347	32.909	-2.718	1.00	12.21	BA	O
ATOM	1548	N	TYR	B	72	-16.191	33.436	-2.268	1.00	12.15	BA	N
ATOM	1549	CA	TYR	B	72	-16.041	32.631	-1.077	1.00	11.11	BA	C
ATOM	1550	CB	TYR	B	72	-16.849	33.253	0.082	1.00	11.96	BA	C
ATOM	1551	CG	TYR	B	72	-16.154	34.469	0.617	1.00	10.49	BA	C
ATOM	1552	CD1	TYR	B	72	-14.982	34.342	1.286	1.00	11.04	BA	C
ATOM	1553	CE1	TYR	B	72	-14.296	35.496	1.834	1.00	12.38	BA	C
ATOM	1554	CZ	TYR	B	72	-14.872	36.727	1.684	1.00	10.45	BA	C
ATOM	1555	OH	TYR	B	72	-14.225	37.819	2.239	1.00	14.01	BA	O
ATOM	1556	CE2	TYR	B	72	-16.073	36.869	0.991	1.00	13.26	BA	C
ATOM	1557	CD2	TYR	B	72	-16.740	35.730	0.480	1.00	12.85	BA	C
ATOM	1558	C	TYR	B	72	-16.492	31.199	-1.315	1.00	10.60	BA	C
ATOM	1559	O	TYR	B	72	-17.140	30.521	-0.462	1.00	11.32	BA	O
ATOM	1560	N	LEU	B	73	-16.052	30.655	-2.449	1.00	10.48	BA	N
ATOM	1561	CA	LEU	B	73	-16.255	29.216	-2.752	1.00	10.92	BA	C
ATOM	1562	CB	LEU	B	73	-17.319	28.975	-3.848	1.00	12.37	BA	C
ATOM	1563	CG	LEU	B	73	-18.740	29.212	-3.319	1.00	14.12	BA	C
ATOM	1564	CD1	LEU	B	73	-19.583	29.148	-4.587	1.00	17.32	BA	C
ATOM	1565	CD2	LEU	B	73	-19.142	28.112	-2.312	1.00	14.16	BA	C
ATOM	1566	C	LEU	B	73	-14.943	28.669	-3.287	1.00	10.96	BA	C
ATOM	1567	O	LEU	B	73	-14.291	29.292	-4.159	1.00	11.72	BA	O
ATOM	1568	N	LYS	B	74	-14.465	27.562	-2.708	1.00	11.70	BA	N
ATOM	1569	CA	LYS	B	74	-13.274	26.853	-3.242	1.00	11.73	BA	C
ATOM	1570	CB	LYS	B	74	-12.971	25.637	-2.366	1.00	13.27	BA	C
ATOM	1571	CG	LYS	B	74	-11.760	24.702	-2.939	1.00	14.76	BA	C
ATOM	1572	CD	LYS	B	74	-12.334	23.249	-2.637	1.00	22.32	BA	C
ATOM	1573	CE	LYS	B	74	-11.848	22.390	-1.527	1.00	22.93	BA	C
ATOM	1574	NZ	LYS	B	74	-11.111	21.161	-2.054	1.00	27.26	BA	N
ATOM	1575	C	LYS	B	74	-13.529	26.484	-4.718	1.00	11.13	BA	C
ATOM	1576	O	LYS	B	74	-14.529	25.920	-5.090	1.00	11.44	BA	O
ATOM	1577	N	PRO	B	75	-12.558	26.853	-5.581	1.00	10.74	BA	N
ATOM	1578	CA	PRO	B	75	-12.732	26.554	-6.988	1.00	11.83	BA	C
ATOM	1579	CB	PRO	B	75	-11.458	27.147	-7.664	1.00	11.27	BA	C
ATOM	1580	CG	PRO	B	75	-11.198	28.389	-6.759	1.00	13.94	BA	C
ATOM	1581	CD	PRO	B	75	-11.493	27.836	-5.334	1.00	12.92	BA	C
ATOM	1582	C	PRO	B	75	-12.811	25.042	-7.221	1.00	11.52	BA	C
ATOM	1583	O	PRO	B	75	-12.104	24.298	-6.533	1.00	12.23	BA	O
ATOM	1584	N	ASP	B	76	-13.655	24.613	-8.133	1.00	11.55	BA	N
ATOM	1585	CA	ASP	B	76	-13.722	23.175	-8.445	1.00	13.01	BA	C
ATOM	1586	CB	ASP	B	76	-12.420	22.705	-9.105	1.00	13.70	BA	C
ATOM	1587	CG	ASP	B	76	-12.633	21.493	-10.003	1.00	17.64	BA	C
ATOM	1588	OD1	ASP	B	76	-13.755	20.996	-10.152	1.00	18.74	BA	O
ATOM	1589	OD2	ASP	B	76	-11.641	21.028	-10.543	1.00	18.70	BA	O
ATOM	1590	C	ASP	B	76	-14.022	22.327	-7.204	1.00	11.95	BA	C
ATOM	1591	O	ASP	B	76	-13.401	21.246	-6.958	1.00	13.25	BA	O
ATOM	1592	N	ASP	B	77	-15.023	22.786	-6.415	1.00	11.60	BA	N

ATOM	1593	CA	ASP	B	77	-15.426	22.072	-5.207	1.00	11.66	BA	C
ATOM	1594	CB	ASP	B	77	-15.848	23.067	-4.110	1.00	10.05	BA	C
ATOM	1595	CG	ASP	B	77	-16.040	22.418	-2.746	1.00	10.58	BA	C
ATOM	1596	OD1	ASP	B	77	-16.241	21.201	-2.726	1.00	11.04	BA	O
ATOM	1597	OD2	ASP	B	77	-15.952	23.131	-1.743	1.00	10.71	BA	O
ATOM	1598	C	ASP	B	77	-16.604	21.144	-5.595	1.00	11.01	BA	C
ATOM	1599	O	ASP	B	77	-17.749	21.584	-5.706	1.00	11.76	BA	O
ATOM	1600	N	SER	B	78	-16.322	19.852	-5.628	1.00	11.40	BA	N
ATOM	1601	CA	SER	B	78	-17.290	18.846	-6.036	1.00	11.33	BA	C
ATOM	1602	CB	SER	B	78	-16.625	17.459	-6.206	1.00	11.07	BA	C
ATOM	1603	OG	SER	B	78	-16.153	16.931	-4.984	1.00	15.40	BA	O
ATOM	1604	C	SER	B	78	-18.475	18.707	-5.052	1.00	10.63	BA	C
ATOM	1605	O	SER	B	78	-19.571	18.206	-5.409	1.00	12.36	BA	O
ATOM	1606	N	ARG	B	79	-18.316	19.173	-3.820	1.00	10.54	BA	N
ATOM	1607	CA	ARG	B	79	-19.339	19.090	-2.750	1.00	11.65	BA	C
ATOM	1608	CB	ARG	B	79	-18.708	19.379	-1.352	1.00	10.95	BA	C
ATOM	1609	CG	ARG	B	79	-17.611	18.352	-0.972	1.00	11.06	BA	C
ATOM	1610	CD	ARG	B	79	-16.859	18.802	0.268	1.00	7.67	BA	C
ATOM	1611	NE	ARG	B	79	-16.169	20.068	-0.041	1.00	9.80	BA	N
ATOM	1612	CZ	ARG	B	79	-15.220	20.637	0.713	1.00	11.93	BA	C
ATOM	1613	NH1	ARG	B	79	-14.903	19.985	1.844	1.00	13.15	BA	N
ATOM	1614	NH2	ARG	B	79	-14.636	21.767	0.349	1.00	10.92	BA	N
ATOM	1615	C	ARG	B	79	-20.474	20.079	-2.982	1.00	9.39	BA	C
ATOM	1616	O	ARG	B	79	-21.533	19.928	-2.386	1.00	11.48	BA	O
ATOM	1617	N	VAL	B	80	-20.249	21.119	-3.804	1.00	10.56	BA	N
ATOM	1618	CA	VAL	B	80	-21.265	22.114	-4.054	1.00	10.92	BA	C
ATOM	1619	CB	VAL	B	80	-20.612	23.444	-4.445	1.00	11.14	BA	C
ATOM	1620	CG1	VAL	B	80	-21.793	24.491	-4.808	1.00	12.79	BA	C
ATOM	1621	CG2	VAL	B	80	-19.715	23.951	-3.278	1.00	12.23	BA	C
ATOM	1622	C	VAL	B	80	-22.246	21.579	-5.069	1.00	10.18	BA	C
ATOM	1623	O	VAL	B	80	-21.886	21.397	-6.239	1.00	12.37	BA	O
ATOM	1624	N	ILE	B	81	-23.508	21.384	-4.677	1.00	9.68	BA	N
ATOM	1625	CA	ILE	B	81	-24.499	20.816	-5.641	1.00	9.98	BA	C
ATOM	1626	CB	ILE	B	81	-25.624	20.247	-4.825	1.00	12.96	BA	C
ATOM	1627	CG1	ILE	B	81	-25.129	19.035	-3.967	1.00	12.15	BA	C
ATOM	1628	CD1	ILE	B	81	-26.144	18.545	-2.927	1.00	13.75	BA	C
ATOM	1629	CG2	ILE	B	81	-26.806	19.821	-5.761	1.00	14.42	BA	C
ATOM	1630	C	ILE	B	81	-24.959	21.945	-6.569	1.00	11.49	BA	C
ATOM	1631	O	ILE	B	81	-25.197	21.708	-7.797	1.00	12.90	BA	O
ATOM	1632	N	ALA	B	82	-25.087	23.159	-6.043	1.00	10.53	BA	N
ATOM	1633	CA	ALA	B	82	-25.565	24.312	-6.835	1.00	11.32	BA	C
ATOM	1634	CB	ALA	B	82	-27.121	24.266	-7.013	1.00	12.66	BA	C
ATOM	1635	C	ALA	B	82	-25.238	25.532	-5.992	1.00	11.11	BA	C
ATOM	1636	O	ALA	B	82	-25.137	25.471	-4.765	1.00	11.04	BA	O
ATOM	1637	N	HIS	B	83	-25.100	26.663	-6.675	1.00	11.25	BA	N
ATOM	1638	CA	HIS	B	83	-24.854	27.943	-5.941	1.00	9.67	BA	C
ATOM	1639	CB	HIS	B	83	-23.380	28.029	-5.501	1.00	12.01	BA	C
ATOM	1640	CG	HIS	B	83	-22.446	28.066	-6.673	1.00	11.58	BA	C
ATOM	1641	ND1	HIS	B	83	-21.889	29.224	-7.223	1.00	14.29	BA	N
ATOM	1642	CE1	HIS	B	83	-21.188	28.863	-8.305	1.00	15.17	BA	C
ATOM	1643	NE2	HIS	B	83	-21.198	27.544	-8.399	1.00	16.15	BA	N
ATOM	1644	CD2	HIS	B	83	-22.033	27.038	-7.422	1.00	12.71	BA	C
ATOM	1645	C	HIS	B	83	-25.169	29.075	-6.826	1.00	11.59	BA	C
ATOM	1646	O	HIS	B	83	-25.024	29.040	-8.058	1.00	11.61	BA	O
ATOM	1647	N	THR	B	84	-25.501	30.189	-6.207	1.00	12.04	BA	N

ATOM	1648	CA	THR	B	84	-25.543	31.523	-6.890	1.00	10.82	BA	C
ATOM	1649	CB	THR	B	84	-26.550	32.472	-6.236	1.00	11.72	BA	C
ATOM	1650	OG1	THR	B	84	-26.032	32.784	-4.916	1.00	12.25	BA	O
ATOM	1651	CG2	THR	B	84	-27.926	31.897	-6.132	1.00	10.63	BA	C
ATOM	1652	C	THR	B	84	-24.154	32.117	-6.799	1.00	10.75	BA	C
ATOM	1653	O	THR	B	84	-23.223	31.598	-6.169	1.00	13.03	BA	O
ATOM	1654	N	LYS	B	85	-23.971	33.264	-7.453	1.00	12.88	BA	N
ATOM	1655	CA	LYS	B	85	-22.794	34.128	-7.196	1.00	12.76	BA	C
ATOM	1656	CB	LYS	B	85	-22.695	35.270	-8.271	1.00	15.51	BA	C
ATOM	1657	CG	LYS	B	85	-22.663	34.693	-9.673	1.00	18.14	BA	C
ATOM	1658	CD	LYS	B	85	-22.295	35.803	-10.662	1.00	23.07	BA	C
ATOM	1659	CE	LYS	B	85	-22.333	35.366	-12.156	1.00	28.38	BA	C
ATOM	1660	NZ	LYS	B	85	-22.047	33.908	-12.359	1.00	27.62	BA	N
ATOM	1661	C	LYS	B	85	-22.952	34.803	-5.819	1.00	13.69	BA	C
ATOM	1662	O	LYS	B	85	-24.083	34.856	-5.242	1.00	12.89	BA	O
ATOM	1663	N	LEU	B	86	-21.858	35.443	-5.360	1.00	11.58	BA	N
ATOM	1664	CA	LEU	B	86	-21.889	36.337	-4.189	1.00	12.13	BA	C
ATOM	1665	CB	LEU	B	86	-20.500	36.431	-3.591	1.00	13.32	BA	C
ATOM	1666	CG	LEU	B	86	-20.474	37.092	-2.188	1.00	9.89	BA	C
ATOM	1667	CD1	LEU	B	86	-20.787	36.116	-1.091	1.00	12.73	BA	C
ATOM	1668	CD2	LEU	B	86	-19.092	37.677	-1.989	1.00	11.72	BA	C
ATOM	1669	C	LEU	B	86	-22.407	37.733	-4.575	1.00	11.07	BA	C
ATOM	1670	O	LEU	B	86	-21.941	38.336	-5.537	1.00	14.73	BA	O
ATOM	1671	N	ILE	B	87	-23.421	38.189	-3.864	1.00	11.93	BA	N
ATOM	1672	CA	ILE	B	87	-24.018	39.529	-4.214	1.00	11.17	BA	C
ATOM	1673	CB	ILE	B	87	-25.493	39.411	-4.785	1.00	11.67	BA	C
ATOM	1674	CG1	ILE	B	87	-26.484	38.819	-3.755	1.00	13.44	BA	C
ATOM	1675	CD1	ILE	B	87	-27.896	38.802	-4.167	1.00	13.35	BA	C
ATOM	1676	CG2	ILE	B	87	-25.404	38.606	-6.115	1.00	12.76	BA	C
ATOM	1677	C	ILE	B	87	-24.010	40.449	-3.042	1.00	11.73	BA	C
ATOM	1678	O	ILE	B	87	-24.017	40.008	-1.883	1.00	12.77	BA	O
ATOM	1679	N	GLY	B	88	-23.921	41.755	-3.339	1.00	12.33	BA	N
ATOM	1680	CA	GLY	B	88	-24.090	42.779	-2.309	1.00	13.29	BA	C
ATOM	1681	C	GLY	B	88	-25.456	43.414	-2.430	1.00	12.48	BA	C
ATOM	1682	O	GLY	B	88	-26.294	43.020	-3.298	1.00	14.71	BA	O
ATOM	1683	N	SER	B	89	-25.667	44.472	-1.622	1.00	13.83	BA	N
ATOM	1684	CA	SER	B	89	-27.021	45.080	-1.690	1.00	15.80	BA	C
ATOM	1685	CB	SER	B	89	-27.270	46.123	-0.599	1.00	15.40	BA	C
ATOM	1686	OG	SER	B	89	-26.464	47.251	-0.886	1.00	19.64	BA	O
ATOM	1687	C	SER	B	89	-27.360	45.672	-3.097	1.00	14.95	BA	C
ATOM	1688	O	SER	B	89	-26.499	46.125	-3.898	1.00	15.24	BA	O
ATOM	1689	N	GLY	B	90	-28.643	45.545	-3.357	1.00	15.18	BA	N
ATOM	1690	CA	GLY	B	90	-29.226	45.974	-4.661	1.00	14.24	BA	C
ATOM	1691	C	GLY	B	90	-28.890	45.040	-5.817	1.00	17.58	BA	C
ATOM	1692	O	GLY	B	90	-29.278	45.308	-6.958	1.00	20.84	BA	O
ATOM	1693	N	GLU	B	91	-28.193	43.929	-5.577	1.00	16.81	BA	N
ATOM	1694	CA	GLU	B	91	-27.932	42.940	-6.602	1.00	16.20	BA	C
ATOM	1695	CB	GLU	B	91	-26.471	42.482	-6.597	1.00	17.35	BA	C
ATOM	1696	CG	GLU	B	91	-25.502	43.630	-6.831	1.00	18.66	BA	C
ATOM	1697	CD	GLU	B	91	-24.049	43.164	-6.962	1.00	28.13	BA	C
ATOM	1698	OE1	GLU	B	91	-23.444	42.633	-5.981	1.00	20.77	BA	O
ATOM	1699	OE2	GLU	B	91	-23.480	43.358	-8.069	1.00	29.56	BA	O
ATOM	1700	C	GLU	B	91	-28.865	41.758	-6.498	1.00	16.96	BA	C
ATOM	1701	O	GLU	B	91	-29.540	41.542	-5.484	1.00	16.36	BA	O
ATOM	1702	N	LYS	B	92	-28.934	41.005	-7.581	1.00	15.96	BA	N

ATOM	1703	CA	LYS	B	92	-29.715	39.760	-7.631	1.00	18.04	BA	C
ATOM	1704	CB	LYS	B	92	-31.064	39.931	-8.352	1.00	18.30	BA	C
ATOM	1705	CG	LYS	B	92	-32.123	40.911	-7.702	1.00	21.94	BA	C
ATOM	1706	CD	LYS	B	92	-33.482	41.174	-8.467	1.00	29.25	BA	C
ATOM	1707	CE	LYS	B	92	-34.724	40.391	-7.893	1.00	33.95	BA	C
ATOM	1708	NZ	LYS	B	92	-35.660	40.970	-6.746	1.00	30.51	BA	N
ATOM	1709	C	LYS	B	92	-28.874	38.734	-8.410	1.00	16.78	BA	C
ATOM	1710	O	LYS	B	92	-27.929	39.039	-9.220	1.00	15.88	BA	O
ATOM	1711	N	ASP	B	93	-29.231	37.469	-8.173	1.00	14.82	BA	N
ATOM	1712	CA	ASP	B	93	-28.676	36.458	-8.961	1.00	13.44	BA	C
ATOM	1713	CB	ASP	B	93	-27.256	36.091	-8.521	1.00	12.76	BA	C
ATOM	1714	CG	ASP	B	93	-26.543	35.239	-9.501	1.00	12.59	BA	C
ATOM	1715	OD1	ASP	B	93	-26.461	35.638	-10.708	1.00	13.80	BA	O
ATOM	1716	OD2	ASP	B	93	-26.059	34.145	-9.151	1.00	14.85	BA	O
ATOM	1717	C	ASP	B	93	-29.548	35.231	-8.851	1.00	13.10	BA	C
ATOM	1718	O	ASP	B	93	-29.998	34.869	-7.781	1.00	13.30	BA	O
ATOM	1719	N	SER	B	94	-29.717	34.520	-9.956	1.00	12.18	BA	N
ATOM	1720	CA	SER	B	94	-30.491	33.278	-9.991	1.00	12.54	BA	C
ATOM	1721	CB	SER	B	94	-31.650	33.367	-11.025	1.00	13.98	BA	C
ATOM	1722	OG	SER	B	94	-32.533	34.394	-10.629	1.00	13.38	BA	O
ATOM	1723	C	SER	B	94	-29.671	32.115	-10.364	1.00	11.49	BA	C
ATOM	1724	O	SER	B	94	-28.774	32.228	-11.202	1.00	11.76	BA	O
ATOM	1725	N	VAL	B	95	-30.004	30.932	-9.827	1.00	11.28	BA	N
ATOM	1726	CA	VAL	B	95	-29.327	29.715	-10.265	1.00	11.76	BA	C
ATOM	1727	CB	VAL	B	95	-28.295	29.160	-9.211	1.00	13.91	BA	C
ATOM	1728	CG1	VAL	B	95	-28.916	28.796	-7.920	1.00	15.24	BA	C
ATOM	1729	CG2	VAL	B	95	-27.525	27.939	-9.787	1.00	17.87	BA	C
ATOM	1730	C	VAL	B	95	-30.408	28.676	-10.560	1.00	11.21	BA	C
ATOM	1731	O	VAL	B	95	-31.367	28.506	-9.775	1.00	11.16	BA	O
ATOM	1732	N	THR	B	96	-30.254	27.945	-11.630	1.00	9.78	BA	N
ATOM	1733	CA	THR	B	96	-31.129	26.828	-11.941	1.00	8.96	BA	C
ATOM	1734	CB	THR	B	96	-31.649	26.984	-13.372	1.00	8.86	BA	C
ATOM	1735	OG1	THR	B	96	-32.547	28.130	-13.290	1.00	11.01	BA	O
ATOM	1736	CG2	THR	B	96	-32.477	25.698	-13.795	1.00	9.82	BA	C
ATOM	1737	C	THR	B	96	-30.237	25.523	-11.857	1.00	10.66	BA	C
ATOM	1738	O	THR	B	96	-29.125	25.514	-12.436	1.00	10.96	BA	O
ATOM	1739	N	PHE	B	97	-30.764	24.465	-11.282	1.00	10.62	BA	N
ATOM	1740	CA	PHE	B	97	-30.058	23.208	-11.256	1.00	10.49	BA	C
ATOM	1741	CB	PHE	B	97	-29.334	23.011	-9.852	1.00	10.62	BA	C
ATOM	1742	CG	PHE	B	97	-30.211	22.935	-8.731	1.00	11.15	BA	C
ATOM	1743	CD1	PHE	B	97	-30.554	24.069	-7.990	1.00	11.32	BA	C
ATOM	1744	CE1	PHE	B	97	-31.287	24.010	-6.856	1.00	14.06	BA	C
ATOM	1745	CZ	PHE	B	97	-31.791	22.793	-6.418	1.00	17.06	BA	C
ATOM	1746	CE2	PHE	B	97	-31.496	21.651	-7.120	1.00	18.33	BA	C
ATOM	1747	CD2	PHE	B	97	-30.612	21.693	-8.242	1.00	12.79	BA	C
ATOM	1748	C	PHE	B	97	-30.997	22.053	-11.594	1.00	11.88	BA	C
ATOM	1749	O	PHE	B	97	-32.178	22.133	-11.412	1.00	11.95	BA	O
ATOM	1750	N	ASP	B	98	-30.387	20.937	-11.962	1.00	11.16	BA	N
ATOM	1751	CA	ASP	B	98	-31.098	19.771	-12.311	1.00	10.66	BA	C
ATOM	1752	CB	ASP	B	98	-30.206	18.820	-13.059	1.00	13.41	BA	C
ATOM	1753	CG	ASP	B	98	-29.961	19.231	-14.432	1.00	16.39	BA	C
ATOM	1754	OD1	ASP	B	98	-30.731	20.034	-15.009	1.00	18.00	BA	O
ATOM	1755	OD2	ASP	B	98	-28.961	18.746	-15.015	1.00	22.05	BA	O
ATOM	1756	C	ASP	B	98	-31.605	19.035	-11.037	1.00	11.23	BA	C
ATOM	1757	O	ASP	B	98	-30.857	18.731	-10.104	1.00	12.82	BA	O

ATOM	1758	N	VAL B 99	-32.889	18.725	-11.007	1.00	11.52	BA	N
ATOM	1759	CA	VAL B 99	-33.499	18.044	-9.868	1.00	12.32	BA	C
ATOM	1760	CB	VAL B 99	-35.072	18.163	-9.916	1.00	12.14	BA	C
ATOM	1761	CG1	VAL B 99	-35.763	17.312	-8.842	1.00	14.65	BA	C
ATOM	1762	CG2	VAL B 99	-35.536	19.622	-9.684	1.00	14.40	BA	C
ATOM	1763	C	VAL B 99	-32.925	16.611	-9.704	1.00	11.38	BA	C
ATOM	1764	O	VAL B 99	-32.910	16.088	-8.560	1.00	11.97	BA	O
ATOM	1765	N	SER B 100	-32.464	16.027	-10.823	1.00	11.82	BA	N
ATOM	1766	CA	SER B 100	-31.810	14.736	-10.759	1.00	13.27	BA	C
ATOM	1767	CB	SER B 100	-31.453	14.276	-12.197	1.00	14.14	BA	C
ATOM	1768	OG	SER B 100	-30.507	15.151	-12.789	1.00	18.24	BA	O
ATOM	1769	C	SER B 100	-30.548	14.763	-9.864	1.00	13.21	BA	C
ATOM	1770	O	SER B 100	-30.056	13.671	-9.525	1.00	15.64	BA	O
ATOM	1771	N	LYS B 101	-30.009	15.921	-9.476	1.00	12.64	BA	N
ATOM	1772	CA	LYS B 101	-28.890	15.938	-8.551	1.00	14.56	BA	C
ATOM	1773	CB	LYS B 101	-28.266	17.368	-8.582	1.00	13.97	BA	C
ATOM	1774	CG	LYS B 101	-27.566	17.601	-9.941	1.00	19.71	BA	C
ATOM	1775	CD	LYS B 101	-26.496	18.629	-9.915	1.00	30.06	BA	C
ATOM	1776	CE	LYS B 101	-25.594	18.439	-11.170	1.00	29.54	BA	C
ATOM	1777	NZ	LYS B 101	-24.486	19.451	-11.203	1.00	37.48	BA	N
ATOM	1778	C	LYS B 101	-29.299	15.608	-7.138	1.00	13.92	BA	C
ATOM	1779	O	LYS B 101	-28.417	15.398	-6.291	1.00	17.93	BA	O
ATOM	1780	N	LEU B 102	-30.603	15.587	-6.848	1.00	12.47	BA	N
ATOM	1781	CA	LEU B 102	-31.135	15.414	-5.460	1.00	12.23	BA	C
ATOM	1782	CB	LEU B 102	-32.235	16.432	-5.100	1.00	13.51	BA	C
ATOM	1783	CG	LEU B 102	-31.784	17.894	-5.327	1.00	11.38	BA	C
ATOM	1784	CD1	LEU B 102	-32.945	18.804	-4.983	1.00	17.77	BA	C
ATOM	1785	CD2	LEU B 102	-30.522	18.245	-4.492	1.00	14.77	BA	C
ATOM	1786	C	LEU B 102	-31.732	14.017	-5.322	1.00	12.29	BA	C
ATOM	1787	O	LEU B 102	-32.326	13.460	-6.292	1.00	13.00	BA	O
ATOM	1788	N	LYS B 103	-31.602	13.458	-4.123	1.00	13.06	BA	N
ATOM	1789	CA	LYS B 103	-32.100	12.108	-3.837	1.00	13.97	BA	C
ATOM	1790	CB	LYS B 103	-31.014	11.273	-3.121	1.00	14.49	BA	C
ATOM	1791	CG	LYS B 103	-31.216	9.756	-3.098	1.00	22.49	BA	C
ATOM	1792	CD	LYS B 103	-29.909	9.087	-2.554	1.00	27.58	BA	C
ATOM	1793	CE	LYS B 103	-28.599	9.893	-2.914	1.00	33.20	BA	C
ATOM	1794	NZ	LYS B 103	-27.312	9.416	-2.244	1.00	31.85	BA	N
ATOM	1795	C	LYS B 103	-33.269	12.259	-2.914	1.00	12.61	BA	C
ATOM	1796	O	LYS B 103	-33.259	13.001	-1.876	1.00	12.73	BA	O
ATOM	1797	N	GLU B 104	-34.315	11.546	-3.217	1.00	13.28	BA	N
ATOM	1798	CA	GLU B 104	-35.435	11.512	-2.293	1.00	14.02	BA	C
ATOM	1799	CB	GLU B 104	-36.557	10.634	-2.883	1.00	13.96	BA	C
ATOM	1800	CG	GLU B 104	-37.329	11.345	-4.032	1.00	18.78	BA	C
ATOM	1801	CD	GLU B 104	-38.564	10.524	-4.403	1.00	20.37	BA	C
ATOM	1802	OE1	GLU B 104	-39.521	10.620	-3.600	1.00	22.40	BA	O
ATOM	1803	OE2	GLU B 104	-38.565	9.819	-5.463	1.00	19.27	BA	O
ATOM	1804	C	GLU B 104	-35.034	11.008	-0.903	1.00	12.89	BA	C
ATOM	1805	O	GLU B 104	-34.211	10.106	-0.738	1.00	15.32	BA	O
ATOM	1806	N	GLY B 105	-35.663	11.601	0.119	1.00	15.17	BA	N
ATOM	1807	CA	GLY B 105	-35.456	11.135	1.476	1.00	16.57	BA	C
ATOM	1808	C	GLY B 105	-34.273	11.836	2.128	1.00	17.64	BA	C
ATOM	1809	O	GLY B 105	-33.999	11.595	3.320	1.00	21.46	BA	O
ATOM	1810	N	GLU B 106	-33.511	12.617	1.360	1.00	15.28	BA	N
ATOM	1811	CA	GLU B 106	-32.349	13.292	1.958	1.00	15.18	BA	C
ATOM	1812	CB	GLU B 106	-31.097	13.126	1.062	1.00	16.31	BA	C

ATOM	1813	CG	GLU	B	106	-29.863	13.832	1.615	1.00	21.72	BA	C
ATOM	1814	CD	GLU	B	106	-28.507	13.208	1.220	1.00	29.28	BA	C
ATOM	1815	OE1	GLU	B	106	-28.487	12.298	0.337	1.00	31.34	BA	O
ATOM	1816	OE2	GLU	B	106	-27.439	13.619	1.840	1.00	28.39	BA	O
ATOM	1817	C	GLU	B	106	-32.653	14.732	2.226	1.00	13.06	BA	C
ATOM	1818	O	GLU	B	106	-33.306	15.406	1.426	1.00	16.27	BA	O
ATOM	1819	N	GLN	B	107	-32.243	15.231	3.388	1.00	12.66	BA	N
ATOM	1820	CA	GLN	B	107	-32.354	16.636	3.713	1.00	14.99	BA	C
ATOM	1821	CB	GLN	B	107	-32.758	16.763	5.203	1.00	14.81	BA	C
ATOM	1822	CG	GLN	B	107	-32.972	18.256	5.586	1.00	18.13	BA	C
ATOM	1823	CD	GLN	B	107	-33.198	18.341	7.065	1.00	16.70	BA	C
ATOM	1824	OE1	GLN	B	107	-32.206	18.290	7.872	1.00	17.43	BA	O
ATOM	1825	NE2	GLN	B	107	-34.449	18.495	7.472	1.00	15.55	BA	N
ATOM	1826	C	GLN	B	107	-31.004	17.315	3.526	1.00	16.59	BA	C
ATOM	1827	O	GLN	B	107	-30.031	16.996	4.275	1.00	17.59	BA	O
ATOM	1828	N	TYR	B	108	-30.916	18.230	2.565	1.00	12.37	BA	N
ATOM	1829	CA	TYR	B	108	-29.665	18.967	2.255	1.00	11.34	BA	C
ATOM	1830	CB	TYR	B	108	-29.775	19.332	0.747	1.00	10.13	BA	C
ATOM	1831	CG	TYR	B	108	-29.743	18.070	-0.125	1.00	9.63	BA	C
ATOM	1832	CD1	TYR	B	108	-28.523	17.522	-0.505	1.00	10.50	BA	C
ATOM	1833	CE1	TYR	B	108	-28.492	16.337	-1.257	1.00	10.16	BA	C
ATOM	1834	CZ	TYR	B	108	-29.686	15.727	-1.591	1.00	10.38	BA	C
ATOM	1835	OH	TYR	B	108	-29.639	14.515	-2.336	1.00	11.96	BA	O
ATOM	1836	CE2	TYR	B	108	-30.896	16.295	-1.249	1.00	10.31	BA	C
ATOM	1837	CD2	TYR	B	108	-30.952	17.405	-0.489	1.00	10.66	BA	C
ATOM	1838	C	TYR	B	108	-29.679	20.252	3.060	1.00	8.57	BA	C
ATOM	1839	O	TYR	B	108	-30.692	20.616	3.645	1.00	10.44	BA	O
ATOM	1840	N	MET	B	109	-28.544	20.960	3.051	1.00	9.14	BA	N
ATOM	1841	CA	MET	B	109	-28.418	22.283	3.712	1.00	9.60	BA	C
ATOM	1842	CB	MET	B	109	-27.288	22.290	4.741	1.00	9.33	BA	C
ATOM	1843	CG	MET	B	109	-27.512	21.341	5.916	1.00	10.36	BA	C
ATOM	1844	SD	MET	B	109	-28.952	21.682	6.941	1.00	12.16	BA	S
ATOM	1845	CE	MET	B	109	-28.475	23.306	7.658	1.00	14.69	BA	C
ATOM	1846	C	MET	B	109	-28.114	23.331	2.627	1.00	9.36	BA	C
ATOM	1847	O	MET	B	109	-27.338	23.085	1.682	1.00	9.96	BA	O
ATOM	1848	N	PHE	B	110	-28.797	24.488	2.810	1.00	9.81	BA	N
ATOM	1849	CA	PHE	B	110	-28.390	25.638	2.056	1.00	9.02	BA	C
ATOM	1850	CB	PHE	B	110	-29.521	26.230	1.186	1.00	10.17	BA	C
ATOM	1851	CG	PHE	B	110	-30.660	26.880	1.953	1.00	10.65	BA	C
ATOM	1852	CD1	PHE	B	110	-30.573	28.241	2.289	1.00	10.43	BA	C
ATOM	1853	CE1	PHE	B	110	-31.629	28.846	2.995	1.00	10.97	BA	C
ATOM	1854	CZ	PHE	B	110	-32.768	28.115	3.323	1.00	12.21	BA	C
ATOM	1855	CE2	PHE	B	110	-32.832	26.781	2.932	1.00	10.23	BA	C
ATOM	1856	CD2	PHE	B	110	-31.787	26.162	2.246	1.00	9.99	BA	C
ATOM	1857	C	PHE	B	110	-27.844	26.674	2.990	1.00	8.72	BA	C
ATOM	1858	O	PHE	B	110	-28.210	26.740	4.185	1.00	10.26	BA	O
ATOM	1859	N	PHE	B	111	-26.934	27.513	2.488	1.00	9.36	BA	N
ATOM	1860	CA	PHE	B	111	-26.264	28.491	3.430	1.00	9.30	BA	C
ATOM	1861	CB	PHE	B	111	-25.235	27.741	4.345	1.00	9.78	BA	C
ATOM	1862	CG	PHE	B	111	-24.440	26.694	3.559	1.00	9.74	BA	C
ATOM	1863	CD1	PHE	B	111	-24.935	25.413	3.382	1.00	10.10	BA	C
ATOM	1864	CE1	PHE	B	111	-24.211	24.497	2.641	1.00	10.53	BA	C
ATOM	1865	CZ	PHE	B	111	-22.950	24.835	2.095	1.00	9.14	BA	C
ATOM	1866	CE2	PHE	B	111	-22.488	26.091	2.280	1.00	9.38	BA	C
ATOM	1867	CD2	PHE	B	111	-23.212	27.018	3.000	1.00	10.30	BA	C

ATOM	1868	C	PHE B 111	-25.507	29.512	2.585	1.00	10.37	BA	C
ATOM	1869	O	PHE B 111	-25.276	29.349	1.359	1.00	9.61	BA	O
ATOM	1870	N	CYS B 112	-25.109	30.597	3.245	1.00	9.60	BA	N
ATOM	1871	CA	CYS B 112	-24.164	31.624	2.702	1.00	9.61	BA	C
ATOM	1872	CB	CYS B 112	-24.620	32.950	3.239	1.00	10.65	BA	C
ATOM	1873	SG	CYS B 112	-23.497	34.314	2.684	1.00	9.61	BA	S
ATOM	1874	C	CYS B 112	-22.766	31.328	3.231	1.00	9.61	BA	C
ATOM	1875	O	CYS B 112	-22.584	30.963	4.383	1.00	9.92	BA	O
ATOM	1876	N	THR B 113	-21.779	31.407	2.310	1.00	9.68	BA	N
ATOM	1877	CA	THR B 113	-20.402	31.136	2.639	1.00	9.83	BA	C
ATOM	1878	CB	THR B 113	-19.704	30.276	1.626	1.00	9.16	BA	C
ATOM	1879	OG1	THR B 113	-19.593	31.004	0.382	1.00	10.55	BA	O
ATOM	1880	CG2	THR B 113	-20.342	28.898	1.472	1.00	9.99	BA	C
ATOM	1881	C	THR B 113	-19.569	32.417	2.918	1.00	9.45	BA	C
ATOM	1882	O	THR B 113	-18.362	32.290	3.177	1.00	10.54	BA	O
ATOM	1883	N	PHE B 114	-20.205	33.601	2.991	1.00	9.37	BA	N
ATOM	1884	CA	PHE B 114	-19.419	34.774	3.405	1.00	8.70	BA	C
ATOM	1885	CB	PHE B 114	-20.264	36.033	3.310	1.00	10.61	BA	C
ATOM	1886	CG	PHE B 114	-19.517	37.311	3.560	1.00	9.40	BA	C
ATOM	1887	CD1	PHE B 114	-19.245	37.717	4.878	1.00	11.56	BA	C
ATOM	1888	CE1	PHE B 114	-18.444	38.873	5.190	1.00	12.82	BA	C
ATOM	1889	CZ	PHE B 114	-18.068	39.655	4.051	1.00	13.28	BA	C
ATOM	1890	CE2	PHE B 114	-18.285	39.251	2.755	1.00	11.31	BA	C
ATOM	1891	CD2	PHE B 114	-19.043	38.081	2.479	1.00	12.28	BA	C
ATOM	1892	C	PHE B 114	-18.939	34.541	4.844	1.00	9.96	BA	C
ATOM	1893	O	PHE B 114	-19.696	33.994	5.627	1.00	9.92	BA	O
ATOM	1894	N	PRO B 115	-17.673	34.929	5.220	1.00	9.60	BA	N
ATOM	1895	CA	PRO B 115	-17.140	34.500	6.522	1.00	9.64	BA	C
ATOM	1896	CB	PRO B 115	-15.803	35.300	6.614	1.00	12.40	BA	C
ATOM	1897	CG	PRO B 115	-15.416	35.337	5.132	1.00	13.66	BA	C
ATOM	1898	CD	PRO B 115	-16.628	35.527	4.340	1.00	11.07	BA	C
ATOM	1899	C	PRO B 115	-18.092	34.904	7.669	1.00	8.91	BA	C
ATOM	1900	O	PRO B 115	-18.495	36.058	7.875	1.00	10.09	BA	O
ATOM	1901	N	GLY B 116	-18.459	33.893	8.421	1.00	9.19	BA	N
ATOM	1902	CA	GLY B 116	-19.312	34.036	9.590	1.00	8.76	BA	C
ATOM	1903	C	GLY B 116	-20.778	33.956	9.352	1.00	8.43	BA	C
ATOM	1904	O	GLY B 116	-21.544	33.702	10.307	1.00	11.75	BA	O
ATOM	1905	N	HIS B 117	-21.236	34.073	8.105	1.00	8.21	BA	N
ATOM	1906	CA	HIS B 117	-22.648	34.126	7.847	1.00	9.42	BA	C
ATOM	1907	CB	HIS B 117	-22.935	34.687	6.445	1.00	9.26	BA	C
ATOM	1908	CG	HIS B 117	-22.644	36.149	6.355	1.00	7.82	BA	C
ATOM	1909	ND1	HIS B 117	-23.095	36.901	5.301	1.00	9.42	BA	N
ATOM	1910	CE1	HIS B 117	-22.621	38.134	5.433	1.00	10.04	BA	C
ATOM	1911	NE2	HIS B 117	-22.014	38.250	6.608	1.00	9.44	BA	N
ATOM	1912	CD2	HIS B 117	-21.977	36.981	7.182	1.00	9.18	BA	C
ATOM	1913	C	HIS B 117	-23.399	32.802	8.022	1.00	9.08	BA	C
ATOM	1914	O	HIS B 117	-24.604	32.847	8.302	1.00	10.45	BA	O
ATOM	1915	N	SER B 118	-22.707	31.687	7.923	1.00	9.02	BA	N
ATOM	1916	CA	SER B 118	-23.454	30.419	8.001	1.00	10.68	BA	C
ATOM	1917	CB	SER B 118	-22.583	29.243	7.491	1.00	12.51	BA	C
ATOM	1918	OG	SER B 118	-21.552	29.051	8.362	1.00	15.43	BA	O
ATOM	1919	C	SER B 118	-23.936	30.099	9.408	1.00	11.02	BA	C
ATOM	1920	O	SER B 118	-24.815	29.246	9.543	1.00	10.76	BA	O
ATOM	1921	N	ALA B 119	-23.495	30.831	10.413	1.00	9.63	BA	N
ATOM	1922	CA	ALA B 119	-24.030	30.616	11.771	1.00	10.12	BA	C

ATOM	1923	CB	ALA	B	119	-23.323	31.544	12.822	1.00	12.45	BA	C
ATOM	1924	C	ALA	B	119	-25.554	30.865	11.757	1.00	10.11	BA	C
ATOM	1925	O	ALA	B	119	-26.332	30.141	12.398	1.00	11.33	BA	O
ATOM	1926	N	LEU	B	120	-26.009	31.890	11.069	1.00	9.15	BA	N
ATOM	1927	CA	LEU	B	120	-27.428	32.247	11.009	1.00	10.97	BA	C
ATOM	1928	CB	LEU	B	120	-27.578	33.748	11.363	1.00	10.75	BA	C
ATOM	1929	CG	LEU	B	120	-27.154	34.126	12.783	1.00	10.29	BA	C
ATOM	1930	CD1	LEU	B	120	-27.469	35.596	13.018	1.00	12.17	BA	C
ATOM	1931	CD2	LEU	B	120	-27.911	33.226	13.822	1.00	13.38	BA	C
ATOM	1932	C	LEU	B	120	-28.038	32.051	9.654	1.00	10.29	BA	C
ATOM	1933	O	LEU	B	120	-29.272	31.858	9.571	1.00	10.12	BA	O
ATOM	1934	N	MET	B	121	-27.261	32.074	8.594	1.00	9.32	BA	N
ATOM	1935	CA	MET	B	121	-27.785	32.064	7.216	1.00	9.59	BA	C
ATOM	1936	CB	MET	B	121	-27.084	33.074	6.291	1.00	9.54	BA	C
ATOM	1937	CG	MET	B	121	-27.164	34.501	6.862	1.00	8.98	BA	C
ATOM	1938	SD	MET	B	121	-26.591	35.743	5.699	1.00	10.54	BA	S
ATOM	1939	CE	MET	B	121	-28.016	35.781	4.545	1.00	11.28	BA	C
ATOM	1940	C	MET	B	121	-27.663	30.660	6.668	1.00	8.62	BA	C
ATOM	1941	O	MET	B	121	-26.754	30.319	5.974	1.00	9.80	BA	O
ATOM	1942	N	LYS	B	122	-28.620	29.821	7.147	1.00	9.56	BA	N
ATOM	1943	CA	LYS	B	122	-28.626	28.413	6.732	1.00	9.61	BA	C
ATOM	1944	CB	LYS	B	122	-27.681	27.541	7.555	1.00	11.26	BA	C
ATOM	1945	CG	LYS	B	122	-28.098	27.113	8.951	1.00	10.58	BA	C
ATOM	1946	CD	LYS	B	122	-28.297	28.276	9.918	1.00	12.38	BA	C
ATOM	1947	CE	LYS	B	122	-28.657	27.664	11.289	1.00	13.34	BA	C
ATOM	1948	NZ	LYS	B	122	-28.841	28.802	12.315	1.00	14.09	BA	N
ATOM	1949	C	LYS	B	122	-30.053	27.898	6.886	1.00	11.06	BA	C
ATOM	1950	O	LYS	B	122	-30.830	28.453	7.652	1.00	10.61	BA	O
ATOM	1951	N	GLY	B	123	-30.376	26.845	6.149	1.00	9.18	BA	N
ATOM	1952	CA	GLY	B	123	-31.657	26.174	6.229	1.00	9.46	BA	C
ATOM	1953	C	GLY	B	123	-31.576	24.858	5.519	1.00	8.71	BA	C
ATOM	1954	O	GLY	B	123	-30.560	24.420	5.076	1.00	9.86	BA	O
ATOM	1955	N	THR	B	124	-32.723	24.178	5.492	1.00	9.53	BA	N
ATOM	1956	CA	THR	B	124	-32.816	22.827	4.884	1.00	10.37	BA	C
ATOM	1957	CB	THR	B	124	-33.581	21.884	5.826	1.00	8.60	BA	C
ATOM	1958	OG1	THR	B	124	-34.929	22.357	6.006	1.00	9.91	BA	O
ATOM	1959	CG2	THR	B	124	-32.941	21.839	7.199	1.00	10.67	BA	C
ATOM	1960	C	THR	B	124	-33.464	22.865	3.528	1.00	11.15	BA	C
ATOM	1961	O	THR	B	124	-34.247	23.768	3.213	1.00	10.75	BA	O
ATOM	1962	N	LEU	B	125	-33.158	21.854	2.732	1.00	10.75	BA	N
ATOM	1963	CA	LEU	B	125	-33.850	21.680	1.420	1.00	10.40	BA	C
ATOM	1964	CB	LEU	B	125	-32.967	22.229	0.306	1.00	10.90	BA	C
ATOM	1965	CG	LEU	B	125	-33.550	22.088	-1.123	1.00	10.20	BA	C
ATOM	1966	CD1	LEU	B	125	-33.111	23.248	-1.980	1.00	11.93	BA	C
ATOM	1967	CD2	LEU	B	125	-33.112	20.776	-1.801	1.00	12.78	BA	C
ATOM	1968	C	LEU	B	125	-34.133	20.168	1.247	1.00	10.80	BA	C
ATOM	1969	O	LEU	B	125	-33.255	19.339	1.534	1.00	11.30	BA	O
ATOM	1970	N	THR	B	126	-35.329	19.866	0.767	1.00	9.69	BA	N
ATOM	1971	CA	ATHR	B	126	-35.632	18.471	0.490	0.50	10.44	BA	C
ATOM	1972	CA	BTHR	B	126	-35.751	18.506	0.602	0.50	11.08	BA	C
ATOM	1973	CB	ATHR	B	126	-36.157	17.700	1.757	0.50	9.77	BA	C
ATOM	1974	CB	BTHR	B	126	-36.604	18.097	1.831	0.50	12.17	BA	C
ATOM	1975	OG1	ATHR	B	126	-36.167	16.302	1.552	0.50	10.20	BA	O
ATOM	1976	OG1	BTHR	B	126	-37.869	18.771	1.831	0.50	17.79	BA	O
ATOM	1977	CG2	ATHR	B	126	-37.616	17.989	2.053	0.50	13.51	BA	C

ATOM	1978	CG2BTHR	B 126	-35.885	18.460	3.136	0.50	9.03	BA	C
ATOM	1979	C THR	B 126	-36.565	18.349	-0.685	1.00	11.28	BA	C
ATOM	1980	O THR	B 126	-37.358	19.263	-0.986	1.00	10.80	BA	O
ATOM	1981	N LEU	B 127	-36.500	17.172	-1.334	1.00	12.31	BA	N
ATOM	1982	CA LEU	B 127	-37.560	16.861	-2.315	1.00	14.47	BA	C
ATOM	1983	CB LEU	B 127	-37.183	15.664	-3.193	1.00	15.89	BA	C
ATOM	1984	CG LEU	B 127	-36.057	15.928	-4.151	1.00	14.36	BA	C
ATOM	1985	CD1 LEU	B 127	-35.576	14.633	-4.783	1.00	17.72	BA	C
ATOM	1986	CD2 LEU	B 127	-36.596	16.795	-5.367	1.00	17.37	BA	C
ATOM	1987	C LEU	B 127	-38.898	16.459	-1.690	1.00	14.78	BA	C
ATOM	1988	O LEU	B 127	-38.981	15.893	-0.561	1.00	16.76	BA	O
ATOM	1989	N LYS	B 128	-39.977	16.725	-2.384	1.00	15.29	BA	N
ATOM	1990	CA LYS	B 128	-41.232	16.100	-1.962	1.00	16.87	BA	C
ATOM	1991	CB LYS	B 128	-41.982	17.058	-1.015	1.00	18.80	BA	C
ATOM	1992	CG LYS	B 128	-43.290	16.488	-0.474	1.00	25.04	BA	C
ATOM	1993	CD LYS	B 128	-43.975	17.491	0.476	1.00	31.02	BA	C
ATOM	1994	CE LYS	B 128	-45.186	16.865	1.140	1.00	33.04	BA	C
ATOM	1995	NZ LYS	B 128	-44.824	16.521	2.543	1.00	36.46	BA	N
ATOM	1996	C LYS	B 128	-42.044	15.830	-3.190	1.00	17.62	BA	C
ATOM	1997	O LYS	B 128	-42.034	16.581	-4.168	1.00	18.77	BA	O
ATOM	1998	OXT LYS	B 128	-42.840	14.831	-3.193	1.00	19.21	BA	O
ATOM	1999	CU CUF	1	-12.589	17.767	-12.503	1.00	19.72	CU	
ATOM	2000	CU CUG	1	-46.986	26.786	-12.948	1.00	16.77	CU	
ATOM	2001	CU CUI	1	-20.359	26.330	-9.881	1.00	28.14	CU	
ATOM	2002	CU CUJ	1	-12.454	-9.224	-8.502	1.00	20.80	CU	
ATOM	2003	CU CUC	1	-3.365	12.626	-25.893	1.00	11.79	CU	
ATOM	2004	CU CUD	1	-24.028	36.223	3.615	1.00	12.77	CU	
ATOM	2005	O3 TRS	E 1	-14.395	17.729	-14.212	1.00	18.99	O	
ATOM	2006	C3 TRS	E 1	-13.744	18.792	-14.981	1.00	17.71	C	
ATOM	2007	C TRS	E 1	-13.303	20.054	-14.270	1.00	19.95	C	
ATOM	2008	N TRS	E 1	-12.245	19.647	-13.311	1.00	23.64	N	
ATOM	2009	C2 TRS	E 1	-12.646	20.904	-15.391	1.00	20.56	C	
ATOM	2010	O2 TRS	E 1	-13.751	21.432	-16.119	1.00	22.04	O	
ATOM	2011	C1 TRS	E 1	-14.578	20.634	-13.603	1.00	21.59	C	
ATOM	2012	O1 TRS	E 1	-14.110	21.779	-12.870	1.00	25.97	O	
ATOM	2013	O3 TRS	H 1	-20.199	24.487	-8.099	1.00	30.83	O	
ATOM	2014	C3 TRS	H 1	-19.170	25.140	-7.327	1.00	26.67	C	
ATOM	2015	C TRS	H 1	-18.054	25.603	-8.297	1.00	28.27	C	
ATOM	2016	N TRS	H 1	-18.705	26.526	-9.288	1.00	27.07	N	
ATOM	2017	C2 TRS	H 1	-16.960	26.287	-7.442	1.00	28.50	C	
ATOM	2018	O2 TRS	H 1	-16.584	25.377	-6.397	1.00	27.89	O	
ATOM	2019	C1 TRS	H 1	-17.533	24.348	-9.027	1.00	30.24	C	
ATOM	2020	O1 TRS	H 1	-16.299	24.679	-9.651	1.00	35.18	O	
ATOM	2021	O3 TRS	K 1	-0.029	20.230	-16.490	1.00	18.22	O	
ATOM	2022	C3 TRS	K 1	0.277	21.622	-16.799	1.00	20.71	C	
ATOM	2023	C TRS	K 1	0.043	22.527	-15.536	1.00	22.13	C	
ATOM	2024	N TRS	K 1	0.628	21.762	-14.406	1.00	21.69	N	
ATOM	2025	C2 TRS	K 1	0.814	23.848	-15.561	1.00	22.32	C	
ATOM	2026	O2 TRS	K 1	2.222	23.659	-15.573	1.00	19.28	O	
ATOM	2027	C1 TRS	K 1	-1.480	22.744	-15.300	1.00	22.41	C	
ATOM	2028	O1 TRS	K 1	-1.683	23.173	-13.933	1.00	25.94	O	
ATOM	2029	CL CLL	1	-5.194	0.875	-23.597	1.00	15.73	CL	
ATOM	2030	O HOH	M 1	-22.510	17.883	-0.926	1.00	10.81	O	
ATOM	2031	O HOH	M 2	-2.165	4.196	-13.909	1.00	10.99	O	
ATOM	2032	O HOH	M 3	-20.520	31.438	6.133	1.00	10.83	O	

ATOM	2033	O	HOH M	4	-1.385	26.078	-24.065	1.00	10.91	O
ATOM	2034	O	HOH M	5	-2.078	1.726	-17.768	1.00	9.95	O
ATOM	2035	O	HOH M	6	-8.473	19.636	-27.883	1.00	11.85	O
ATOM	2036	O	HOH M	7	0.962	15.797	-29.987	1.00	9.05	O
ATOM	2037	O	HOH M	9	-36.002	34.541	2.125	1.00	13.19	O
ATOM	2038	O	HOH M	10	-13.390	21.275	-21.901	1.00	12.95	O
ATOM	2039	O	HOH N	1	-20.833	19.771	7.832	1.00	12.95	O
ATOM	2040	O	HOH N	2	-12.967	13.183	-12.294	1.00	12.62	O
ATOM	2041	O	HOH N	3	-8.107	15.983	-28.633	1.00	10.13	O
ATOM	2042	O	HOH N	4	-15.976	25.929	-0.964	1.00	12.98	O
ATOM	2043	O	HOH N	5	-2.220	5.034	-11.270	1.00	10.43	O
ATOM	2044	O	HOH N	6	-21.514	14.876	-20.808	1.00	10.05	O
ATOM	2045	O	HOH N	7	-1.070	17.610	-15.517	1.00	14.59	O
ATOM	2046	O	HOH O	1	-25.593	24.437	10.325	1.00	13.43	O
ATOM	2047	O	HOH O	2	-36.340	21.029	4.046	1.00	16.61	O
ATOM	2048	O	HOH O	3	-4.551	14.538	-35.314	1.00	9.72	O
ATOM	2049	O	HOH O	4	-1.042	7.397	-10.154	1.00	11.14	O
ATOM	2050	O	HOH O	5	-24.206	34.295	11.020	1.00	14.60	O
ATOM	2051	O	HOH O	6	-17.046	30.997	5.078	1.00	16.55	O
ATOM	2052	O	HOH O	7	-6.842	16.636	-17.845	1.00	15.56	O
ATOM	2053	O	HOH O	8	-21.835	20.585	10.202	1.00	15.34	O
ATOM	2054	O	HOH O	9	-9.130	4.100	-28.845	1.00	14.12	O
ATOM	2055	O	HOH O	10	-24.649	20.864	7.999	1.00	15.01	O
ATOM	2056	O	HOH O	11	-31.453	33.201	8.486	1.00	15.99	O
ATOM	2057	O	HOH O	12	-17.633	19.030	5.741	1.00	13.66	O
ATOM	2058	O	HOH O	13	-30.825	30.499	11.327	1.00	17.02	O
ATOM	2059	O	HOH O	14	-18.513	0.628	-25.118	1.00	13.84	O
ATOM	2060	O	HOH O	15	2.199	1.315	-19.230	1.00	12.43	O
ATOM	2061	O	HOH P	1	-24.615	26.082	-9.539	1.00	15.99	O
ATOM	2062	O	HOH P	2	-40.846	23.157	-6.349	1.00	15.67	O
ATOM	2063	O	HOH P	3	-6.651	5.194	-30.153	1.00	13.38	O
ATOM	2064	O	HOH P	4	-1.401	-0.970	-18.666	1.00	15.35	O
ATOM	2065	O	HOH P	5	-24.888	26.928	11.187	1.00	17.48	O
ATOM	2066	O	HOH P	6	-22.602	4.558	-7.668	1.00	14.83	O
ATOM	2067	O	HOH P	7	-7.308	29.148	-20.073	1.00	14.00	O
ATOM	2068	O	HOH Q	1	-22.907	16.084	-2.926	1.00	14.58	O
ATOM	2069	O	HOH Q	2	-20.612	18.932	-28.178	1.00	12.41	O
ATOM	2070	O	HOH Q	3	-38.257	31.921	-11.324	1.00	11.56	O
ATOM	2071	O	HOH Q	4	-3.317	27.564	-21.347	1.00	14.77	O
ATOM	2072	O	HOH Q	5	-32.592	30.613	7.106	1.00	17.55	O
ATOM	2073	O	HOH Q	6	1.301	15.395	-15.162	1.00	15.21	O
ATOM	2074	O	HOH Q	7	-23.330	14.643	-18.717	1.00	16.37	O
ATOM	2075	O	HOH Q	8	-3.464	8.371	-9.145	1.00	13.93	O
ATOM	2076	O	HOH Q	9	-27.671	15.912	3.558	1.00	15.78	O
ATOM	2077	O	HOH Q	10	-8.100	19.013	-30.500	1.00	13.47	O
ATOM	2078	O	HOH Q	11	-21.091	24.632	9.247	1.00	19.56	O
ATOM	2079	O	HOH Q	12	-31.312	44.929	5.835	1.00	18.02	O
ATOM	2080	O	HOH Q	13	-15.097	29.927	-6.741	1.00	15.55	O
ATOM	2081	O	HOH Q	14	-33.325	37.145	-7.814	1.00	14.78	O
ATOM	2082	O	HOH Q	15	-44.506	21.975	-9.864	1.00	18.12	O
ATOM	2083	O	HOH Q	16	6.041	19.012	-20.273	1.00	16.12	O
ATOM	2084	O	HOH Q	17	2.897	10.134	-12.310	1.00	15.76	O
ATOM	2085	O	HOH Q	18	-13.806	19.761	-2.624	1.00	17.22	O
ATOM	2086	O	HOH Q	19	-0.970	11.033	-10.705	1.00	18.93	O
ATOM	2087	O	HOH Q	20	-9.203	16.896	-31.994	1.00	16.71	O

ATOM	2088	O	HOH	Q	21	-1.511	24.748	-26.645	1.00	15.99	O
ATOM	2089	O	HOH	Q	22	-20.120	15.875	-29.718	1.00	15.15	O
ATOM	2090	O	HOH	Q	23	1.183	21.917	-24.797	1.00	16.33	O
ATOM	2091	O	HOH	Q	24	-29.923	38.732	12.574	1.00	17.44	O
ATOM	2092	O	HOH	S	1	-7.436	-3.420	-17.844	1.00	16.28	O
ATOM	2093	O	HOH	S	2	-17.981	10.472	-9.064	1.00	19.24	O
ATOM	2094	O	HOH	S	3	-8.127	16.514	-34.583	1.00	15.19	O
ATOM	2095	O	HOH	S	4	-9.301	22.548	-16.206	1.00	18.79	O
ATOM	2096	O	HOH	S	5	-23.220	22.932	9.574	1.00	18.06	O
ATOM	2097	O	HOH	S	6	-24.914	14.483	5.038	1.00	17.43	O
ATOM	2098	O	HOH	S	7	-18.936	24.075	-26.528	1.00	15.72	O
ATOM	2099	O	HOH	S	8	-17.117	18.142	-14.499	1.00	17.81	O
ATOM	2100	O	HOH	S	9	3.628	5.705	-17.611	1.00	21.70	O
ATOM	2101	O	HOH	S	10	-25.921	32.222	-10.866	1.00	17.15	O
ATOM	2102	O	HOH	S	11	-10.962	29.011	6.364	1.00	19.32	O
ATOM	2103	O	HOH	R	1	-17.757	27.706	10.444	1.00	19.59	O
ATOM	2104	O	HOH	R	2	-14.150	11.448	-10.491	1.00	18.99	O
ATOM	2105	O	HOH	R	3	3.235	20.543	-14.637	1.00	18.35	O
ATOM	2106	O	HOH	R	4	-27.484	21.119	-12.347	1.00	17.77	O
ATOM	2107	O	HOH	R	5	-25.198	14.723	-3.702	1.00	21.03	O
ATOM	2108	O	HOH	R	6	-33.591	16.634	-13.560	1.00	16.17	O
ATOM	2109	O	HOH	R	7	-20.849	8.264	-30.333	1.00	16.63	O
ATOM	2110	O	HOH	R	8	-35.273	37.125	-4.236	1.00	16.72	O
ATOM	2111	O	HOH	R	10	-7.066	11.893	-11.155	1.00	19.81	O
ATOM	2112	O	HOH	R	11	-4.758	-2.311	-23.346	1.00	16.31	O
ATOM	2113	O	HOH	R	12	-34.369	15.291	-0.919	1.00	20.24	O
ATOM	2114	O	HOH	R	13	-40.156	20.804	-15.225	1.00	17.21	O
ATOM	2115	O	HOH	R	14	-18.475	-2.099	-6.192	1.00	15.81	O
ATOM	2116	O	HOH	R	15	1.479	8.869	-10.359	1.00	13.93	O
ATOM	2117	O	HOH	R	16	-28.020	28.673	-13.370	1.00	20.77	O
ATOM	2118	O	HOH	R	17	-17.929	45.633	-2.149	1.00	17.99	O
ATOM	2119	O	HOH	R	19	-21.995	13.898	1.680	1.00	19.40	O
ATOM	2120	O	HOH	R	20	3.028	23.610	-18.084	1.00	18.31	O
ATOM	2121	O	HOH	T	1	-0.761	17.920	-31.126	1.00	12.45	O
ATOM	2122	O	HOH	T	2	-32.453	28.087	9.963	1.00	16.90	O
ATOM	2123	O	HOH	T	3	-8.527	14.037	-12.112	1.00	17.75	O
ATOM	2124	O	HOH	T	4	-10.557	25.487	-29.822	1.00	18.73	O
ATOM	2125	O	HOH	T	5	-32.575	37.084	-10.404	1.00	15.44	O
ATOM	2126	O	HOH	T	6	-44.312	13.929	-8.179	1.00	21.48	O
ATOM	2127	O	HOH	T	7	-16.680	17.993	3.434	1.00	21.53	O
ATOM	2128	O	HOH	T	8	-39.342	27.852	0.753	1.00	21.03	O
ATOM	2129	O	HOH	T	9	-25.048	11.537	-25.955	1.00	17.48	O
ATOM	2130	O	HOH	T	10	-1.719	1.773	-15.043	1.00	21.18	O
ATOM	2131	O	HOH	T	11	4.480	20.893	-19.031	1.00	20.48	O
ATOM	2132	O	HOH	T	12	-21.044	32.180	-4.783	1.00	22.11	O
ATOM	2133	O	HOH	T	13	-16.653	12.138	-11.168	1.00	19.96	O
ATOM	2134	O	HOH	U	2	-4.836	-4.639	-16.787	1.00	19.32	O
ATOM	2135	O	HOH	U	3	-18.099	-4.662	-6.878	1.00	16.76	O
ATOM	2136	O	HOH	U	4	-18.919	18.185	8.943	1.00	20.16	O
ATOM	2137	O	HOH	U	5	-30.455	36.194	11.212	1.00	20.65	O
ATOM	2138	O	HOH	U	6	-43.521	20.322	-12.148	1.00	16.75	O
ATOM	2139	O	HOH	U	7	-34.420	13.489	-8.214	1.00	21.56	O
ATOM	2140	O	HOH	U	8	-29.521	3.004	-15.755	1.00	18.13	O
ATOM	2141	O	HOH	V	1	-33.695	40.556	10.112	1.00	21.59	O
ATOM	2142	O	HOH	V	2	-12.324	27.448	-23.227	1.00	20.10	O

ATOM	2143	O	HOH V	3	-16.423	-0.190	-6.400	1.00	19.46	O
ATOM	2144	O	HOH V	4	-40.549	32.938	-9.993	1.00	16.37	O
ATOM	2145	O	HOH V	5	2.109	-1.429	-28.892	1.00	18.98	O
ATOM	2146	O	HOH V	6	-48.964	26.843	-11.461	1.00	17.61	O
ATOM	2147	O	HOH V	7	-23.987	30.517	-10.117	1.00	19.75	O
ATOM	2148	O	HOH V	8	-19.680	23.331	-19.957	1.00	19.58	O
ATOM	2149	O	HOH V	9	-26.530	24.901	-11.345	1.00	18.75	O
ATOM	2150	O	HOH V	10	-26.115	29.538	15.363	1.00	22.29	O
ATOM	2151	O	HOH V	11	-38.741	21.580	3.344	1.00	20.06	O
ATOM	2152	O	HOH V	12	-34.346	9.778	-5.451	1.00	20.58	O
ATOM	2153	O	HOH V	13	-12.561	-11.077	-10.052	1.00	21.18	O
ATOM	2154	O	HOH V	14	-27.283	13.412	-2.701	1.00	20.44	O
ATOM	2155	O	HOH V	15	-9.243	25.551	3.744	1.00	21.24	O
ATOM	2156	O	HOH V	16	-25.576	14.960	-23.254	1.00	17.70	O
ATOM	2157	O	HOH V	17	-12.466	-4.300	-9.133	1.00	17.22	O
ATOM	2158	O	HOH V	18	-12.338	23.852	-13.018	1.00	21.24	O
ATOM	2159	O	HOH V	19	-29.634	17.313	7.084	1.00	22.51	O
ATOM	2160	O	HOH V	20	5.598	15.122	-18.802	1.00	21.66	O
ATOM	2161	O	HOH V	21	-10.006	28.669	-19.777	1.00	19.65	O
ATOM	2162	O	HOH V	22	-12.110	-2.209	-21.954	1.00	21.45	O
ATOM	2163	O	HOH V	23	7.656	5.832	-20.233	1.00	18.94	O
ATOM	2164	O	HOH W	1	-17.623	31.220	7.718	1.00	20.70	O
ATOM	2165	O	HOH W	3	-38.198	22.657	-14.766	1.00	16.39	O
ATOM	2166	O	HOH W	4	-19.088	-5.898	-9.369	1.00	23.18	O
ATOM	2167	O	HOH W	5	-11.626	18.632	-32.858	1.00	20.35	O
ATOM	2168	O	HOH W	6	-37.845	13.473	-0.132	1.00	23.13	O
ATOM	2169	O	HOH W	7	-16.513	-3.149	-15.080	1.00	21.11	O
ATOM	2170	O	HOH W	8	0.608	2.001	-31.192	1.00	19.32	O
ATOM	2171	O	HOH W	9	-26.033	3.641	-12.403	1.00	17.41	O
ATOM	2172	O	HOH W	10	-25.802	15.654	-6.372	1.00	22.13	O
ATOM	2173	O	HOH W	11	2.664	12.866	-12.102	1.00	20.32	O
ATOM	2174	O	HOH W	12	-21.673	20.131	-25.859	1.00	18.92	O
ATOM	2175	O	HOH W	13	-7.116	17.971	-15.215	1.00	22.99	O
ATOM	2176	O	HOH W	14	-16.587	29.714	9.499	1.00	30.11	O
ATOM	2177	O	HOH W	15	-19.633	21.809	-7.736	1.00	23.00	O
ATOM	2178	O	HOH W	16	-25.763	22.399	-10.496	1.00	21.21	O
ATOM	2179	O	HOH W	17	-14.083	-2.815	-20.401	1.00	24.10	O
ATOM	2180	O	HOH X	1	-23.795	17.254	-7.379	1.00	21.68	O
ATOM	2181	O	HOH X	2	-36.414	18.215	5.425	1.00	22.86	O
ATOM	2182	O	HOH X	3	-17.439	-6.501	-11.656	1.00	23.44	O
ATOM	2183	O	HOH X	4	-21.988	45.223	-3.722	1.00	20.13	O
ATOM	2184	O	HOH X	5	-27.584	48.665	6.928	1.00	23.86	O
ATOM	2185	O	HOH X	6	-2.756	-3.280	-18.770	1.00	21.56	O
ATOM	2186	O	HOH X	7	-24.477	18.138	-20.671	1.00	21.49	O
ATOM	2187	O	HOH X	8	-40.497	36.916	3.238	1.00	21.42	O
ATOM	2188	O	HOH X	9	8.966	19.362	-24.880	1.00	20.88	O
ATOM	2189	O	HOH X	10	-23.697	16.074	8.956	1.00	23.60	O
ATOM	2190	O	HOH X	11	-8.599	-3.005	-28.451	1.00	23.12	O
ATOM	2191	O	HOH X	12	-31.443	43.066	-4.296	1.00	22.20	O
ATOM	2192	O	HOH X	13	-23.613	12.171	-14.740	1.00	22.10	O
ATOM	2193	O	HOH X	14	-11.320	-2.378	-25.817	1.00	21.79	O
ATOM	2194	O	HOH X	15	-16.339	20.680	-9.078	1.00	21.91	O
ATOM	2195	O	HOH X	16	-43.554	17.364	-17.963	1.00	23.55	O
ATOM	2196	O	HOH X	17	-21.761	14.017	-14.268	1.00	24.63	O
ATOM	2197	O	HOH Y	1	-35.669	17.258	8.899	1.00	22.05	O

ATOM	2198	O	HOH Y	2	-32.587	38.945	12.158	1.00	20.08	O
ATOM	2199	O	HOH Y	3	-40.096	32.455	-7.404	1.00	20.96	O
ATOM	2200	O	HOH Y	4	-9.791	24.095	-14.103	1.00	26.73	O
ATOM	2201	O	HOH Y	5	-14.915	39.523	6.992	1.00	23.62	O
ATOM	2202	O	HOH Y	6	-49.362	23.161	-11.901	1.00	18.92	O
ATOM	2203	O	HOH Y	7	-5.083	27.289	-26.513	1.00	20.73	O
ATOM	2204	O	HOH Y	8	-13.601	18.863	-5.245	1.00	21.94	O
ATOM	2205	O	HOH Y	9	-27.493	42.106	-10.079	1.00	21.32	O
ATOM	2206	O	HOH Y	10	-11.001	10.984	-36.713	1.00	19.82	O
ATOM	2207	O	HOH Y	11	-24.624	3.239	-20.884	1.00	18.66	O
ATOM	2208	O	HOH Y	12	-9.048	27.172	5.904	1.00	24.00	O
ATOM	2209	O	HOH Y	13	-0.238	19.139	-13.253	1.00	23.65	O
ATOM	2210	O	HOH Y	14	-17.407	14.641	1.205	1.00	20.10	O
ATOM	2211	O	HOH Y	15	-7.079	16.550	-12.747	1.00	23.95	O
ATOM	2212	O	HOH Y	17	-33.933	38.763	4.405	1.00	25.10	O
ATOM	2213	O	HOH Y	18	-9.260	31.270	6.033	1.00	28.93	O
ATOM	2214	O	HOH Y	19	-16.702	27.839	13.117	1.00	28.18	O
ATOM	2215	O	HOH Y	20	-14.821	41.702	5.200	1.00	22.58	O
ATOM	2216	O	HOH Y	21	-8.412	25.868	1.198	1.00	28.42	O
ATOM	2217	O	HOH Y	22	-10.970	14.110	-10.698	1.00	25.28	O
ATOM	2218	O	HOH Z	1	4.227	16.849	-14.885	1.00	22.60	O
ATOM	2219	O	HOH Z	2	-11.191	30.135	-10.002	1.00	21.87	O
ATOM	2220	O	HOH Z	3	-15.628	1.812	-4.329	1.00	20.69	O
ATOM	2221	O	HOH Z	4	-14.680	15.445	-31.568	1.00	28.30	O
ATOM	2222	O	HOH Z	5	-10.884	38.058	-2.736	1.00	21.25	O
ATOM	2223	O	HOH Z	6	5.989	4.546	-18.312	1.00	23.34	O
ATOM	2224	O	HOH Z	7	-16.323	22.525	-11.425	1.00	24.40	O
ATOM	2225	O	HOH Z	8	-23.821	0.494	-9.268	1.00	27.78	O
ATOM	2226	O	HOH Z	9	-13.678	17.506	-0.941	1.00	23.08	O
ATOM	2227	O	HOH Z	10	-24.417	12.737	-17.383	1.00	23.32	O
ATOM	2228	O	HOH Z	11	-8.600	26.446	-13.236	1.00	22.57	O
ATOM	2229	O	HOH Z	12	-14.825	28.330	-17.581	1.00	25.49	O
ATOM	2230	O	HOH Z	13	-34.155	44.033	8.540	1.00	22.43	O
ATOM	2231	O	HOH Z	14	-36.249	44.239	-0.546	1.00	23.25	O
ATOM	2232	O	HOH Z	15	-9.507	28.901	-13.727	1.00	23.77	O
ATOM	2233	O	HOH Z	17	-18.176	21.827	-13.394	1.00	25.81	O
ATOM	2234	O	HOH Z	19	-14.850	40.497	1.476	1.00	20.80	O
ATOM	2235	O	HOH Z	20	-1.096	5.299	-34.106	1.00	23.94	O
ATOM	2236	O	HOH Z	21	-30.788	13.564	5.307	1.00	22.57	O
ATOM	2237	O	HOH Z	22	-32.814	47.461	11.041	1.00	24.07	O
ATOM	2238	O	HOH Z	23	-38.615	17.887	6.636	1.00	23.21	O
ATOM	2239	O	HOH Z	24	-24.060	17.508	-29.875	1.00	23.59	O
ATOM	2240	O	HOH Z	25	-19.662	21.364	11.837	1.00	26.32	O
ATOM	2241	O	HOH Z	26	1.733	1.385	-15.470	1.00	25.79	O
ATOM	2242	O	HOH Z	27	-15.238	19.241	7.100	1.00	25.69	O
ATOM	2243	O	HOH Z	28	-40.177	20.520	-17.999	1.00	26.12	O
ATOM	2244	O	HOH Z	29	-25.162	27.347	13.891	1.00	26.01	O
ATOM	2245	O	HOH a	1	-29.131	-7.163	-12.501	1.00	31.46	O
ATOM	2246	O	HOH a	2	-42.178	27.037	-12.200	1.00	18.87	O
ATOM	2247	O	HOH a	3	-7.501	-2.315	-14.111	1.00	27.61	O
ATOM	2248	O	HOH a	4	-17.432	48.418	-1.772	1.00	23.79	O
ATOM	2249	O	HOH a	5	-18.537	26.108	-23.379	1.00	20.64	O
ATOM	2250	O	HOH a	6	-33.409	47.380	0.223	1.00	24.71	O
ATOM	2251	O	HOH a	7	-40.723	31.880	-3.163	1.00	23.15	O
ATOM	2252	O	HOH a	8	-7.798	-5.801	-28.449	1.00	19.36	O

ATOM	2253	O	HOH a	9	-31.877	25.489	-18.299	1.00	22.08	O
ATOM	2254	O	HOH a	10	-36.903	8.446	0.033	1.00	21.49	O
ATOM	2255	O	HOH a	11	-17.631	48.338	2.261	1.00	23.37	O
ATOM	2256	O	HOH a	12	-27.892	-4.302	-18.921	1.00	20.45	O
ATOM	2257	O	HOH a	13	-33.070	48.916	14.821	1.00	31.11	O
ATOM	2258	O	HOH a	14	-40.457	25.548	-0.650	1.00	28.88	O
ATOM	2259	O	HOH a	15	-30.831	47.368	-1.878	1.00	25.07	O
ATOM	2260	O	HOH a	16	-4.876	-2.854	-20.424	1.00	23.18	O
ATOM	2261	O	HOH a	17	-12.117	22.574	5.278	1.00	30.19	O
ATOM	2262	O	HOH a	18	-19.428	14.302	-12.692	1.00	24.17	O
ATOM	2263	O	HOH a	19	-12.330	26.167	-11.464	1.00	24.95	O
ATOM	2264	O	HOH a	20	-10.761	-4.932	-12.705	1.00	24.40	O
ATOM	2265	O	HOH b	1	-19.382	30.346	9.146	1.00	23.79	O
ATOM	2266	O	HOH b	2	-22.332	27.120	10.247	1.00	25.31	O
ATOM	2267	O	HOH b	3	-37.655	15.532	-11.825	1.00	25.71	O
ATOM	2268	O	HOH b	4	-17.747	29.381	-7.849	1.00	31.29	O
ATOM	2269	O	HOH b	5	-9.458	24.378	-5.683	1.00	25.89	O
ATOM	2270	O	HOH b	6	5.543	21.255	-16.173	1.00	25.40	O
ATOM	2271	O	HOH b	7	-12.367	14.256	-32.613	1.00	27.52	O
ATOM	2272	O	HOH b	8	-24.032	51.474	2.622	1.00	22.68	O
ATOM	2273	O	HOH b	9	-42.605	13.778	-15.461	1.00	29.72	O
ATOM	2274	O	HOH b	10	-22.245	15.671	-16.494	1.00	27.52	O
ATOM	2275	O	HOH b	11	-10.191	26.890	-0.433	1.00	23.91	O
ATOM	2276	O	HOH c	1	-25.866	10.333	-18.323	1.00	20.86	O
ATOM	2277	O	HOH c	2	-23.734	47.044	-3.153	1.00	24.67	O
ATOM	2278	O	HOH c	3	-19.525	38.013	-7.114	1.00	22.02	O
ATOM	2279	O	HOH c	5	-26.377	0.613	-21.878	1.00	21.74	O
ATOM	2280	O	HOH c	6	-9.462	19.921	-14.702	1.00	22.60	O
ATOM	2281	O	HOH c	7	-11.419	29.234	9.075	1.00	30.43	O
ATOM	2282	O	HOH c	8	-37.051	41.800	-1.236	1.00	22.27	O
ATOM	2283	O	HOH c	9	4.755	2.373	-28.722	1.00	26.25	O
ATOM	2284	O	HOH c	10	-21.986	16.880	-5.192	1.00	24.31	O
ATOM	2285	O	HOH c	11	-30.446	38.056	-11.766	1.00	25.12	O
ATOM	2286	O	HOH c	12	-38.349	37.810	1.744	1.00	29.82	O
ATOM	2287	O	HOH c	13	-36.589	23.226	-17.089	1.00	34.13	O
ATOM	2288	O	HOH c	14	-46.663	17.171	-12.132	1.00	21.15	O
ATOM	2289	O	HOH c	15	5.529	10.082	-11.852	1.00	24.90	O
ATOM	2290	O	HOH c	16	-14.394	26.717	-10.031	1.00	25.26	O
ATOM	2291	O	HOH c	17	-4.829	29.237	-24.084	1.00	28.76	O
ATOM	2292	O	HOH c	18	-13.832	29.312	-8.984	1.00	25.44	O
ATOM	2293	O	HOH c	19	8.869	9.106	-29.396	1.00	23.06	O
ATOM	2294	O	HOH d	1	-41.331	26.020	-5.382	1.00	25.32	O
ATOM	2295	O	HOH d	2	-10.081	41.384	-3.506	1.00	24.95	O
ATOM	2296	O	HOH d	3	-25.693	17.475	-24.211	1.00	21.08	O
ATOM	2297	O	HOH d	4	-40.544	15.560	1.727	1.00	23.02	O
ATOM	2298	O	HOH d	5	-26.916	12.775	-24.636	1.00	21.61	O
ATOM	2299	O	HOH e	1	-13.806	27.904	-25.699	1.00	28.08	O
ATOM	2300	O	HOH e	2	-20.648	23.852	-17.339	1.00	24.59	O
ATOM	2301	O	HOH e	3	-18.757	24.044	-29.109	1.00	24.80	O
ATOM	2302	O	HOH e	4	-21.788	4.648	-28.837	1.00	22.34	O
ATOM	2303	O	HOH e	6	-23.944	49.196	-1.254	1.00	26.24	O
ATOM	2304	O	HOH e	7	-21.522	50.616	-1.066	1.00	22.44	O
ATOM	2305	O	HOH e	8	-9.652	20.764	-32.123	1.00	27.01	O
ATOM	2306	O	HOH e	9	-39.938	25.820	-3.228	1.00	28.42	O
ATOM	2307	O	HOH e	10	-46.226	20.081	-12.047	1.00	26.84	O

ATOM	2308	O	HOH e	11	-17.953	15.584	-3.604	1.00	27.61	O
ATOM	2309	O	HOH e	12	-15.201	11.068	-32.086	1.00	29.02	O
ATOM	2310	O	HOH e	13	-4.987	5.214	-32.711	1.00	25.74	O
ATOM	2311	O	HOH e	14	-14.736	49.437	0.746	1.00	23.71	O
ATOM	2312	O	HOH e	15	-20.958	28.526	12.575	1.00	31.56	O
ATOM	2313	O	HOH e	16	-32.167	49.931	1.240	1.00	26.39	O
ATOM	2314	O	HOH e	17	-27.499	9.937	-16.286	1.00	27.90	O
ATOM	2315	O	HOH e	18	-14.915	8.446	-4.404	1.00	23.41	O
ATOM	2316	O	HOH e	19	-13.558	23.694	9.533	1.00	29.20	O
ATOM	2317	O	HOH e	20	-7.802	30.520	-26.483	1.00	26.98	O
ATOM	2318	O	HOH e	21	-40.870	37.727	5.946	1.00	25.74	O
ATOM	2319	O	HOH e	22	-47.879	21.429	-10.252	1.00	25.36	O
ATOM	2320	O	HOH f	1	-24.516	12.539	-29.906	1.00	24.61	O
ATOM	2321	O	HOH f	2	-19.591	25.322	12.359	1.00	32.69	O
ATOM	2322	O	HOH f	3	-42.582	22.992	-3.320	1.00	25.14	O
ATOM	2323	O	HOH f	4	-46.499	31.800	-8.078	1.00	30.15	O
ATOM	2324	O	HOH f	5	-34.635	46.759	8.412	1.00	30.26	O
ATOM	2325	O	HOH f	6	-25.876	15.093	-20.139	1.00	24.72	O
ATOM	2326	O	HOH f	7	-19.360	14.894	-1.342	1.00	27.71	O
ATOM	2327	O	HOH f	8	-17.489	22.159	-30.522	1.00	29.27	O
ATOM	2328	O	HOH f	9	-28.671	25.689	-15.317	1.00	27.68	O
ATOM	2329	O	HOH f	10	-19.780	49.293	3.637	1.00	20.21	O
ATOM	2330	O	HOH f	11	1.081	4.387	-35.609	1.00	28.33	O
ATOM	2331	O	HOH f	12	-19.461	46.426	-4.195	1.00	24.96	O
ATOM	2332	O	HOH f	13	-40.562	33.972	3.519	1.00	33.71	O
ATOM	2333	O	HOH f	14	9.215	11.598	-31.394	1.00	24.71	O
ATOM	2334	O	HOH f	15	-13.867	21.909	7.295	1.00	29.96	O
ATOM	2335	O	HOH f	16	-14.323	28.028	9.094	1.00	30.40	O
ATOM	2336	O	HOH f	17	-11.775	28.432	-12.422	1.00	28.72	O
ATOM	2337	O	HOH f	18	-17.383	28.695	-23.593	1.00	31.39	O
ATOM	2338	O	HOH f	19	-24.658	10.079	-9.101	1.00	28.35	O
ATOM	2339	O	HOH f	20	-27.971	14.489	-12.152	1.00	32.57	O
ATOM	2340	O	HOH f	21	8.094	3.026	-20.522	1.00	29.99	O
ATOM	2341	O3	TRS g	1	-13.129	15.785	-8.550	1.00	38.20	O
ATOM	2342	C3	TRS g	1	-14.245	15.390	-9.319	1.00	36.32	C
ATOM	2343	C	TRS g	1	-14.270	16.293	-10.543	1.00	35.85	C
ATOM	2344	N	TRS g	1	-13.083	16.108	-11.453	1.00	32.42	N
ATOM	2345	C2	TRS g	1	-15.551	16.000	-11.339	1.00	36.49	C
ATOM	2346	O2	TRS g	1	-16.676	16.637	-10.743	1.00	42.41	O
ATOM	2347	C1	TRS g	1	-14.189	17.735	-10.018	1.00	34.75	C
ATOM	2348	O1	TRS g	1	-13.770	18.583	-11.073	1.00	27.24	O
ATOM	2349	O	HOH h	1	-42.589	28.615	-8.385	1.00	24.31	O