

Synthesis of Functionalized Benzoxaboroles

A THESIS

SUBMITTED TO THE FACULTY OF

UNIVERSITY OF MINNESOTA

BY

SRAVAN K JONNALAGADDA

IN PARTIAL FULFILLMENT OF THE REQUIREMENTS

FOR THE DEGREE OF

MASTER OF SCIENCE

Venkatram R Mereddy

OCTOBER 2014

© Sravan Jonnalagadda 2014

ACKNOWLEDGEMENTS

Foremost, I would like to express my appreciation and deepest gratitude to Dr. Venkatram R. Mereddy, my thesis advisor, for his valuable advice, continuous support and expert guidance. I am also thankful to him for his great amounts of time given to achieve my success throughout.

I would also like to thank Dr. Lester Drewes and Dr. Joseph Johnson for their valuable help in my professional development.

I would also like to thank the Department of Chemistry and Biochemistry for its financial support for the past three years. My thanks go to the faculty in the Department of Chemistry and Biochemistry for the knowledge they have taught me during this time. I thank all the members of my group for their support given me for the past three years.

Abstract

Synthesis of Functionalized Benzoxaboroles

Sravan K. Jonnalagadda

Master of Science (Department of Chemistry and Biochemistry),

University of Minnesota.

2-Formylphenylboronic acids upon reaction with activated olefins such as acrylates, methyl vinyl ketone, and acrylonitrile, etc. via Baylis-Hillman reaction to provide functionalized benzoxaboroles. The corresponding homologated benzoxaboroles were synthesized via Barbier allylation reaction of 2-formylphenylboronic acids with α -bromomethylacrylates. Several novel benzoxaborole derivatives were synthesized starting from 2-formylphenylboronic acid utilizing Passerini reaction and aldol reaction protocols as the key step.

An efficient methodology for the preparation of α -hydroxyamides via boric acid mediated addition of isonitriles on to aldehydes has been developed. The reaction of isonitriles with α -boronobenzaldehyde takes place under intramolecular catalysis conditions to provide functionalized benzoxaboroles.

Table of Contents

I. ACKNOWLEDGEMENTS	i
II. ABSTRACT	ii
III. TABLE OF CONTENTS	iii
IV. LIST OF SCHEMES	iv
V. LIST OF FIGURES	vii
VI. LIST OF ABBREVIATIONS	viii
VII. CHAPTER 1: BENZOXABOROLES	1
VIII. CHAPTER 2: NOVEL METHODOLOGIES FOR THE SYNTHESIS OF BENZOXABOROLES	20
IX. RESULTS AND DISCUSSION	21
X. CHAPTER 3: SYNTHESIS OF BENZOXABOROLES VIA PASSERINI REACTION	33
XI. RESULTS AND DISCUSSION	34
XII. CONCLUSIONS	42
XIII. SPECTRAL CHARACTERIZATION	43
XIV. REFERENCES	70
XV. APPENDIX	74

LIST OF SCHEMES

Scheme	Title of the scheme	Pg.No.
Scheme 1.1	Synthesis of benzoxaborole	3
Scheme 1.2	Synthesis of substituted benzoxaborole	3
Scheme 1.3	Synthesis of 3-substituted benzoxaborole	4
Scheme 1.4	Synthesis of benzoxaborole via multicomponent coupling of alkynes	4
Scheme 1.5	Synthesis of benzoboroxoles as anti-trypanosomal agents	5
Scheme 1.6	Synthesis of benzoboroxoles as anti-trypanosomal agents	7
Scheme 1.7	Synthesis of polymeric benzoxaboroles	8
Scheme 1.8	Synthesis of 3-amino-substituted benzoxaboroles	9
Scheme 1.9	Practical synthesis of benzoxaboroles	9
Scheme 1.10	Practical synthesis of chiral benzoxaboroles	10
Scheme 1.11	Chalcone–benzoxaborole hybrid molecules potent antitrypanosomal agents	11
Scheme 1.12	Synthesis of AN2690 and its analogs for the potential treatment of onychomycosis	12
Scheme 1.13	Synthesis and structure–activity relationships of novel benzoxaboroles as a new class of antimalarial agents	14

Scheme 1.14 Synthesis and SAR of novel benzoxaboroles as a new class of β -lactamase inhibitors	15
Scheme 1.15 Discovery of Novel Benzoxaborole-Based Potent Antitrypanosomal Agent	16
Scheme 1.16 Discovery and structure–activity study of a novel benzoxaborole anti-inflammatory agent (AN2728) for the potential topical treatment of psoriasis and atopic dermatitis	17
Scheme 1.17 Suzuki coupling–macrolactamization approach to the <i>AB-COD</i> bicyclic system of vancomycin	18
Scheme 1.18 Synthesis of amidobenzoxaboroles	19
Scheme 2.0 Baylis-Hillman Reaction	22
Scheme 2.1 Baylis Hillman reaction of β -Boronoaldehydes	23
Scheme 2.2 Preparation of α -cyclohexenonyl benzoxaborole	24
Scheme 2.3 Reaction of β -Boronoaldehydes with Acrolein	24
Scheme 2.4 Synthesis of Homologated Benzoxaboroles	25
Scheme 2.5 Synthesis of β -ketobenzoxaboroles	27
Scheme 3.1 Synthesis of functionalized benzoxaboroles	39

LIST OF FIGURES

Figure 1 Functionalized Benzoxaboroles	2
Figure 2 X-ray Crystal Structure of 13d	26
Figure 3 β -Ketobenzoxaboroles via aldol reaction	27

LIST OF TABLES

Table 1 Boric acid mediated addition of isonitriles and aldehydes for the synthesis of α -hydroxyamides	35
-----------------------------------------------------------------------------------------------------------------------	----

LIST OF ABBREVIATIONS

NBS	N-Bromosuccinimide
Cp	cyclopentyl
p-TsOH	p-Toluene Sulfonic acid
BnOH	Benzyl alcohol
NaH	Sodium hydride
BuLi	Butyl Lithium
B(iPrO) ₃	Triisopropyl borate
NaBH ₄	Sodium borohydrate
THF	Tetrahydrofuran
DMF	N, N - Dimethylformamide
Et ₃ N or TEA	Triethyl amine
TBDMS-Cl	Tertiary- butyldimethylsilyl chloride
DIBAL-H	Diisobutylaluminium hydride
MOM	Methoxymethyl
THP	Tetrahydropyran
DHP	Dihydropyran
PCC	Pyridinium chlorochromate
DCM	Dichloromethane
Pd(dppf)Cl ₂	(1,1'-Bis(diphenylphosphino)ferrocene)palladium(II) dichloride
Pd(PPh ₃) ₄	tetrakis(triphenylphosphine)palladium(0)
B(OMe) ₃	Trimethoxy borane

BBr ₃	Boron tribromide
NIS	N-iodo succinimide
BH	Baylis-Hillman
EWG	Electron with-drawing group
DABCO	1,4-diazabicyclo[2.2.2] octane
PBu ₃	Tributyl phosphine
EtOAc	Ethylacetate
MeOH	Methanol
TLC	Thin layer chromatography
LDA	Lithium diisopropylamide

Chapter 1: Benzoxaboroles

Introduction

Boron is the fifth element in the periodic table. Boron containing compounds have been highly utilized in several household purposes such as cleaning, insecticide and medicinal uses such as antiseptic, antibacterial, multiple enzyme inhibitors^{1,2}, etc. Boron containing compound, bortezomib, is a novel chemotherapeutic for multiple myeloma³. Apart from these medicinal uses, several boranes had been utilized in boron-neutron capture therapy in cancer treatment⁴. In industry, it is especially utilized in nuclear power plants as a poison to reduce the rate of fission. In material science, boron in the form of borates has been utilized in semiconductors.

Apart from boric acid and some other borates, most of the synthetic boron compounds are air sensitive with a few exceptions such as sodium borohydride. Benzoxaboroles **1** are cyclic hemiesters of phenylboronic acids⁵ which are stable compared to the acyclic boronic acids. There are only few scientists working on the synthesis of these hemiesters. Despite benzoxaborole had been known for ~60 years, these compounds are routinely used in cross coupling reactions in organic synthesis¹.

Benzoxaboroles also gained reputation for its medicinal usage⁶. For example, AN2728 **2** is used as *anti*-inflammatory activity against psoriasis, a common skin disease. 5-Fluoro-1,3-dihydro-1-hydroxy-2,1-benzoxaborole,

AN2690 **3**, is a broad-spectrum antifungal drug. 5-chloro-substituted benzoxaborole, AN2718 **4**, is being developed for the topical treatment of *tinea pedis* infection (**Figure 1**).

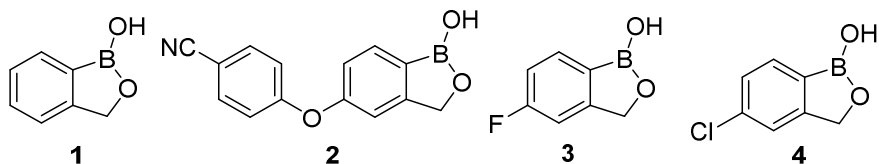
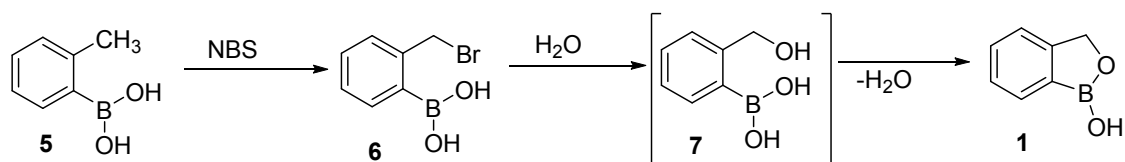


Figure 1 Functionalized Benzoxaboroles

In material science, benzoxaboroles has been used in the construction of molecular receptors, design of self-assembled molecular structures, building blocks in crystal engineering, use as steroid conjugates for molecular imprinting, dyes, biosensors of α -hydroxy-carboxylic acids, and biocides for plastic biodegradation etc.⁷ Because of the sugar-like structures, benzoxaboroles have been found to selectively complex with natural oligosaccharides to effect direct glycosidation. They are known for their selective recognition of cell surface glycoconjugates.⁸

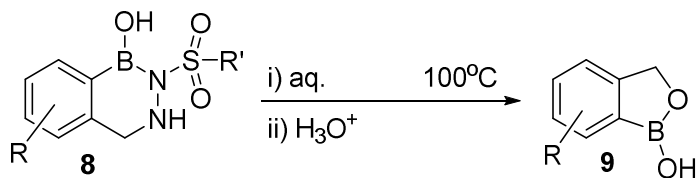
The following are some of the synthetic methods that have been reported for the preparation of benzoxaboroles and their development as medicinal agents.

2-Methylphenylboronic acid **5** was brominated using N-bromosuccinimide, followed by hydrolysis of the bromide **6** to benzyl alcohol intermediate **7**, which upon intramolecular esterification yielded unsubstituted benzoxaborole **1** (Scheme 1.1).⁹



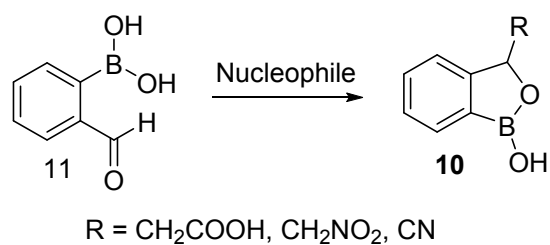
Scheme-1.1 Synthesis of benzoxaborole

Hydrolysis of 1,2-dihydro-1-hydroxy-2,3,1-benzodiazaborines **8** in presence of aqueous sodium hydroxide furnished corresponding benzoxaboroles **9** (Scheme 1.2).¹⁰



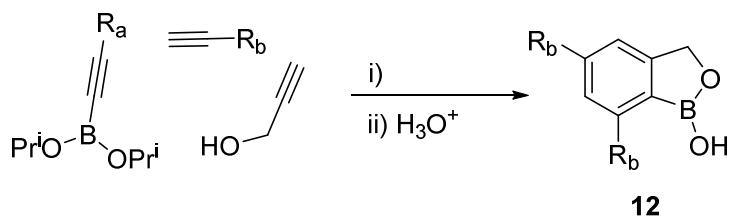
Scheme-1.2 Synthesis of substituted benzoxaborole

3-substituted benzoxaboroles **10** were obtained upon the treatment of o-formylphenylboronic acid **11** with nucleophiles such as malonic acid, nitromethane, sodium cyanide, etc. (**Scheme 1.3**).¹¹



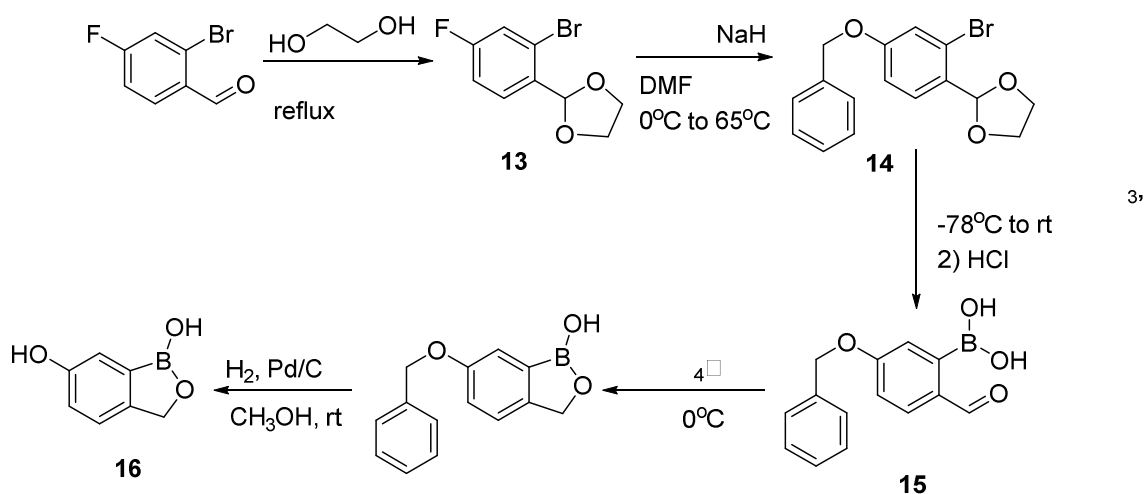
Scheme-1.3 Synthesis of 3-substituted benzoxaborole

Multicomponent coupling of alkynes in presence of a transition metal catalyst resulted in cyclotrimerization of alkynes furnishing substituted benzoxaboroles **12** (**Scheme 1.4**).¹²



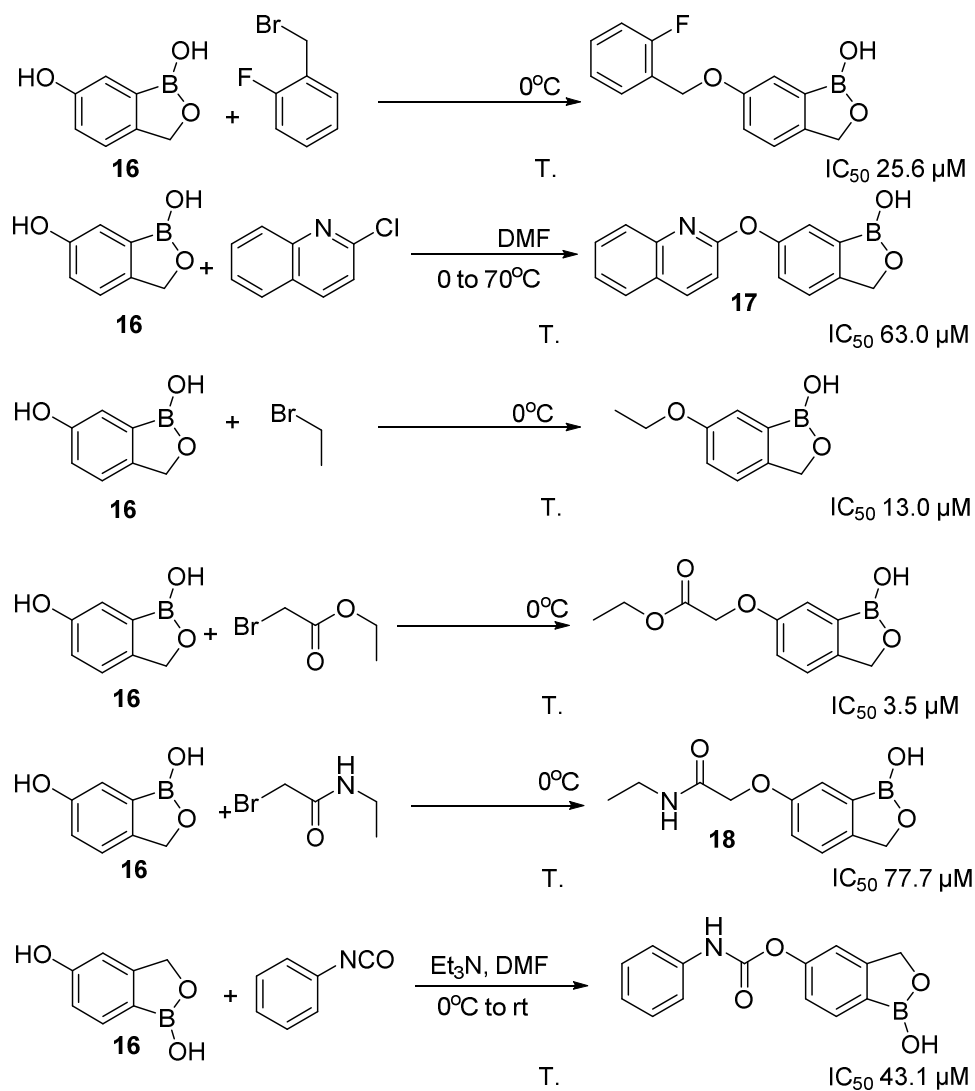
Scheme-1.4 Synthesis of benzoxaborole via multicomponent coupling of alkynes

Hydroxybenzboroxole **16** was synthesized and used as common intermediate in the preparation of various aryl ethers. These ethers have been evaluated as potential anti-Trypanosomal agents. The formyl group in 2-bromo-4-fluorobenzaldehyde was protected with ethylene glycol as its acetal **13**. Nucleophilic displacement of fluoride with benzyl alcohol furnished 2-(4-(benzyloxy)-2-bromophenyl)-1,3-dioxolane **14**. Reaction of **14** with *n*-butyl lithium followed by triisopropylborate, resulted in the formation of boronic acid, which upon acidification resulted in the deprotection of the acetal-protecting group to furnish the aldehyde **15**. Reduction of **15** with NaBH₄ yielded 6-(*O*-benzyl)-benzoxaborole, which was further reduced with Pd/C to obtain 6-hydroxybenzoxaborole **16** (Scheme 1.5).¹³



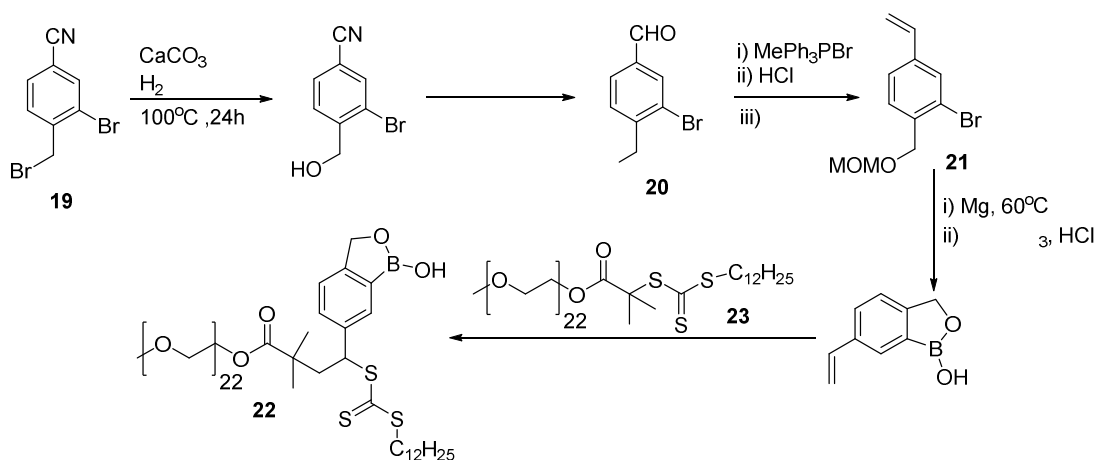
Scheme-1.5: Synthesis of benzoboroxoles as anti-trypanosomal agents

Hydroxybenzoxaborole **16** was coupled to corresponding benzyl bromides, alkyl bromides, or α -bromoacetates in presence of sodium hydride at 0°C to obtain the benzyl ethers, alkyl ethers and α -alkoxyacetates respectively. Diaryl ethers **17** were prepared from **16** upon reaction with NaH at 70°C. Treatment of **16** with α -bromoacetamides in the presence of K₂CO₃ with NaI as catalyst yielded α -bromoacetamides **18**. Carbamates were prepared from **16** by coupling with corresponding isocyanates in the presence of Et₃N in DMF at 0°C (**Scheme 1.6**).¹³



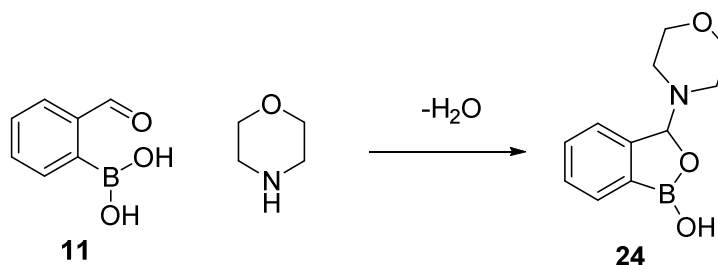
Scheme-1.6: Synthesis of benzoxaboroles as anti-trypanosomal agents

Kim *et. al.* reported the synthesis of polymers of benzoxaboroles as monosaccharide-responsive release of insulin from polymersomes of polyboroxole block copolymers at neutral pH (**Scheme 1.7**). The synthesis was initiated via the treatment of 3-bromo-4-(bromomethyl)benzonitrile **19** with calcium carbonate in presence of water-dioxane at 100°C to obtain 3-bromo-4-(hydroxymethyl) benzonitrile. The hydroxy group was protected by tert-butyltrimethylsilyl (TBDMS-Cl) group and the reduction of cyanide with DiBAL-H furnished the aldehyde **20**. Wittig olefination of **20** with methyltriphenylphosphonium bromide followed by the deprotection of TBDMS and re-protection using methoxymethyl (MOM) chloride furnished MOM ether **21**. The aryl bromide was converted into its Grignard by reacting with magnesium at 60°C and subsequently treated with trimethylborate to furnish the benzoxaborole monomer upon acidic work up in the presence of HCl. The monomer was further converted into the polymer by treating **22** with xanthate **23** (**Scheme 1.7**).¹⁴



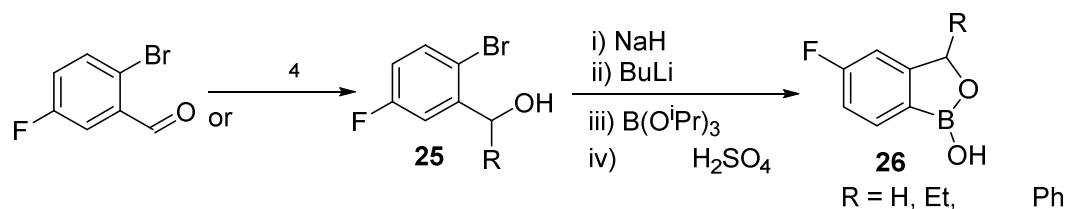
Scheme-1.7: Synthesis of polymeric benzoxaboroles

Morpholinobenzoxaborole **24** was obtained upon the reaction of *o*-formylphenylboronic acid **11** with morpholine (**Scheme 1.8**).¹⁵



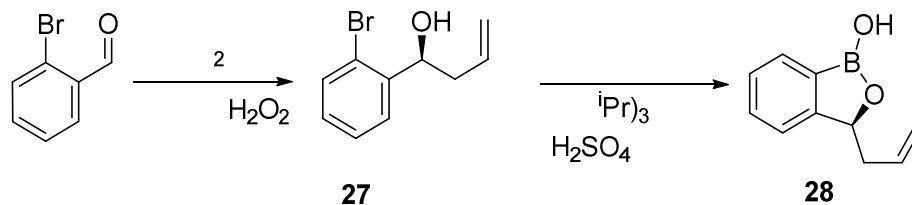
Scheme-1.8: Synthesis of 3-amino-substituted benzoxaboroles

2-Bromo-5-fluorobenzaldehyde was reacted with sodium borohydride or Grignard reagents to obtain substituted alcohols **25**. This was further treated with sodium hydride, followed by *n*-butyl lithium and triisopropylborate and 10% sulfuric acid to furnish corresponding 5-fluoro-3-substituted benzoxaboroles **26** (**Scheme 1.9**).¹⁶



Scheme-1.9: Practical synthesis of benzoxaboroles

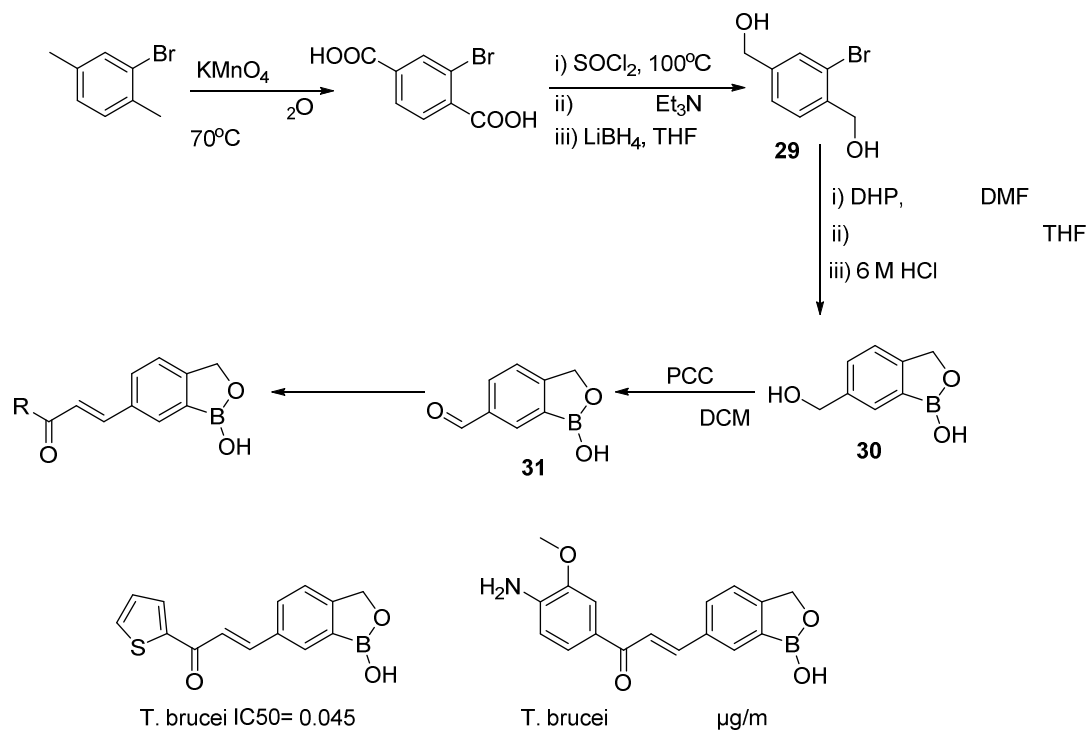
Similarly, chiral benzoxaboroles were prepared by the reaction of aldehydes with asymmetric allylating reagent such as Ipc_2BALLy . 2-bromobenzaldehyde was treated with Ipc_2BALL to obtain (S)-1-(2-bromophenyl)but-3-en-1-ol (**27**), which was cyclized to (S)-3-allylbenzo[*c*][1,2]oxaborol-1(3H)-ol (**28**) upon reaction with sodium hydride, *n*-butyl lithium, triisopropylborate, and sulfuric acid (**Scheme 1.10**).¹⁶



Scheme-1.10: Practical synthesis of chiral benzoxaboroles

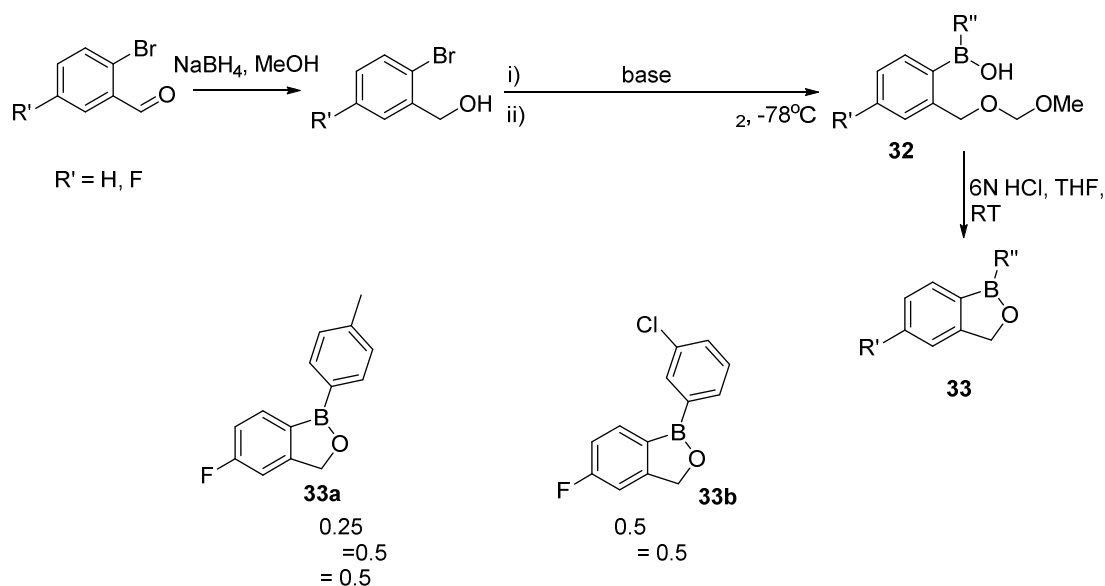
Dimethylbromobenzene was oxidized to dicarboxylic acid using KMnO_4 in tertiary butanol and water. Conversion of dicarboxylic acid into the corresponding acid chloride was achieved using thionyl chloride, which upon reaction with methanol in triethylamine resulted in the formation of the dimethyl ester. Reduction of the diester with lithium borohydride furnished the diol **29**. Protection of the alcohol groups as THP acetal followed by the treatment with *n*-butyl lithium and triisopropylborate, and subsequent deprotection and cyclization in 6 M HCl furnished 5-hydroxymethylbenzoxaborole **30**. This was further oxidized with PCC to obtain 5-formyl benzoxaborole **31**. Chalcone–benzoxaborole hybrid molecules were prepared from this aldehyde **31** by aldol condensation with substituted methylketones in presence of sodium hydroxide in ethanol-water. Some of these

molecules showed excellent anti-trypanosomal activity against *T. brucei* with IC₅₀ values ranging 0.045 µg/mL (**Scheme 1.11**).¹⁷



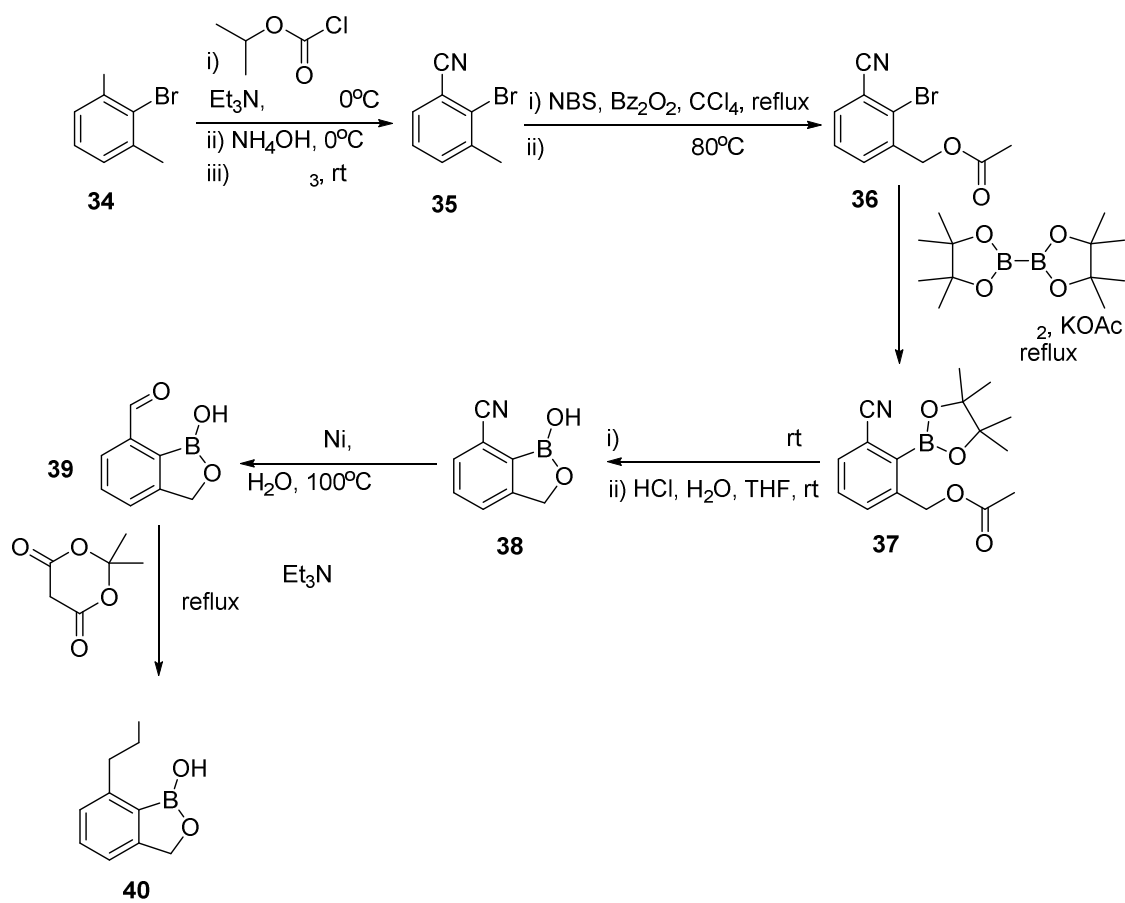
Scheme-1.11: Chalcone–benzoxaborole hybrid molecules as potent anti-trypanosomal agents

2-bromo-5-fluorobenzaldehyde was reduced to alcohol with sodium borohydride in methanol. The hydroxy group was protected as the methoxymethyl ether (MOM) and treated with butyllithium at -78°C , in presence of B-phenylboronic acid ethylene glycol ester to obtain the borinic acid **32**. The protecting group was removed under acidic conditions and the free alcohol spontaneously cyclized to give the target compound **33**.^{6c} Some of these derivatives showed antifungal properties on *C. albicans* at a conc of $0.25\ \mu\text{g}/\text{mL}$ and *C. neoformans* at $0.5\ \mu\text{g}/\text{mL}$ (**Scheme 1.12**).



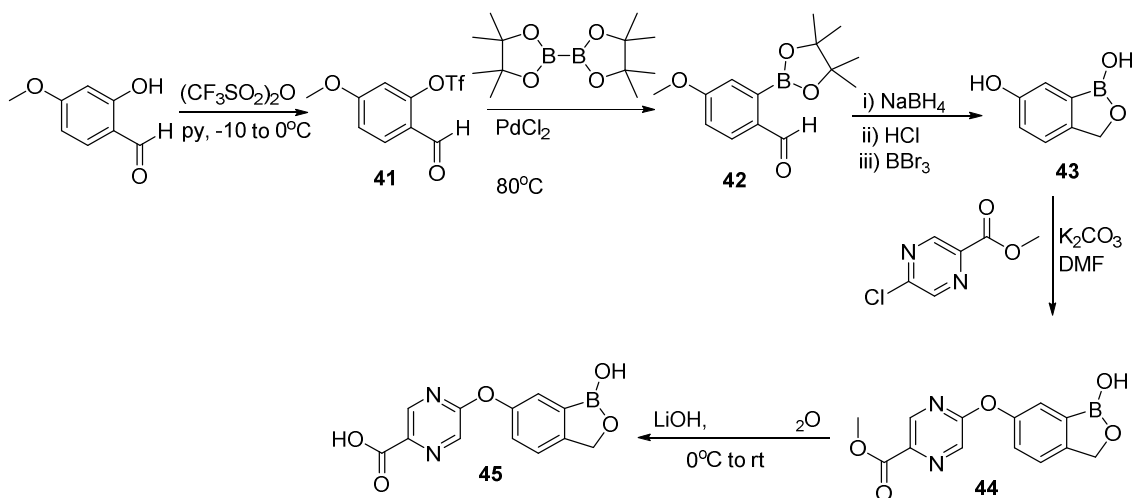
Scheme-1.12: Synthesis of AN2690 and its analogs for the potential treatment of onychomycosis

2-Bromo-3-methylbenzoic acid **34** was converted to cyano compound **35** by amide formation followed by dehydration. Benzylic bromination on the methyl group with NBS followed by nucleophilic substitution with potassium acetate afforded the acetate **36** that upon catalytic borylation with bispinacolatodiboron in the presence of Pd catalyst yielded the arylboronate **37**. Alkaline hydrolysis of the acetate followed by acidification generated the cyano benzoboroxole **38**. Treatment of **38** with Raney nickel in aqueous formic acid gave the aldehyde **39**. The target compound **40** was obtained by reacting the aldehyde with 2,2-dimethyl-1,3-dioxane-4,6-dione in the presence of HCOOH and triethylamine.¹⁸ Some of these compounds were active against malaria parasite Plasmodium falciparum with IC₅₀ values in the range of 0.026 μM (**Scheme 1.13**).



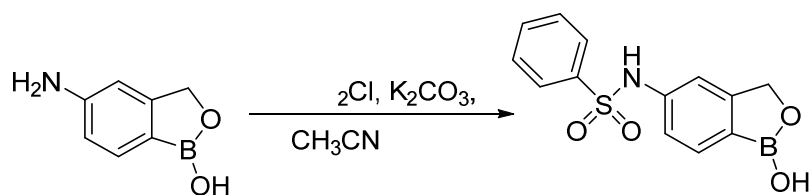
Scheme-1.13: Synthesis and structure–activity relationships of novel benzoxaboroles as a new class of antimalarial agents

2-Hydroxy-4-methoxybenzaldehyde was converted to triflate **41** via treatment with triflic anhydride. Catalytic borylation with bispinacolatodiboron provided compound **42**, which was reduced with NaBH₄ in methanol-THF and upon acid-catalyzed cyclization, was converted to 6-methoxy benzoxaborole **43**. Demethylation of **43** with BBr₃ followed by treatment with methyl 5-chloropyrazine-2-carboxylate in potassium carbonate and DMF afforded the ester **44**, which was further hydrolyzed in basic conditions to furnish the corresponding carboxylic acid **45**.¹⁹ Some of these compounds showed activity on *E. cloacae* P99AmpC (MIC = 1–2 µg/mL) and *E. coli* SYN2549 CMY-2 (MIC = <0.5 µg/mL) (Scheme 1.14).

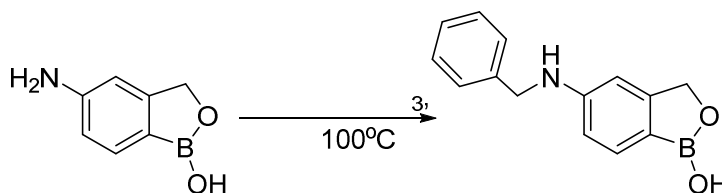


Scheme-1.14: Synthesis and SAR of novel benzoxaboroles as a new class of β -lactamase inhibitors

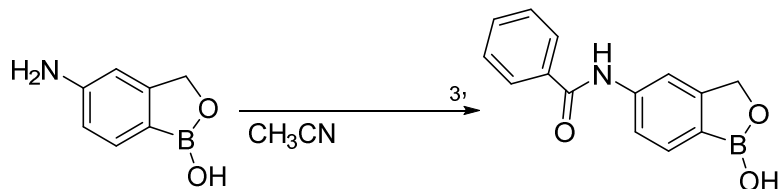
Coupling of 5-amino-benzoxaborole with phenylsulfonylchloride, benzyl bromide and benzoyl chloride provided corresponding sulfonamide, aminomethylene and amide respectively and these compounds showed growth inhibition in *T. brucei* with IC_{50} values ranging between 0.04-1.2 $\mu\text{g/mL}$ (**Scheme 1.15**).²⁰



T. brucei Growth Inhibition $IC_{50} = 0.02 \mu\text{g/mL}$



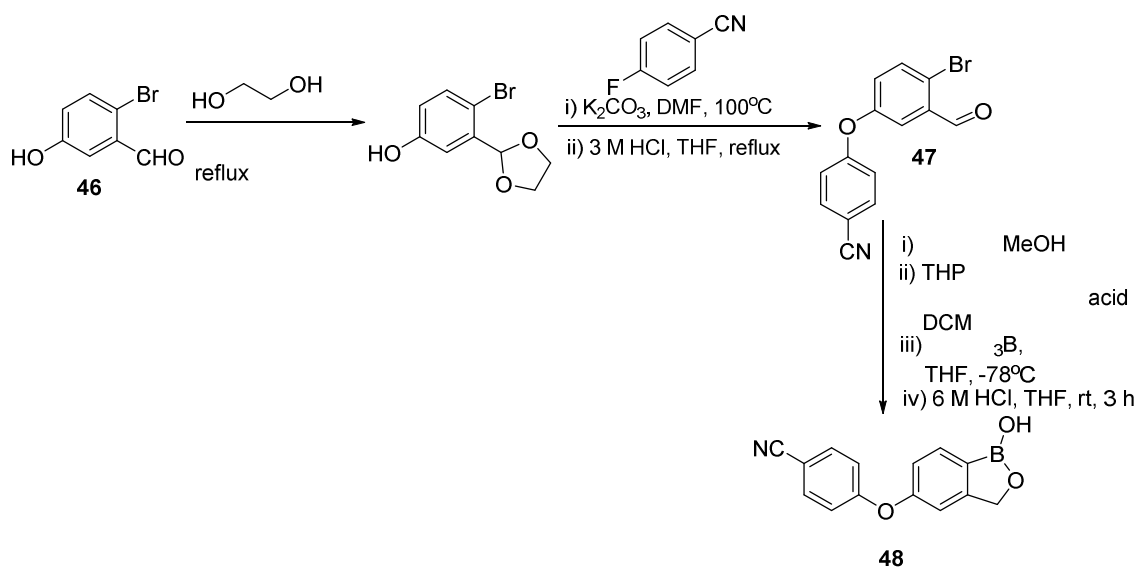
T. brucei Growth Inhibition $IC_{50} = 1.21 \mu\text{g/mL}$



T. brucei Growth Inhibition $IC_{50} = 0.04 \mu\text{g/mL}$

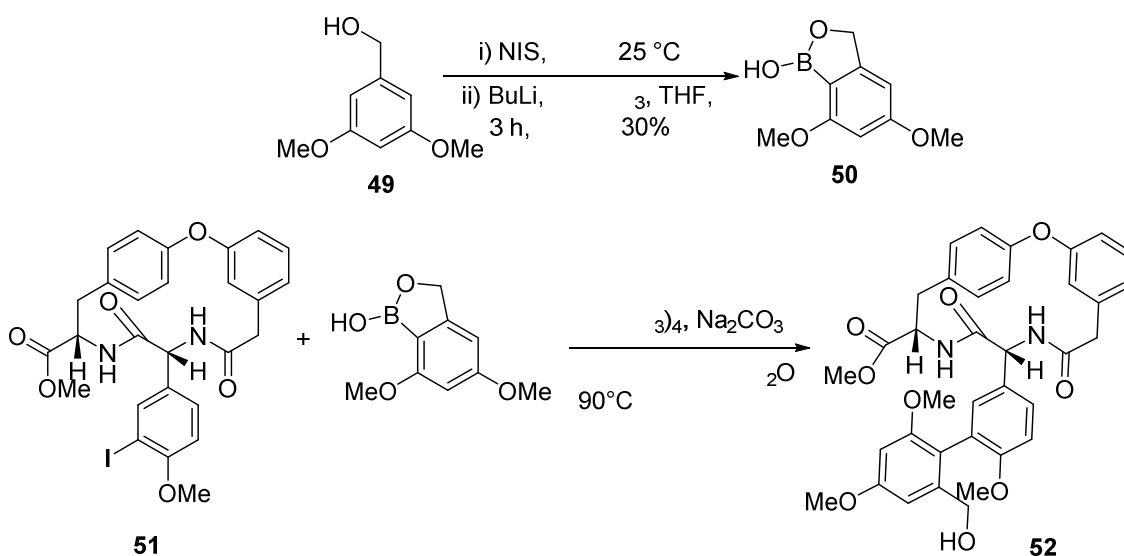
Scheme-1.15: Discovery of Novel Benzoxaborole-Based Potent Antitrypanosomal Agent

The aldehyde group of 2-bromo-5-hydroxybenzaldehyde **46** was protected with ethyleneglycol and treated with 4-fluorobenzonitrile to form 4-(4-bromo-3-formylphenoxy) benzonitrile **47**. Aldehyde **47** was reduced to alcohol using sodium borohydride and protected as a THP ether in presence of 3,4-dihydro-2H-pyran and camphorsulfonic acid. The boron atom was then introduced by halogen–metal exchange with n-butyl lithium using triisopropylborate, which upon reaction with HCl led to the deprotection of the THP acetal and the resultant alcohol spontaneously cyclized to afford 4-((1-hydroxy-1,3-dihydrobenzo[c][1,2]oxaborol-5-yl)oxy)benzonitrile **48** (**Scheme 1.16**).²¹ This molecule showed activity against PDE4 (IC₅₀ 0.49 μM) and TNF-α (IC₅₀ 0.54 μM).



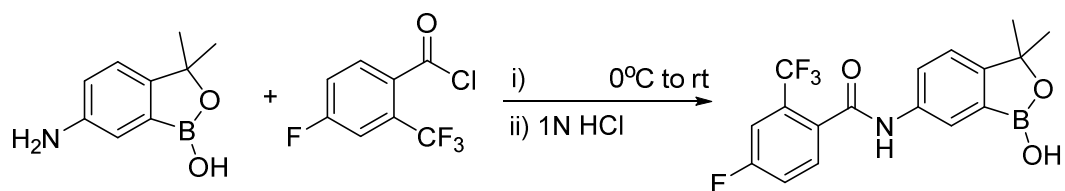
Scheme-1.16: Synthesis of anti-inflammatory agent (AN2728) for the potential topical treatment of psoriasis and atopic dermatitis

3,5-dimethoxybenzyl alcohol **49** was converted to corresponding benzoxaborole **50** by *N*-iodosuccinimide iodination followed by reaction with *n*-butyllithium and trimethoxyborate. Suzuki coupling of aryl iodide **51** with benzoxaborole **52** was facilitated by Pd(PPh₃)₄ catalyst in presence of sodium carbonate in methanol-toluene-water system at 90°C (**Scheme 1.17**).²²



Scheme-1.17: Suzuki coupling–macrolactamization approach to the AB-COD bicyclic system of vancomycin

6-amino-3,3-dimethyl-3H-benzo[c][1,2]oxaborol-1-ol acetate salt was treated with 2-trifluoromethyl-4-fluorobenzoyl chloride in presence of trimethylamine in DCM at 0°C to afford SCYX-7158 (**53**) (**Scheme 1.18**), which showed activity against *Trypanosoma brucei rhodesiense* (IC_{50} = 0.294 μ g/mL).²³



Scheme-1.18: Synthesis of amidobenzoxaboroles

Owing to the importance of cyclic boronic acids, we undertook the project involving the development of novel methodologies for the synthesis of benzoxaboroles containing wide array of functional groups. Our results are reported in the succeeding chapters.

Chapter 2: Novel Methodologies for the Synthesis of Functionalized Benzoxaboroles

Introduction

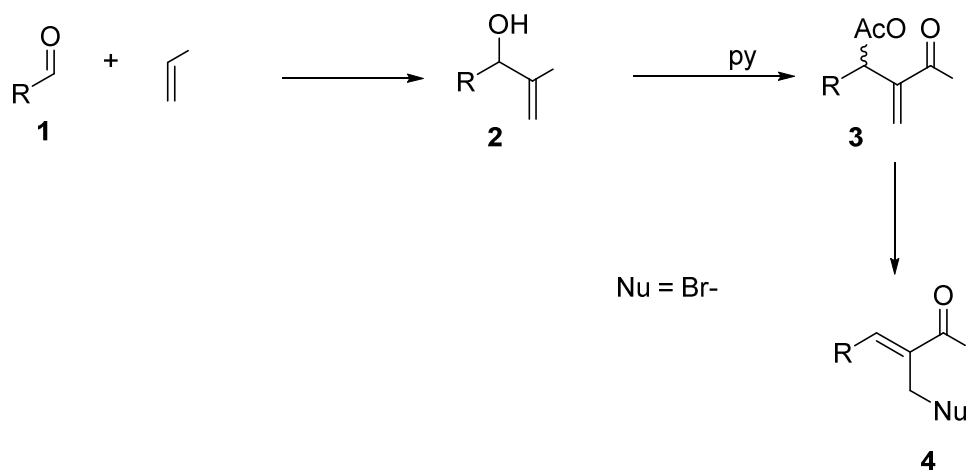
Being a relatively new compound, benzoxaborole has been utilized in synthetic methods and applications, but most of them involve tedious synthesis and in almost all the cases, boron has been introduced in the very last step. Apart from designing novel methods for the synthesis of benzoxaborole, we undertook the project due to its impressive application regime in medicinal chemistry²⁴. In this chapter, we describe the development of three convenient, flexible and novel protocols for the synthesis of densely functionalized benzoxaboroles based on Baylis-Hillman chemistry, Barbier allylation, and aldol reaction protocols. In all the cases, we used 2-formyl boroaldehyde, a commercial available compound as a synthon.

Results and Discussion

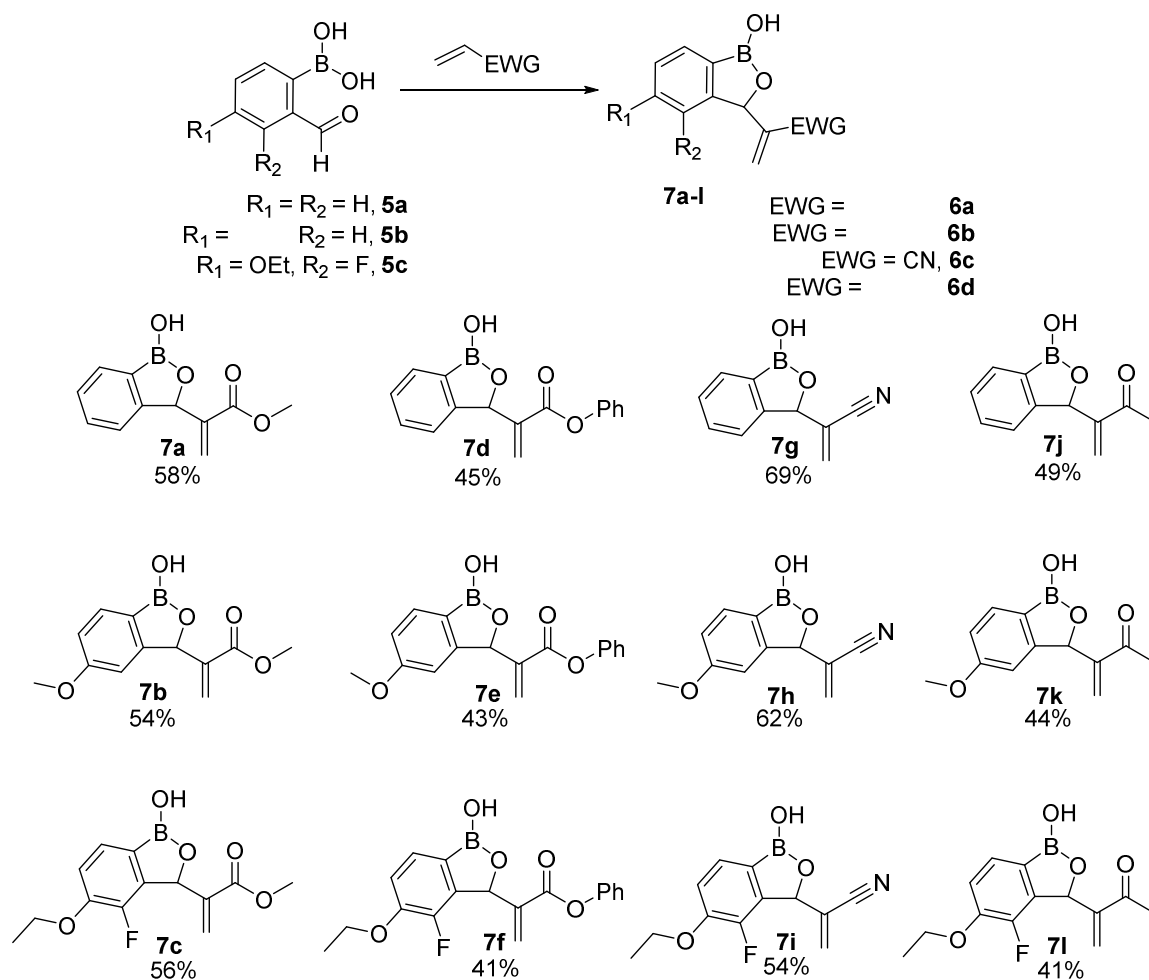
Baylis-Hillman (BH) reaction is one of the best reactions in introducing C-C bond coupling in one step²⁵. This reaction is atom efficient and produces activated allyl alcohols which are highly useful in the isomerization upon reaction with several nucleophiles affording valuable synthons. BH reaction usually involves an aldehyde and electron deficient olefin in the presence of hindered tertiary amine like 1,4-diazabicyclo[2.2.2]octane (DABCO). These reactions usually don't require any solvent. In this case, we first utilized formylboronic acids **5a-c** as electrophiles and methyl acrylate **6a**, phenyl acrylate **6b**, acrylonitrile **6c**, methyl vinyl ketone **6d**, and acrolein **8** as electron deficient olefins. Aldehyde **5a** when reacted with 2 parts of methyl acrylate in presence of 10% DABCO resulted in poor yields of the desired product, leaving most of the unreacted starting material. The yield did not improve significantly even when doubling the amount of the catalyst DABCO. But, with stoichiometric amount of DABCO, compound **7a** was obtained with 58% yield after purification via silica gel column chromatography.

The above reaction with simple benzaldehyde usually requires more than a week for the completion. Similar reaction with 4-boronobenzaldehyde resulted in meagre yield even after two weeks. The increased rate of reactivity with 2-boronobenzaldehyde **5a** could be attributed to the activation of carbonyl oxygen via coordination with proximal Lewis acidic boronic acid group.

The other reactions with **5b-c** with methyl acrylate was even slower (5-7 days) because of the alkoxy substituents on the aromatic ring affording the products **7b-c** in 54% and 56% yields respectively. Similar reactions of **5a-c** with phenyl acrylate **6b** yielded 41-45% of corresponding benzoxaboroles **7d-f** (**Scheme 2.1**).



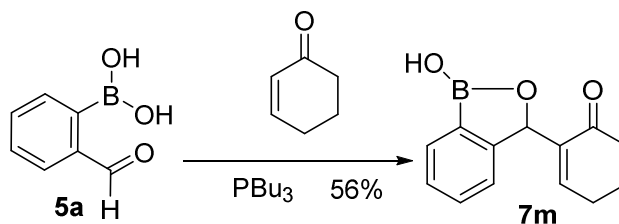
Scheme 2.0: Baylis-Hillman Reaction



Scheme 2.1. Baylis Hillman reaction of β -Boronoaldehydes

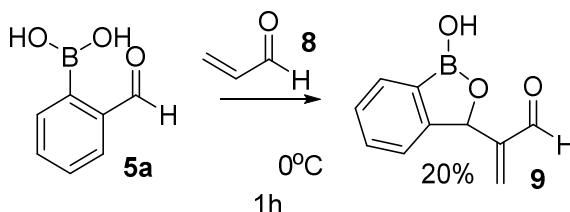
The similar BH reaction with better electron withdrawing group acrylonitrile **6c** was facile with all the boronoaldehydes **5a-c** and the products **7g-i** were obtained in moderate to good yields. Introduction of even stronger electron withdrawing groups resulted in poor yields of **7j-l** (41-49%) due to the higher polymerization rate of methyl vinyl ketone **6d** (Scheme 2.1).

We also carried out BH reaction with the aldehyde **5a** and cyclohexenone in presence of tributylphosphene to obtain **7m** in 56% yield (**Scheme 2.2**).



Scheme 2.2: Preparation of α -cyclohexenonyl benzoxaborole

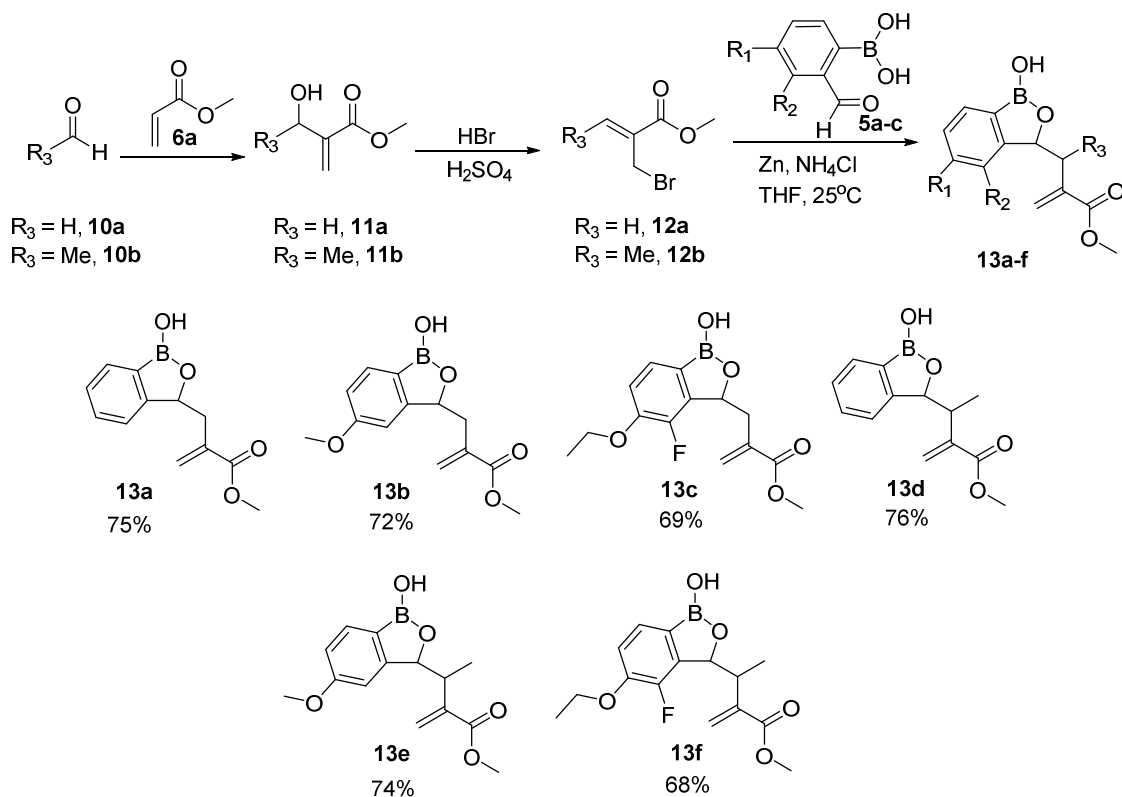
We also utilized acrolein **8**, with boronobenzaldehyde **5a** which resulted in **9** with only 20% yield with 60% of unreacted starting material. The low yield of **9** is due to the fact that acrolein is very sensitive monomer and susceptible for rapid polymerization. Longer reaction time or excessive acrolein resulted in polymerization of the acrolein and did not improve the yields. Hence the reaction was conducted in THF solvent at 0°C to minimize the polymerization. (**Scheme 2.3**).



Scheme 2.3. Reaction of β -Boronoaldehydes with Acrolein

We have utilized Barbier type allylation, another C-C bond forming reaction involving allyl bromides and aldehydes in the presence of metals like

zinc and indium forming a complex with ammonium chloride to synthesize functionalized benzoboroxoles. We synthesized the BH derived bromides **12a-b** by treating the corresponding aldehydes **10a** and **10b** with methyl acrylate **6a** to form allylic alcohols **11a-b**, followed by the reaction with HBr-H₂SO₄. These bromides were then utilized for allylation of boronobenzaldehydes **5a-c** using Barbier allylation conditions to obtain homologated benzoxaboroles **13a-f** in decent yields (**Scheme 2.4**). As expected, products had predominantly *syn* stereochemistry. The relative configuration was confirmed by single crystal X-ray analysis of **13d** (**Figure 2**).



Scheme 2.4. Synthesis of Homologated Benzoxaboroles

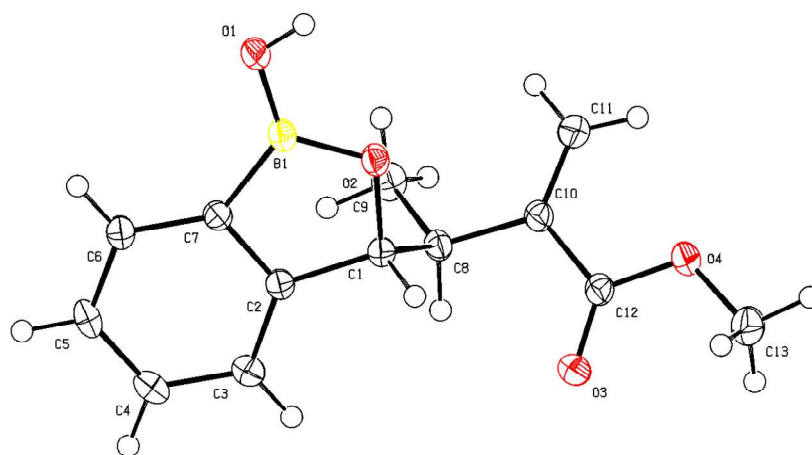
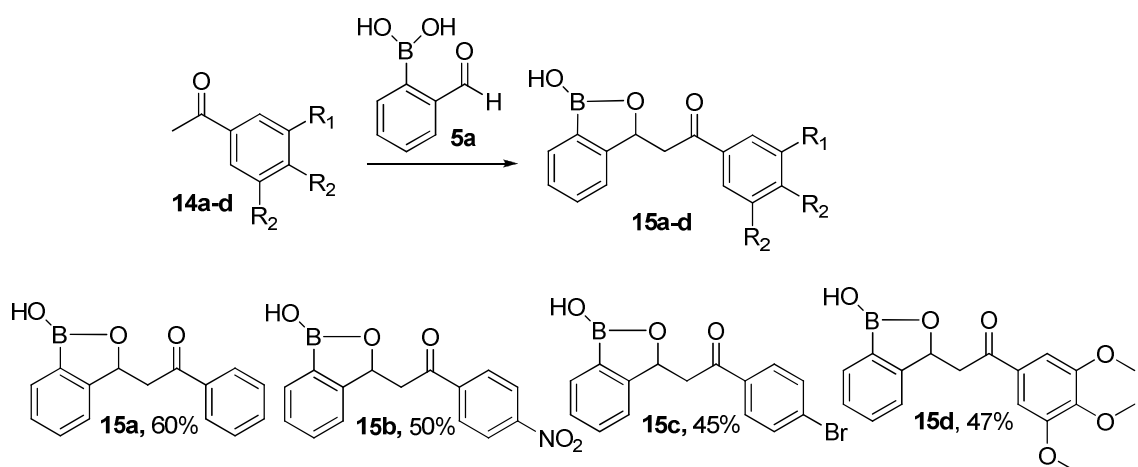


Figure 2. X-ray Crystal Structure of **13d**

We have also utilized aldol reaction with boronobenzaldehyde **5a** and methyl ketones **14a-d** to obtain β -ketobenzoxaboroles **15a-d**. With bases such as sodium hydride, sodium hydroxide, etc., the reaction did not go to completion and the product was impure. When LDA is used at -78°C to generate enolate, the reaction resulted in aldol product **15a** in 60% yield (brsm) (**Scheme 2.5**). Other substituted acetophenones such as *p*-nitroacetophenone, *p*-bromoacetophenone, and 3,4,5-trimethoxyacetophenone upon reaction with 2-boronobenzaldehyde resulted in corresponding products **15b-d**. Irrespective of the amount of base used, the reaction was still incomplete with ~30-50% of starting material left. Benzoxaboroles **15e-g** were obtained upon similar reaction using acetone, dimethyl and diethyl malonates with boronobenzaldehyde **6a** which resulted in 40-62% yields (**Figure 3**).



Scheme 2.5: Synthesis of β -ketobenzoxaboroles

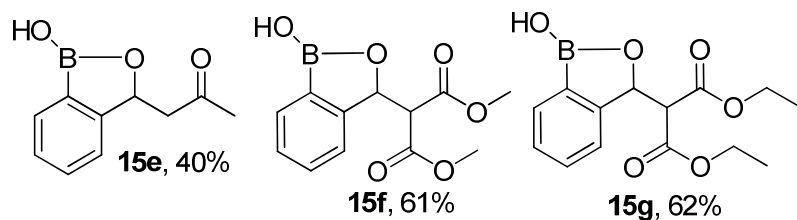


Figure 3: β -Ketobenzoxaboroles via aldol reaction

All the synthesized compounds were tested for their antimicrobial efficacy as well as general cytotoxicity. None of the compounds exhibited any biological activity. This concludes that the substitution of carbon on the benzylic carbon of benzoxaborole lead to the loss of activity.

Experimental Procedures

1. Representative procedure for the preparation of benzoxaboroles via

Baylis-Hillman reaction 7a-l: To a stirred suspension of 2-boronobenzaldehyde **5a** (0.3 g, 2.0 mmol) and methyl acrylate **6a** (0.7 mL, 8.0 mmol) was added DABCO (224 mg, 2.0 mmol) and stirred for 2 days at room temperature. Upon completion (TLC), the reaction mixture was quenched with dilute HCl and worked up with ethyl acetate (3 x 10 mL). The combined organic layers were dried (MgSO₄), concentrated *in vacuo*, and purified via silica gel column chromatography (hexane:acetone, 3:1) to obtain 0.25 g (58%) of benzoboroxole **7a** as a white powder. *mp*: 91-93°C, (Found: C, 60.62%; H, 5.25%; C₁₂H₁₃BO₄ requires: C, 60.60%; H, 5.09%); ¹H NMR (400 MHz, CDCl₃): δ 9.45 (s, 1H), 7.70-7.72 (m, 1H), 7.40-7.44 (m, 1H), 7.25-7.35 (m, 2H), 6.16 (s, 1H), 5.93 (s, 1H), 5.79-5.80 (m, 1H), 3.67 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 166.2, 155.5, 140.6, 131.5, 131.2, 128.1, 126.6, 122.3, 79.3, 52.5; ESI-MS: 217 [(M-H)⁺, 100%].

2. Representative procedure for the preparation of benzoxaboroles via

Barbier reaction 13a-f: To a stirred solution of 2-boronobenzaldehyde **5a** (0.3 g, 2.0 mmol) and zinc (0.19 g, 3.0 mmol), in THF (5.0 mL) was added methyl α-bromomethylacrylate **12a** (0.72 g, 4.0 mmol), and saturated NH₄Cl (1 mL) and stirred overnight at room temperature. Upon completion (TLC), the reaction mixture was filtered over celite, and worked up with water and ethyl acetate (3 x 10 mL). The combined organic layers were dried (MgSO₄), concentrated *in*

vacuo, and purified via silica gel column chromatography (hexane:acetone, 3:1) to obtain 0.35 g (75%) of benzoboroxole **13a** as a viscous liquid. (Found: C, 62.01%; H, 5.52%; C₁₂H₁₃BO₄ requires: C, 62.11%; H, 5.65%); ¹H NMR (400 MHz, CDCl₃): δ 9.24 (s, 1H), 7.67 (d, *J* = 7.2 Hz, 1H), 7.41-7.45 (m, 1H), 7.29-7.36 (m, 2H), 6.12 (d, *J* = 1.2 Hz, 1H), 5.69 (d, *J* = 1.2 Hz, 1H), 5.26 (dd, *J* = 4.0, 8.4 Hz, 1H), 3.64 (s, 3H), 2.89 (dd, *J* = 8.4, 14.4 Hz, 1H), 2.39 (dd, *J* = 8.4, 14.8 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 167.6, 156.8, 136.9, 131.2, 131.1, 128.2, 127.9, 122.2, 79.1, 52.5, 39.2; ESI-MS: 231 (M-H)⁺, 189 (100%).

3. Representative procedure for the preparation of benzoxaboroles via aldol reaction 15a: To a stirred solution of acetophenone **14a** (10 mmol) in 20 mL THF was added LDA (12 mmol) at -78°C and stirred for 1 h. A solution of 2-boronobenzaldehyde **5a** (9 mmol) in 5 mL THF was added to the reaction at -78°C, and stirred overnight at room temperature. Upon completion (TLC), the reaction was quenched with NH₄Cl and worked up with ethyl acetate. The combined organic layers were dried (MgSO₄), concentrated *in vacuo*, and purified by silica gel column chromatography to obtain the pure benzoxaborole **15a** in 60% yield. ¹H NMR (500 MHz, DMSO-d₆): δ 9.21 (s, 1H), 8.03 (d, *J* = 8.0 Hz, 2H), 7.74 (d, *J* = 7.5 Hz, 1H), 7.65-7.68 (m, 1H), 7.48-7.57 (m, 4H), 7.37-7.40 (m, 1H), 5.71 (dd, *J* = 4.0, 8.8 Hz, 1H), 3.58 (dd, *J* = 3.5 Hz, 17.5 Hz, 1H), 3.34 (dd, *J* = 8.5 Hz, 17.0 Hz, 1H); ¹³C NMR (125 MHz, CDCl₃): 198.6, 157.5, 137.7, 134.3, 131.6, 131.4, 129.7, 129.2, 128.2, 122.5, 77.7, 46.3; ESI-MS: 251 [(M-H)⁺, 100%].

Antimicrobial Assay

Due to their importance as antimicrobial agents, the synthesized molecules have been evaluated for their activity as antibacterial and antifungal agents. For initial testing, *Pseudomonas aeruginosa* and *Streptococcus thermophiles* have been selected as gram negative and gram positive bacterial strains respectively. For antifungal activity, we chose *Candida albicans* and *Aspergillus niger*. Standard Kirby-Bauer disk susceptibility method was used for all the species. Mueller Hinton agar plates were used to grow the colonies of microbes. The microbial suspension was inoculated onto the agar plates and the sterile disc saturated with 20 μL test compound was placed on the plate. The final concentration of the compound was maintained at 5 and 1 $\mu\text{g}/\text{disc}$. These plates were incubated at 37° C for 24-48 hours. The diameter of the zone of inhibition was measured in millimeters using calipers to determine the antimicrobial activity. None of the synthesized compounds showed any antimicrobial activity compared to compound **1**, which exhibited moderate activity.

Cytotoxicity Assay

These compounds were also tested for their toxicity on MCF-7 breast cancer cell line using sulforhodamine-B assay. Briefly, 5×10^4 cells/mL were plated in a 48-well plate and incubated for 18-24 hours. The test compounds were dissolved in DMSO and added to the wells and incubated for 3 more days. The growth media

was removed on the third day and the wells were washed with 1% phosphate buffered saline solution. The wells were dried and the SRB in 1% acetic acid was added. The plate was incubated for ~45 minutes and the wells were dried and dissolve in tris base (10mM, pH 10.2). The absorbance was recorded at 540nm, which gives the % survival of the cells. The assay was carried out in triplicates. All the compounds were tested at 100 and 50 μ M. None of the synthesized compounds were toxic even at 100 μ M concentration. This concludes that these molecules are generally non-toxic in nature.

Conclusions

In conclusion, we have successfully devised three methodologies using boronobenzaldehyde to synthesize functionalized benzoboroxoles via Baylis-Hillman, Barbier-type allylation and Aldol chemistry. These protocols can be applied in the synthesis of various benzoboroxoles in large scale. The synthesized molecules were evaluated for their antimicrobial efficacy. These methodologies can be used to develop synthons for various synthetic products.

Chapter 3: Boric Acid Mediated Synthesis of α -Hydroxyamides and Synthesis of Benzoxaboroles via Passerini Reaction

Introduction

Boric acid, usually available in nature in the form of borax and boracite. It is used for industrial as well as household purposes due to its non-toxic nature and it is very inexpensive. Hence, it is usually considered as a green material²⁶. Various borates and organoboranes can be synthesized starting from boric acid. Due to the vacant orbital, it can be used as a mild Lewis acid catalyst in several organic reactions²⁷.

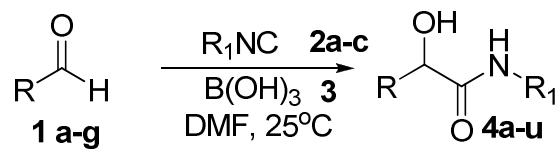
Passerini and Ugi reactions are well-studied multicomponent coupling reactions involving aldehyde/imine, acid and isonitrile²⁸. α -Hydroxyamides can be easily oxidized to prepare keto acids/amides which are biologically relevant since pyruvic acid is one of the important metabolites in glycolysis. Hence, α -Hydroxyamides are important synthons for the synthesis of various agents in medicinal chemistry²⁹. In this chapter, we have developed a novel methodology for the synthesis of several α -hydroxyamides using boric acid as a catalyst for the addition of isonitriles on to carbonyl compounds³⁰.

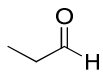
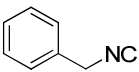
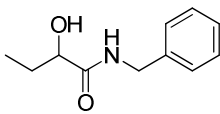
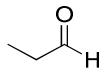

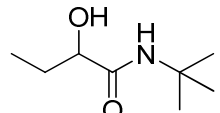
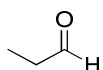
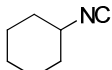
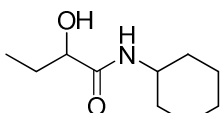
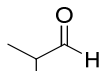
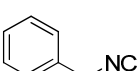
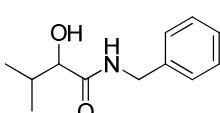
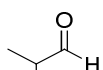

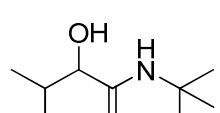
Results and Discussion

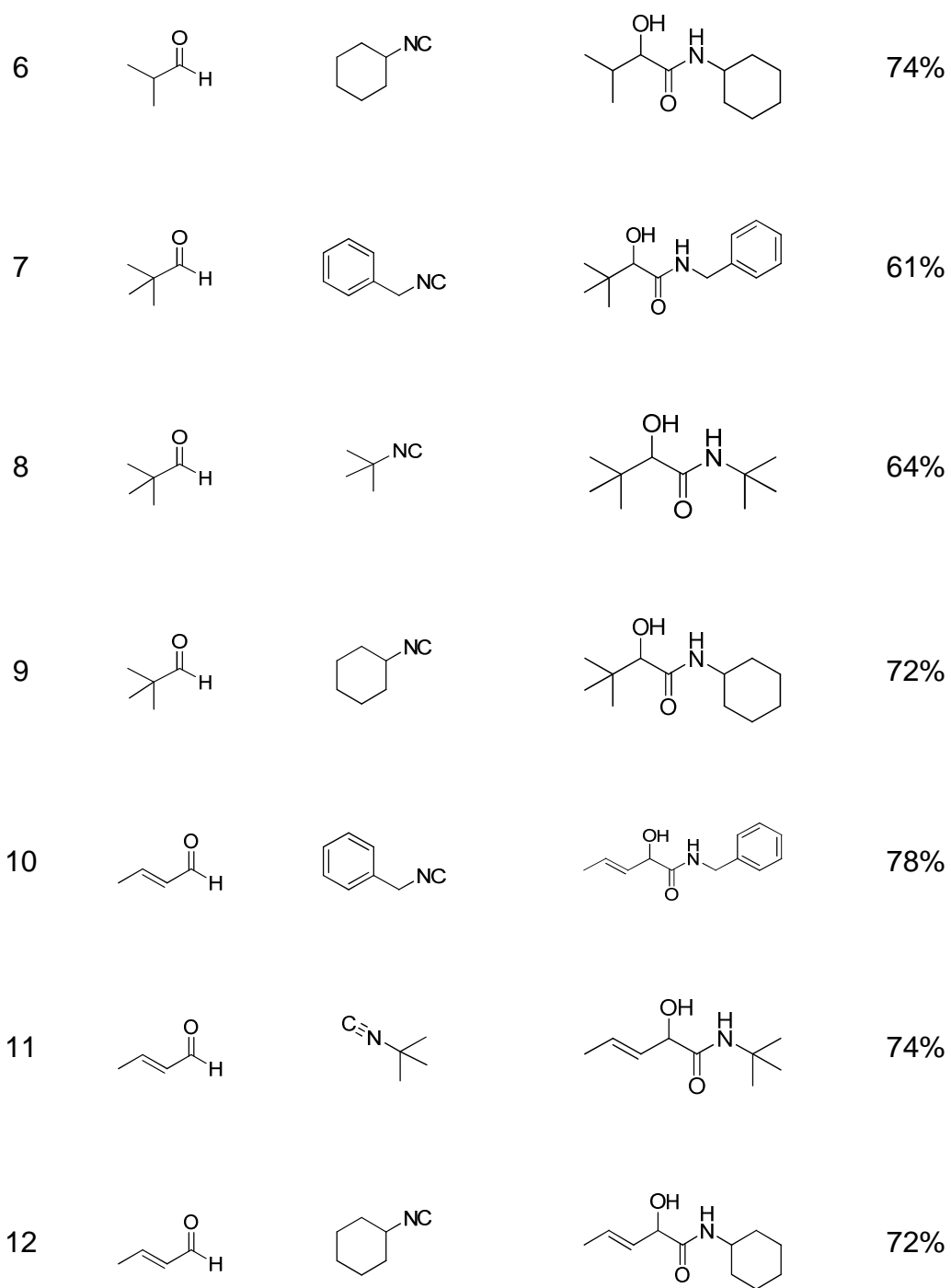
We envisioned the usage of boric acid to couple isonitriles and aldehydes to synthesize α -hydroxyamides. For this purpose, benzyl, cyclohexyl and tert-butyl isonitriles **2a-c** were taken as representative examples for isonitriles. Aldehydes **1a-g** containing aliphatic, aromatic and heterocycles were utilized. For initial purposes, propionaldehyde and benzyl isonitrile were utilized for the optimization. Solvent study was conducted to figure out the suitable solvent for the reaction with maximum yield. Different solvents such as THF, CH₂Cl₂, EtOAc, DMF, DMSO, Dioxane, MeOH and water were used. The amount of boric acid was varied in each reaction to obtain decent yield. DMF and 1 equivalent of boric acid were found to provide α -hydroxyamides with good yields. The reaction with aliphatic aldehydes and aliphatic isonitriles was found to be fast with the generation of small amount of heat whereas the reactions with aromatic systems tended to be relatively slow.

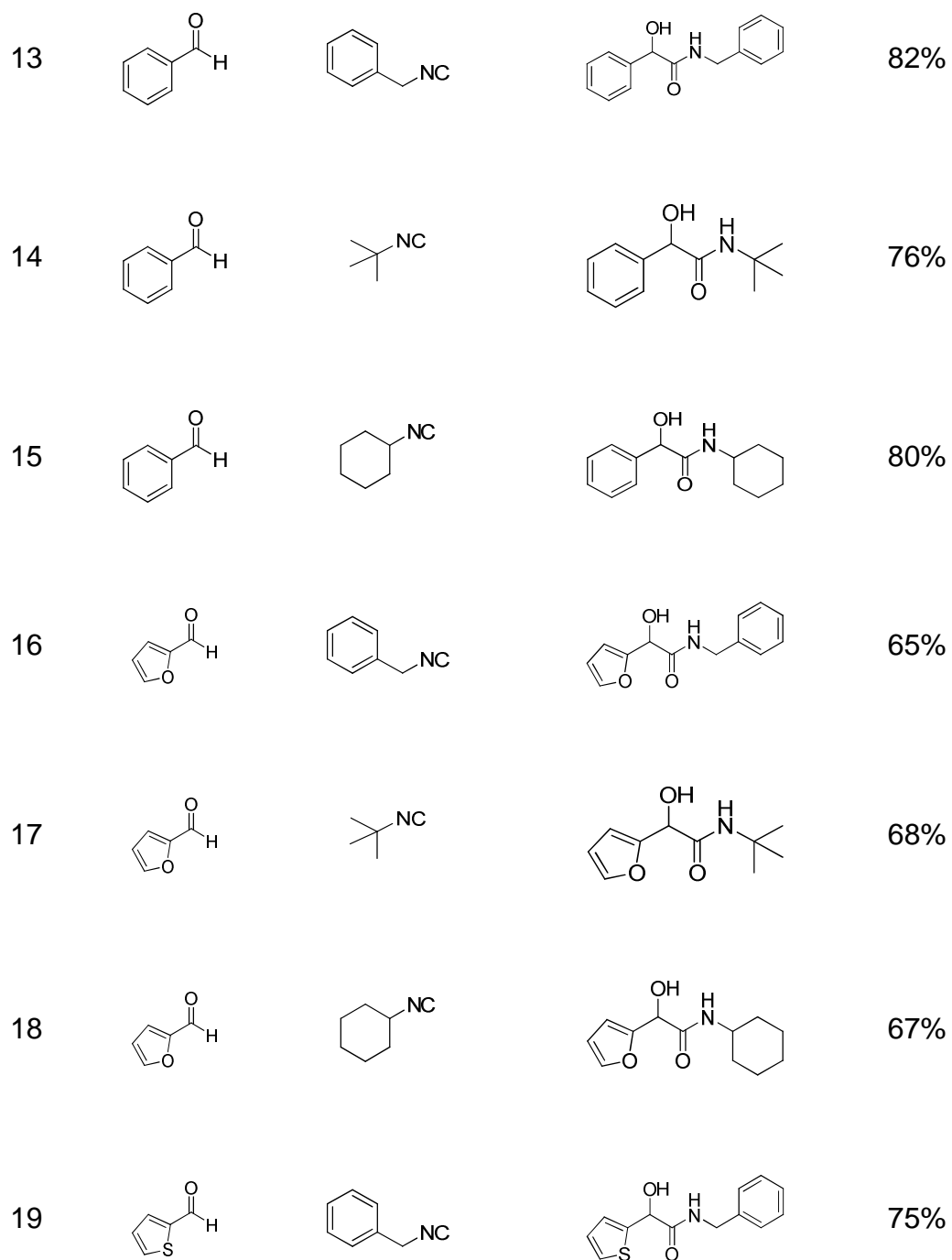
Similar reactions were performed replacing the aldehyde with ketones such as 3-pentanone and acetophenone. In this case, the reactions did not produce the product in acceptable yields even when the boric acid is increased to two equivalents at 80° C.

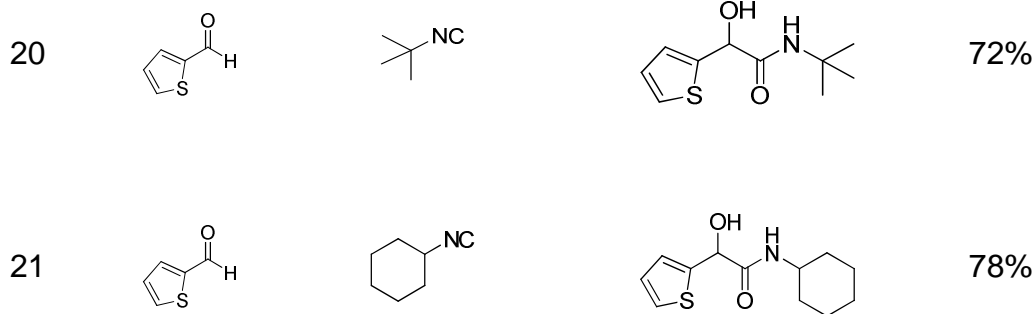
Table 1. Boric acid mediated addition of isonitriles and aldehydes for the synthesis of α -hydroxyamides.



Entry	Aldehyde	Isonitrile	Product	Yield
1				78%
2				74%
3				75%
4				70%
5				71%

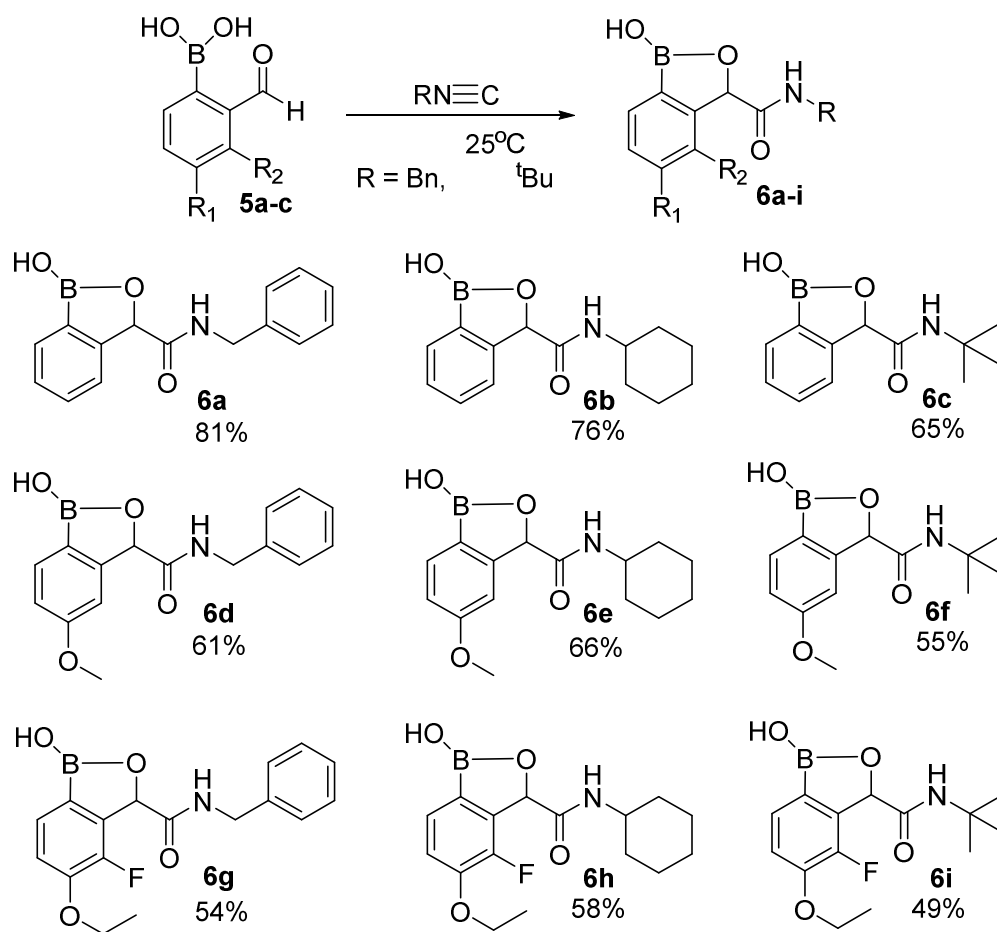






We then explored this method by replacing simple aldehydes with 2-formyl phenylboronic acid **5**. Due to the presence of borinic acid group ortho to aldehyde, we expected that **5** could self-catalyze the reaction with isocyanide. To our surprise, the reaction progressed and benzoxaborole was formed in 84% yield by dehydration. This was evident by the characteristic peak of B-OH at ~9ppm in $^1\text{H-NMR}$.

In a similar way, benzoboroxoles **6b** and **6c** were obtained upon reaction of **5** with isocyanides **2b** and **2c** respectively (**Scheme 3.1**). Also, products **6a-l** were synthesized by the reaction of aldehydes **5a-c** with isocyanides **2a-c** (**Scheme 3.1**).³¹



Scheme 3.1: Synthesis of functionalized benzoxaboroles.

The above synthesized structural entities can be utilized in the applications of material chemistry as well as synthetic chemistry as these compounds act as intermediates for Suzuki cross coupling reactions.³²

Experimental Procedures

1. Representative procedure for the preparation of hydroxyamide 4a: To a stirred solution of propion-aldehyde **1a** (0.14 mL, 2.0 mmol) in 2mL DMF was added benzyl isonitrile **2a** (0.24 mL, 2.0 mmol), and boric acid (0.12 g, 2.0 mmol) and stirred overnight at room temperature. Upon completion (TLC), the reaction mixture was worked up with water and diethyl ether (3 x 25 mL). The combined organic layers were dried (MgSO₄), concentrated *in vacuo*, and purified by column chromatography (silica gel, hexane:acetone, 4:1) to obtain 0.30 g (78%) of hydroxy amide **4a**. ¹H NMR (500 MHz, CDCl₃): 7.22-7.32 (m, 5H), 7.18 (bs, 1H), 4.47 (t, *J* = 4.5 Hz, 1H), 4.42 (dd, *J* = 4.0, 15.0 Hz, 1H), 4.36 (dd, *J* = 4.0, 15.0 Hz, 1H), 4.05-4.08 (m, 1H), 1.78-1.90 (m, 1H), 1.62-1.72 (m, 1H), 0.95 (t, *J* = 7.5 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃): 174.5, 138.2, 128.9, 127.9, 127.7, 73.2, 43.3, 28.1, 9.4; ESI-MS: 216 [(M+Na)⁺, 100%], 194 (M+H)⁺.

2. Representative procedure for the preparation of benzoxaborole 6c: To a stirred solution of boronoaldehyde **5** (0.3 g, 2.0 mmol) in 2mL DMF was added *tert*-butyl isonitrile (0.23 mL, 2.0 mmol), and stirred overnight at room temperature. Upon completion (TLC), the reaction mixture was worked up with water and ethyl acetate (3 x 25 mL). The combined organic layers were dried (MgSO₄), concentrated *in vacuo*, and purified by column chromatography (silica gel, hexane:ethyl acetate, 3:1) to obtain 0.30 g (65%) of benzoxaborole **6c**. (Found: C, 61.53; H, 8.10; N, 6.02 %; C₁₂H₁₆BNO₃ requires: C, 61.84; H, 6.92; N, 6.01%); ¹H NMR (500 MHz, DMSO-d₆): 9.33 (bs, 1H), 7.67 (d, *J* = 7.2 Hz, 1H),

7.62 (d, $J = 7.6$ Hz, 1H), 7.36 (t, $J = 7.4$ Hz, 1H), 7.26 (t, $J = 7.4$ Hz, 1H), 6.52 (s, 1H), 5.27 (s, 1H), 1.21 (s, 9H); ^{13}C NMR (125 MHz, CDCl_3): 169.4, 152.3, 131.3, 130.8, 128.1, 122.9, 80.2, 51.1, 28.9; ESI-MS: 232 $[(\text{M}-\text{H})^+]$, 100%].

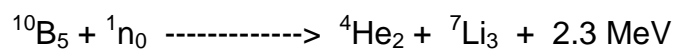
Conclusions

In conclusion, we have introduced a novel methodology for the synthesis of α -hydroxyamides using boric acid as catalyst upon the α -addition of isonitriles to aldehydes. We have also extended this protocol for the synthesis of functionalized benzoxaboroles via intramolecular cyclization utilized Passerini type multicomponent coupling reaction. These synthesized compounds act as excellent synthons and intermediates for the applications in organic and medicinal chemistry.

Overall conclusion

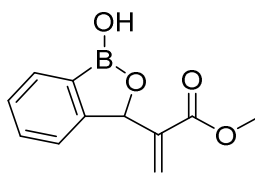
As these compounds did not exhibit any antimicrobial efficacy or cytotoxicity, they can be used for boron neutron capture therapy (BNCT) since these compounds mimic glycolytic metabolites such as fructose.

Boron neutron capture therapy is a binary therapy in which, the tumor cells are loaded with high concentration of ^{10}B and later activated by applying a radiation field of low energy neutrons. ^{10}B is non-radioactive and is present in 20% of natural boron. Neutron capture results in the formation of excited ^{11}B nuclei, which then undergo fission to produce high linear energy transfer (LET) particles such as α - ($^4\text{He}_2$), $^7\text{Li}_3$ and γ -particles. Cell death is triggered by the release of these heavily charged particles (He and Li) that create ionization tracks along their trajectories.



Binding studies and boron accumulation studies have to be carried out to determine the potency of these compound for their use in BNCT.

Spectral Characterization

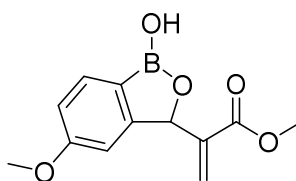


7a

Methyl 2-(1-hydroxy-1,3-dihydrobenzo[c][1,2]oxaborol-3-yl)acrylate 7a: yield = 58%

^1H NMR (500MHz, DMSO- d_6) δ 9.39 (s, 1H), 7.75 (d, $J = 7.5$ Hz, 1H), 7.46 (t, $J = 7.5$ Hz, 1H), 7.37 (t, $J = 7$ Hz, 1H), 7.30 (d, $J = 7.5$ Hz, 1H), 6.20 (s, 1H), 5.98 (s, 1H), 5.84 (s, 1H), 3.72 (s, 3H)

^{13}C NMR (125 MHz, DMSO- d_6) δ 166.2, 155.6, 140.8, 131.5, 131.3, 128.1, 126.6, 122.3, 79.3, 52.6

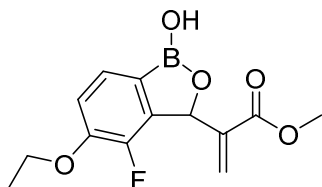


7b

Methyl 2-(1-hydroxy-5-methoxy-1,3-dihydrobenzo[c][1,2]oxaborol-3-yl)acrylate 7b: yield = 54%

^1H NMR (500MHz, DMSO- d_6) δ 9.19 (s, 1H), 7.63 (d, $J = 8$ Hz, 1H), 6.94 (d, $J = 8$ Hz, 1H), 6.80 (s, 1H), 6.19 (s, 1H), 5.89 (s, 1H), 5.81 (s, 1H), 3.76 (s, 3H), 3.72 (s, 3H)

^{13}C NMR (125 MHz, DMSO- d_6) δ 166.3, 162.5, 157.9, 140.8, 132.6, 126.6, 115.1, 107.2, 78.8, 55.8, 52.6

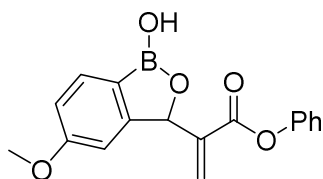


7c

Methyl 2-(5-ethoxy-4-fluoro-1-hydroxy-1,3-dihydrobenzo[c][1,2]oxaborol-3-yl) acrylate 7c: yield = 56%

^1H NMR (500MHz, DMSO- d_6) δ 9.37 (s, 1H), 7.52 (d, $J = 7$ Hz, 1H), 7.19 (t, $J = 5$ Hz, 1H), 6.22 (s, 1H), 6.04 (s, 1H), 5.89 (s, 1H), 4.17 (q, $J = 6$ Hz, 2H), 3.70 (s, 3H), 1.38 (t, $J = 6$ Hz, 3H)

^{13}C NMR (125 MHz, DMSO- d_6) δ 165.8, 144.3, 142.8, 140.6, 139.3, 128.6, 127.7, 115.9, 76.8, 65.3, 52.6, 15.3

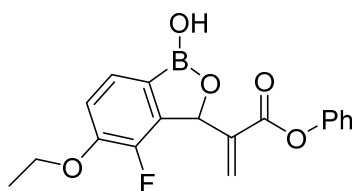


7e

Phenyl 2-(1-hydroxy-5-methoxy-1,3-dihydrobenzo[c][1,2]oxaborol-3-yl)acrylate 7e: yield = 48%

^1H NMR (500MHz, DMSO- d_6) δ 9.24 (s, 1H), 7.66 (d, J = 8 Hz, 1H), 7.44 (t, J = 8 Hz, 2H), 7.29 (t, J = 7 Hz, 1H), 7.12 (d, J = 9 Hz, 2H), 6.96 (d, J = 8.5 Hz, 1H), 6.88 (s, 1H), 6.49 (s, 1H), 6.08 (s, 1H), 6.00 (s, 1H), 3.78 (s, 3H)

^{13}C NMR (125 MHz, DMSO- d_6) δ 164.5, 162.6, 157.9, 150.9, 140.2, 132.6, 130.3, 129.2, 126.8, 122.3, 115.3, 107.1, 79.1, 55.9



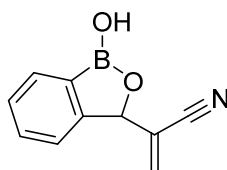
7f

Phenyl 2-(5-ethoxy-4-fluoro-1-hydroxy-1,3-dihydrobenzo[c][1,2]oxaborol-3-yl)acrylate 7f: yield = 41%

^1H NMR (500MHz, DMSO- d_6) δ 9.40 (s, 1H), 7.50 (d, J = 8 Hz, 3H), 7.44 (t, J = 8 Hz, 2H), 7.30 (t, J = 8 Hz, 1H), 7.25 (t, J = 7.5 Hz, 1H), 7.08 (d, J = 8.5 Hz, 2H),

6.55 (s, 1H), 6.18 (s, 1H), 6.08 (s, 1H), 4.187(q, $J = 7$ Hz, 2H), 1.37 (t, $J = 7$ Hz, 3H)

^{13}C NMR (125 MHz, DMSO- d_6) δ 164.1, 150.8, 138.9, 130.9, 130.3, 126.8, 122.3, 116.0, 77.0, 65.4, 15.3

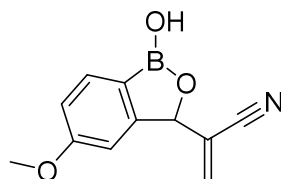


7g

2-(1-hydroxy-1,3-dihydrobenzo[c][1,2]oxaborol-3-yl)acrylonitrile 7g: yield = 69%

^1H NMR (500MHz, DMSO- d_6) δ 9.63 (s, 1H), 7.80 (d, $J = 7$ Hz, 1H), 7.56 (t, $J = 7$ Hz, 1H), 7.46 (t, $J = 7$ Hz, 1H), 7.35 (d, $J = 8$ Hz, 1H), 6.49 (s, 1H), 6.32 (s, 1H), 5.87 (s, 1H)

^{13}C NMR (125 MHz, DMSO- d_6) δ 153.0, 134.3, 131.8, 131.2, 128.8, 124.0, 122.4, 116.5, 80.2



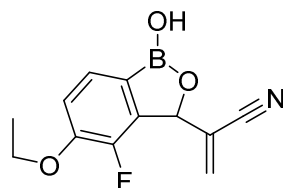
7h

2-(1-hydroxy-5-methoxy-1,3-dihydrobenzo[c][1,2]oxaborol-3-yl)acrylonitrile

7h: yield = 62%

^1H NMR (500MHz, DMSO- d_6) δ 9.42 (s, 1H), 7.67 (d, J = 8 Hz, 1H), 7.01 (d, J = 7.5 Hz, 1H), 6.85 (s, 1H), 6.47 (s, 1H), 6.32 (s, 1H), 5.78 (s, 1H), 3.80 (s, 3H)

^{13}C NMR (125 MHz, DMSO- d_6) δ 162.9, 155.8, 134.6, 132.8, 124.2, 116.8, 116.3, 107.1, 80.1, 56.0

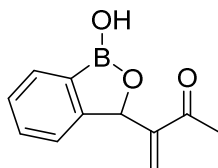


7i

2-(5-ethoxy-4-fluoro-1-hydroxy-1,3-dihydrobenzo[c][1,2]oxaborol-3-yl)acrylonitrile 7i: yield = 54%

^1H NMR (500MHz, DMSO- d_6) δ 9.65 (s, 1H), 7.54 (d, J = 7.5 Hz, 1H), 7.29 (t, J = 7.5 Hz, 1H), 6.50 (s, 1H), 6.39 (s, 1H), 6.01 (s, 1H), 4.18 (q, J = 7.5 Hz, 2H), 1.36 (t, J = 7 Hz, 1H)

^{13}C NMR (125 MHz, DMSO- d_6) δ 149.5, 149.2, 148.0, 145.0, 139.3, 139.2, 135.6, 128.2, 128.1, 122.9, 117.0, 116.5, 79.9, 77.6, 65.5, 15.2

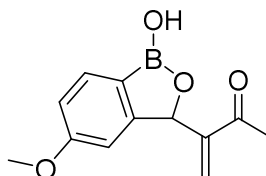


7j

3-(1-hydroxy-1,3-dihydrobenzo[c][1,2]oxaborol-3-yl)but-3-en-2-one 7j: yield = 49%

^1H NMR (500MHz, DMSO- d_6) δ 9.38 (s, 1H), 7.73 (d, J = 7.5 Hz, 1H), 7.43 (t, J = 7.5 Hz, 3H), 7.35 (t, J = 7 Hz, 1H), 7.20 (d, J = 7 Hz, 1H), 6.28 (s, 1H), 6.05 (s, 1H), 5.90 (s, 1H), 2.39 (s, 3H)

^{13}C NMR (125 MHz, DMSO- d_6) δ 199.9, 156.4, 148.4, 131.5, 131.3, 128.0, 126.8, 122.5, 78.1, 27.3

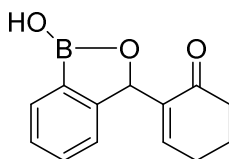


7k

3-(1-hydroxy-5-methoxy-1,3-dihydrobenzo[c][1,2]oxaborol-3-yl)but-3-en-2-one 7k: yield = 44%

^1H NMR (500MHz, DMSO- d_6) δ 9.19 (s, 1H), 7.63 (d, $J = 8$ Hz, 1H), 6.92 (d, $J = 8$ Hz, 1H), 6.72 (s, 1H), 6.28 (s, 1H), 6.98 (s, 1H), 5.89 (s, 1H), 3.74 (s, 1H), 2.40 (s, 3H)

^{13}C NMR (125 MHz, DMSO- d_6) δ 200.0, 162.5, 158.8, 148.5, 132.6, 126.8, 115.0, 107.4, 77.7, 55.8, 27.3



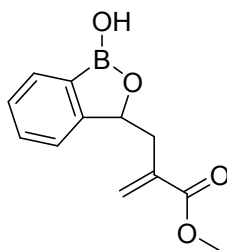
7m

2-(1-hydroxy-1,3-dihydrobenzo[c][1,2]oxaborol-3-yl)cyclohex-2-en-1-one

7m: yield = 56%

^1H NMR (500MHz, DMSO- d_6) δ 9.30 (s, 1H), 7.71 (d, $J = 7$ Hz, 1H), 7.42 (t, $J = 7$ Hz, 1H), 7.33 (t, $J = 7$ Hz, 1H), 7.22 (d, $J = 7$ Hz, 1H), 6.82 (t, $J = 4$ Hz, 1H), 5.98 (s, 1H), 2.42 (t, $J = 7$ Hz, 2H), 2.34 (m, 2H), 1.89 (m, 2H)

^{13}C NMR (125 MHz, DMSO- d_6) δ 198.7, 157.0, 147.1, 139.0, 131.5, 131.1, 127.8, 122.5, 77.4, 25.8, 22.9



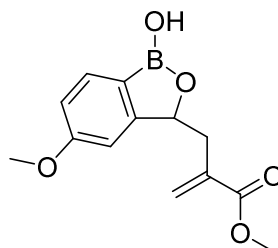
13a

Methyl 2-((1-hydroxy-1,3-dihydrobenzo[c][1,2]oxaborol-3-yl)methyl)acrylate

13a: yield = 75%

^1H NMR (500MHz, DMSO- d_6) δ 9.17 (s, 1H), 7.71 (d, J = 7 Hz, 1H), 7.48 (t, J = 7.5 Hz, 1H), 7.40 (d, J = 7.5 Hz, 1H), 7.35 (t, J = 7.5 Hz, 1H), 6.17 (s, 1H), 5.76 (s, 1H), 5.31 (dd, J = 7 Hz, 1H), 3.98 (s, 3H), 2.94 (d, J = 14.5, 1H), 2.43 (dd, J = 8.5 Hz, 1H)

^{13}C NMR (125 MHz, DMSO- d_6) δ 167.7, 157.4, 156.8, 137.1, 131.2, 128.2, 127.9, 122.2, 79.2, 52.5,



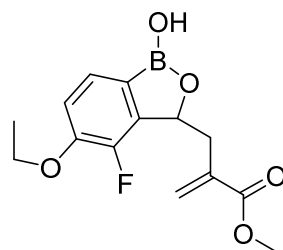
13b

methyl 2-((1-hydroxy-5-methoxy-1,3-dihydrobenzo[c][1,2]oxaborol-3-

yl)methyl) acrylate 13b: yield = 72%

^1H NMR (500MHz, DMSO- d_6) δ 8.96 (s, 1H), 7.58 (d, $J = 8$ Hz, 1H), 6.94 (s, 1H), 6.91 (d, $J = 8$ Hz, 1H), 6.16 (s, 1H), 5.76 (s, 1H), 5.22 (dd, $J = 4$ Hz, 1H), 3.80 (s, 3H), 3.70 (s, 3H), 2.94 (dd, $J = 6$ Hz, 1H), 2.40 (dd, $J =$, 1H)

^{13}C NMR (125 MHz, DMSO- d_6) δ 167.6, 162.4, 159.4, 137.2, 132.4, 128.1, 115.3, 106.8, 78.8, 55.9, 52.5

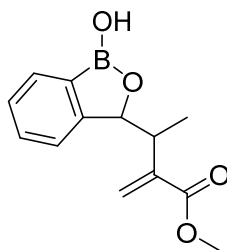


13c

Methyl 2-((5-ethoxy-4-fluoro-1-hydroxy-1,3-dihydrobenzo[*c*][1,2]oxaborol-3-yl) methyl)acrylate 13c: yield = 69%

^1H NMR (500MHz, DMSO- d_6) δ 9.19 (s, 1H), 7.43 (d, $J = 8$ Hz, 1H), 7.17 (t, $J = 7$ Hz, 1H), 6.14 (s, 1H), 5.71 (s, 1H), 5.44 (dd, $J = 2.5, 8$, 1H), 4.15 (q, $J = 6.5$ Hz, 2H), 3.66 (s, 3H), 3.03 (d, $J = 12$ Hz, 1H), 2.47 (dd, $J = 8.5$ Hz, 14.5, 1H), 1.36 (t, $J = 6$ Hz, 3H)

^{13}C NMR (125 MHz, DMSO- d_6) δ 167.4, 149.1, 147.7, 145.8, 142.6, 142.5, 136.6, 128.6, 127.7, 115.9, 76.5, 65.3, 52.5, 15.3

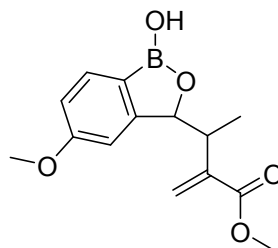


13d

Methyl 3-(1-hydroxy-1,3-dihydrobenzo[c][1,2]oxaborol-3-yl)-2-methylenebutanoate 13d: yield = 76%

^1H NMR (500MHz, DMSO- d_6) δ 9.24(s, 1H), 7.73 (d, J = 8 Hz, 1H), 7.48 (t, J = 6 Hz, 1H), 7.36 (t, J = 7 Hz, 2H), 6.30 (s, 1H), 5.84 (s, 1H), 5.28 (s, 1H), 3.75 (s, 3H), 3.16(q, J = 3.5, 1H), 0.67 (d, J = 7 Hz, 3H)

^{13}C NMR (125 MHz, DMSO- d_6) δ 167.7, 156.0, 143.0, 131.5, 131.1, 127.4, 126.4, 121.9, 81.9, 52.7, 39.7, 39.3, 12.4

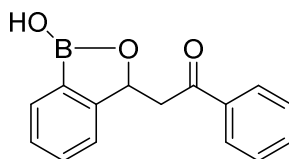


13e

Methyl 3-(1-hydroxy-5-methoxy-1,3-dihydrobenzo[c][1,2]oxaborol-3-yl)-2-methylenebutanoate 13e: yield = 74%

^1H NMR (500MHz, DMSO- d_6) δ 9.02 (s, 1H), 7.61 (d, $J = 8$ Hz, 1H), 6.93 (dd, $J = 2, 8$ Hz, 1H), 6.89 (s, 1H), 6.29 (s, 1H), 5.85 (s, 1H), 5.21 (d, $J = 3$ Hz, 1H), 3.81 (s, 3H), 3.76 (s, 3H), 3.17-3.3.13 (m, 1H), 0.68 (d, $J = 7$ Hz, 3H)

^{13}C NMR (125 MHz, DMSO- d_6) δ 167.7, 162.6, 158.6, 143.1, 132.4, 126.3, 115.2, 106.5, 81.6, 55.9, 52.7, 39.2, 31.4, 12.3



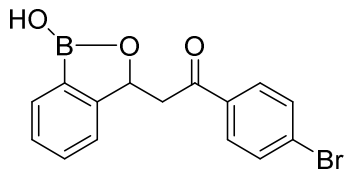
15a

2-(1-hydroxy-1,3-dihydrobenzo[c][1,2]oxaborol-3-yl)-1-phenylethan-1-one

15a: yield = 60%

^1H NMR (500MHz, DMSO- d_6) δ 9.20 (s, 1H), 8.03 (dd, $J = 1, 7.5$, 2H), 7.74 (d, $J = 7.5$, 1H), 7.68-7.65 (m, 1H), 7.56-7.48 (m, 4H), 7.40-7.37 (m, 1H), 5.71 (dd, $J = 3.75, 8.75$, 1H), 3.63-3.55 (m, 1H), 3.38-3.33 (m 1H)

^{13}C NMR (125 MHz, DMSO- d_6) δ 198.1, 157.0, 137.2, 133.8, 131.2, 130.9, 130.8, 129.2, 128.7, 127.7, 122.0, 77.3, 70.6, 45.8



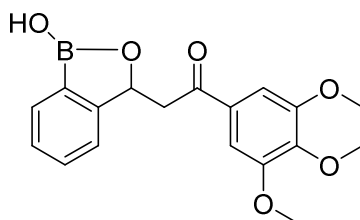
15c

1-(4-bromophenyl)-2-(1-hydroxy-1,3-dihydrobenzo[c][1,2]oxaborol-3-

yl)ethan-1-one 15c: yield = 45%

^1H NMR (500MHz, DMSO- d_6) δ 9.2 (s, 1H), 7.95 (d, J = 8, 2H), 7.76-7.72 (m, 3H), 7.51-7.38 (m, 3H), 5.68 (dd, J = 3.5, 5, 1H), 3.58 (dd, J = 3.5, 17, 1H), 3.33 (dd, J = 9, 17, 1H)

^{13}C NMR (125 MHz, DMSO- d_6) δ 197.0, 157.1, 153.6, 142.4, 132.6, 131.1, 130.9, 127.7, 122.1, 106.3, 77.3, 60.6, 56.5, 45.6

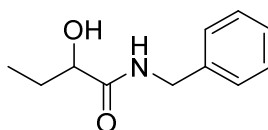


15d

2-(1-hydroxy-1,3-dihydrobenzo[*c*][1,2]oxaborol-3-yl)-1-(3,4,5-trimethoxyphenyl)ethan-1-one 15d: yield = 47%

^1H NMR (500MHz, DMSO- d_6) δ 9.2 (s, 1H), 7.74 (d, J = 7, 1H), 7.52 (t, J = 7.75, 2H), 7.38 (t, J = 7, 1H), 7.31 (s, 2H), 5.71 (dd, J = 4, 8.5, 1H), 3.86 (s, 3H), 3.76 (s, 6H), 3.47 (m, 2H)

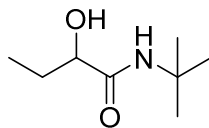
^{13}C NMR (125 MHz, DMSO- d_6) δ 197.0, 157.1, 153.3, 142.4, 132.6, 131.1, 130.9, 127.7, 122.1, 106.3, 77.3, 60.6, 56.5, 45.6



1

***N*-benzyl-2-hydroxybutanamide 1:** yield = 78%

^{13}C NMR (125 MHz, CDCl_3) δ 174.5, 138.2, 129.0, 127.9, 127.8, 73.2, 43.3, 28.0, 9.4

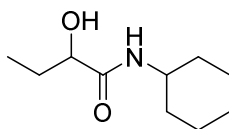


2

***N*-(tert-butyl)-2-hydroxybutanamide 2:** yield = 74%

^1H NMR (500MHz, CDCl_3) δ 6.67 (br, s, 1H), 4.53 (d, $J = 5$ Hz, 1H), 3.90 (m, 1H), 1.78 (m, 1H), 1.64 (m, 1H), 1.32 (s, 9H), 0.91 (t, $J = 8$ Hz)

^{13}C NMR (125 MHz, CDCl_3) δ 174.0, 73.0, 51.0, 28.8, 27.9, 9.1

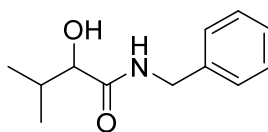


3

***N*-cyclohexyl-2-hydroxybutanamide 3:** yield = 75%

^1H NMR (500MHz, CDCl_3) δ 6.59 (br, d, 1H), 4.03 (t, $J = 3.5$, 1H), 3.72 (m, 2H), 1.86 (m, 4H), 1.65 (m, 4H), 1.18 (m, 3H), 0.96 (t, $J = 7$ Hz, 3H)

^{13}C NMR (125 MHz, CDCl_3) δ 173.3, 73.0, 48.1, 33.3, 28.1, 25.7, 25.0, 9.2

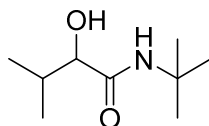


4

N-benzyl-2-hydroxy-3-methylbutanamide 4: yield = 70%

^1H NMR (500MHz, CDCl_3) δ 7.35-7.28 (m, 5H), 7.00 (br, s 1H), 4.46 (dd, $J = 6$ Hz, 2H), 4.02 (s, 1H), 2.20 (m, 1H), 1.03 (d, $J = 7$ Hz, 3H), 0.87 (d, $J = 7$ Hz, 3H)

^{13}C NMR (125 MHz, CDCl_3) δ 173.7, 138.3, 129.0, 128.0, 127.8, 76.6, 43.4, 32.1, 19.4, 15.8

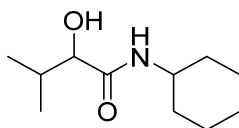


5

N-(tert-butyl)-2-hydroxy-3-methylbutanamide 5: yield = 71%

^1H NMR (500MHz, CDCl_3) δ 6.53(br, s, 1H), 3.92(d, $J = 5.5$ Hz, 1H), 3.80(m, 1H), 2.10(m, 1H), 1.35(s, 9H), 0.99(d, $J = 6.5$ Hz, 3H), 0.84(d, $J = 7$ Hz, 3H)

^{13}C NMR (125 MHz, CDCl_3) δ 173.3, 76.3, 51.1, 32.0, 29.0, 19.5, 15.6



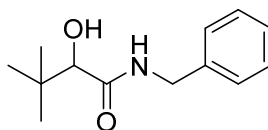
56

6

***N*-cyclohexyl-2-hydroxy-3-methylbutanamide 6**: yield = 74%

^1H NMR (500MHz, CDCl_3) δ 6.57 (br, d, $J = 7.5$ Hz, 1H), 3.90 (d, $J = 8$ Hz, 1H), 3.76 (m, 1H), 3.59 (d, $J = 5$ Hz, 1H), 2.14 (m, 1H), 1.89 (m, 2H), 1.71 (m, 2H), 1.61 (m, 1H), 1.36 (m, 2H), 1.16 (m, 3H), 1.01 (d, $J = 7$ Hz, 3H), 0.84 (d, $J = 6.5$, 3H)

^{13}C NMR (125 MHz, CDCl_3) δ 172.9, 76.2, 48.0, 33.4, 33.2, 32.1, 25.7, 25.0, 19.4, 15.6

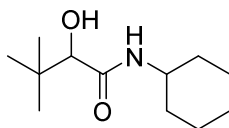


7

***N*-benzyl-2-hydroxy-3,3-dimethylbutanamide 7**: yield = 61%

^1H NMR (500MHz, CDCl_3) δ 7.33-7.28 (m, 5H), 6.81 (br, s, 1H), 4.44 (dd, $J = 5.5$ Hz, 2H), 3.74 (d, $J = 5$ Hz, 1H), 3.51 (d, $J = 5.5$ Hz, 1H), 0.90 (s, 9H)

^{13}C NMR (125 MHz, CDCl_3) δ 173.0, 138.3, 128.9, 128.0, 127.7, 79.8, 43.4, 35.4, 26.3

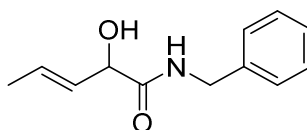


57

9

***N*-cyclohexyl-2-hydroxy-3,3-dimethylbutanamide 9:** yield = 72%

¹H NMR (500MHz, CDCl₃) δ 6.08 (br, s, 1H), 3.80 (br, 1H), 3.65 (d, *J* = 6 Hz, 1H), 3.18 (d, *J* = 5.5 Hz, 1H), 1.90 (m, 2H), 1.73-1.61 (m, 2H), 1.42-1.16 (m, 6H), 0.99 (s, 9H)

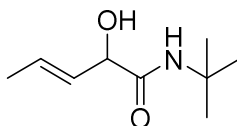


10

***(E)*-*N*-benzyl-2-hydroxypent-3-enamide 10:** yield = 78%

¹H NMR (500MHz, CDCl₃) δ 7.36-7.26 (m, 5H), 6.82 (br, s, 1H), 5.88 (dd, *J* = 6.5 Hz, 1H), 5.60 (dd, *J* = 7 Hz, 1H), 4.53 (d, *J* = 7 Hz, 1H), 4.45 (d, *J* = 6 Hz, 2H), 3.71 (br, s, 1H), 1.74 (d, *J* = 6.5 Hz, 3H)

¹³C NMR (125 MHz, CDCl₃) δ 172.8, 138.0, 131.0, 129.1, 129.0, 127.9, 127.8, 73.18, 43.6, 18.0

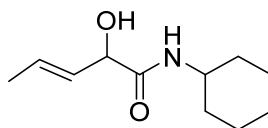


11

***(E)*-*N*-(tert-butyl)-2-hydroxypent-3-enamide 11:** yield = 74%

^1H NMR (500MHz, CDCl_3) δ 5.99(br, s, 1H), 5.88(m, 1H), 5.54(dd, $J = 7.5$ Hz, 1H), 4.35(d, $J = 11$ Hz, 3.39(d, $J = 3.5$ Hz, 1H), 1.77(d, 7.5 Hz, 3H), 1.37(s, 9H)

^{13}C NMR (125 MHz, CDCl_3) δ 171.7, 131.3, 129.6, 73.3, 51.6, 29.0, 18.0

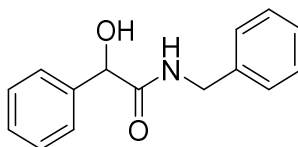


12

(E)-N-cyclohexyl-2-hydroxy-3-enamide 12: yield = 72%

^1H NMR (500MHz, CDCl_3) δ 5.88 (br, s, 1H) 5.81 (dd, $J = 6.5$ Hz, 1H), 5.47 (dd, $J = 7.5$ Hz, 1H), 4.36 (d, $J = 7.5$ Hz, 1H), 3.70(m, 1H), 1.84 (d, $J = 12.5$, 3H), 1.69 (d, $J = 6.5$, 3H), 1.67-0.76 (m, 8H)

^{13}C NMR (125 MHz, CDCl_3) δ 171.5, 131.3, 129.4, 73.1, 48.6, 33.3, 25.7, 25.0, 18.0

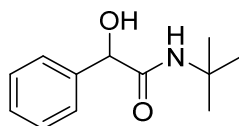


13

N-benzyl-2-hydroxy-2-phenylacetamide 13: yield = 80%

^1H NMR (500MHz, CDCl_3) δ 7.34-7.12(m, 10H), 4.95(s, 1H), 4.65(br, s, 1H), 4.31(s, 2H)

^{13}C NMR (125 MHz, CDCl_3) δ 172.0, 139.9, 138.0, 128.9, 128.6, 128.0, 127.8, 127.7, 127.0, 74.4, 43.5

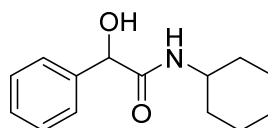


14

***N*-(tert-butyl)-2-hydroxy-2-phenylacetamide 14:** yield = 76%

^1H NMR (500MHz, CDCl_3) δ 7.36-7.29 (m, 5H), 6.21 (br, s, 1H), 4.84 (s, 1H), 4.22 (br, s, 1H), 1.31 (s, 9H)

^{13}C NMR (125 MHz, CDCl_3) δ 171.5, 140.1, 129.1, 128.8, 127.1, 74.5, 51.7, 28.8

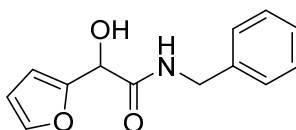


15

***N*-cyclohexyl-2-hydroxy-2-phenylacetamide 15:** yield = 80%

^1H NMR (500MHz, CDCl_3) δ 7.25 (m, 5H), 6.31 (s, 1H), 4.81 (s, 1H), 4.22 (s, 1H), 3.61 (m, 1H), 1.73-0.71 (m, 10H)

^{13}C NMR (125 MHz, CDCl_3) δ 140.2, 128.8, 128.4, 127.0, 74.2, 48.4, 33.0, 32.9, 25.7, 25.0

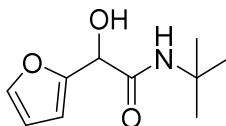


16

***N*-benzyl-2-(furan-2-yl)-2-hydroxyacetamide 16:** yield = 65%

^1H NMR (500MHz, CDCl_3) δ 7.32 (d, $J = 2$ Hz 1H), 7.27-7.12 (m, 5H), 6.54 (br s, 1H), 6.30 (d, $J = 20$, 2H), 5.08(s, 1H), 4.43 (m, 2H), 3.63(br, 1H),

^{13}C NMR (125 MHz, CDCl_3) δ 170.4, 152.0, 143.2, 137.8, 129.0, 127.9, 127.8, 110.8, 108.9, 68.1, 43.7



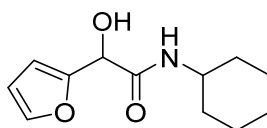
17

***N*-(tert-butyl)-2-(furan-2-yl)-2-hydroxyacetamide 17:** yield = 68%

^1H NMR (500MHz, CDCl_3) δ 7.412(t, $J = 1$ Hz, 1H), 6.37(d, $J = 3$ Hz, 2H), 6.10(br, s, 1H), 5.00(s, 1H), 3.96(br, s, 1H), 1.37(s, 9H)

61

^{13}C NMR (125 MHz, CDCl_3) δ 169.0, 152.5, 143.1, 110.8, 108.5, 68.0, 51.9, 28.9

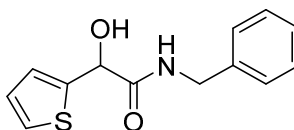


18

N-cyclohexyl-2-(furan-2-yl)-2-hydroxyacetamide 18: yield = 67%

^1H NMR (500MHz, CDCl_3) δ 7.41 (d, $J = 1.5$ Hz, 1H), 6.40-6.37 (m, 2H), 6.2 (d, $J = 3.5$ Hz, 1H), 3.92(br, s, 1H), 3.84-3.78 (m, 1H), 1.95-1.87 (m, 2H), 1.77-1.60 (m, 4H), 1.42-1.33 (m, 2H), 1.27-1.14 (m, 3H)

^{13}C NMR (125 MHz, CDCl_3) δ 169.0, 152.2, 143.1, 110.9, 108.6, 67.9, 48.9, 33.1, 25.7, 24.9

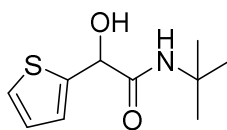


19

N-benzyl-2-hydroxy-2-(thiophen-2-yl)acetamide 19: yield = 75%

^1H NMR (500MHz, CDCl_3) δ 7.28 (m, 6H), 7.07 (d, $J = 3.5$, 1H), 7.04 (br, s, 1H), 6.95 (t, $J = 5$ Hz, 1H), 5.30 (s, 1H), 4.65 (br, s, 1H), 4.41 (t, $J = 5.5$ Hz, 2H)

^{13}C NMR (125 MHz, CDCl_3) δ 171.8, 161.7, 142.9, 137.8, 129.0, 128.0, 127.9, 127.1, 126.1, 70.4, 43.7

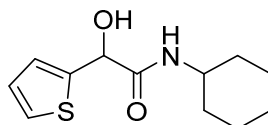


20

***N*-(tert-butyl)-2-hydroxy-2-(thiophen-2-yl)acetamide 20:** yield = 72%

^1H NMR (500MHz, CDCl_3) δ 7.28(d, $J = 5.5$ Hz, 1H), 7.06(d, $J = 4.5$ Hz, 1H), 6.98(t, $J = 4.5$ Hz, 1H), 6.34(br,s, 1H), 5.13(s,1H), 4.48(br, s, 1H), 1.35(s, 9H)

^{13}C NMR (125 MHz, CDCl_3) δ 170.7, 143.3, 127.0, 126.1, 126.0, 70.4, 51.7, 28.8

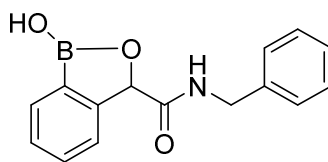


21

***N*-cyclohexyl-2-hydroxy-2-(thiophen-2-yl)acetamide 21:** yield = 78%

^1H NMR (500MHz, CDCl_3) δ 7.31 (d, $J = 4$ Hz, 1H), 7.11 (d, $J = 3.5$, 1H), 7.00(t, $J = 3$ Hz, 1H), 6.25 (br, d, $J = 6.5$, 1H), 4.09 (d, $J = 4$ Hz, 1H), 3.80 (m, 1H), 1.91-1.90 (m, 2H), 1.73-1.60 (m, 3H), 1.41-1.32(m, 2H), 1.21-1.16(m, 2H)

^{13}C NMR (125 MHz, CDCl_3) δ 171.2, 143.0, 127.1, 126.3, 126.2, 70.2, 48.8, 33.0, 25.7, 24.9



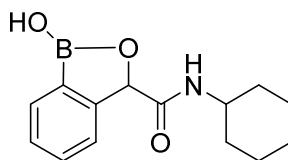
6a

***N*-benzyl-1-hydroxy-1,3-dihydrobenzo[*c*][1,2]oxaborole-3-carboxamide 6a:**

yield = 81%

^1H NMR (500MHz, DMSO- d_6) δ 9.55 (s, 1H), 8.57 (t, J = 8 Hz, 1H), 7.76 (d, J = 7 Hz, 1H), 7.56 (d, J = 7.5 Hz, 2H), 7.49 (t, J = 7 Hz, 1H), 7.41 (t, J = 7.5 Hz, 2H), 7.30-7.19 (m, 3H), 5.56 (s, 1H), 4.31 (dd, J = 6.5 Hz, 1H), 4.22 (dd, J = 6 Hz, 1H)

^{13}C NMR (125 MHz, DMSO- d_6) δ 170.2, 153.1, 140.0, 131.6, 131.2, 128.9, 128.5, 127.9, 127.5, 122.6, 80.4, 42.6



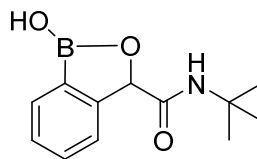
6b

***N*-cyclohexyl-1-hydroxy-1,3-dihydrobenzo[*c*][1,2]oxaborole-3-carboxamide**

6b: yield = 76%

^1H NMR (500MHz, DMSO- d_6) δ 9.42 (br, s, 1H), 7.82 (d, J = 8 Hz, 1H), 7.76 (d, J = 7.5 Hz, 1H), 7.51-7.38 (m, 3H), 5.42 (s, 1H), 3.56 (m, 1H), 1.79-0.85 (m, 10H)

^{13}C NMR (125 MHz, DMSO- d_6) δ 168.9, 153.3, 131.5, 131.1, 128.3, 122.3, 80.2, 79.9, 48.3, 32.9, 32.8, 25.8, 25.4, 25.3



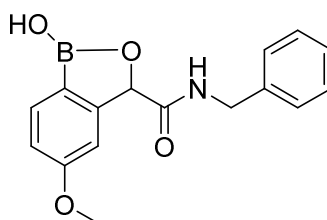
6c

***N*-(tert-butyl)-1-hydroxy-1,3-dihydrobenzo[*c*][1,2]oxaborole-3-carboxamide**

6c: yield = 65%

^1H NMR (500MHz, DMSO- d_6) δ 9.42 (br, s, 1H), 7.71 (d, J = 7 Hz, 1H), 7.56-7.45 (m, 3H), 7.40-7.38 (m, 1H), 5.42 (s, 1H), 1.24 (s, 9H)

^{13}C NMR (125 MHz, DMSO- d_6) δ 169.1, 153.4, 131.4, 131.0, 128.3, 122.3, 80.3, 79.9, 51.0, 29.1

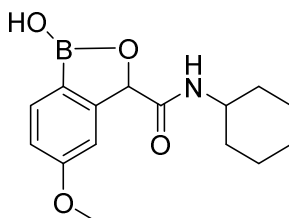


6d

***N*-benzyl-1-hydroxy-5-methoxy-1,3-dihydrobenzo[*c*][1,2]oxaborole-3-carboxamide 6d:** yield = 61%

^1H NMR (500MHz, DMSO- d_6) δ 9.36(s, 1H), 8.56 (t, $J = 6$ Hz, 1H), 7.65 (d, $J = 8$ Hz, 1H), 7.25 (m, 5H), 7.06 (d, $J = 2.5$, 1H), 6.97 (dd, $J = 2.5, 8.5$, 1H), 5.49 (s, 1H), 4.36 (dd, $J = 6.5$ Hz, 15, 1H), 4.19 (dd, $J = 6$ Hz, 15, 1H), 3.78 (s, 3H)

^{13}C NMR (125 MHz, DMSO- d_6) δ 170.1, 162.5, 155.5, 140.1, 132.5, 128.9, 128.0, 127.5, 115.8, 107.0, 80.1, 55.9, 42.6

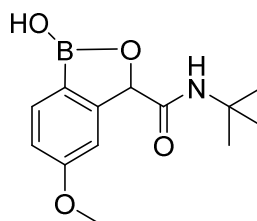


6e

N-cyclohexyl-1-hydroxy-5-methoxy-1,3-dihydrobenzo[c][1,2]oxaborole-3-carboxamide 6e: yield = 55%

^1H NMR (500MHz, DMSO- d_6) δ 9.30 (s, 1H), 7.84 (d, $J = 8$ Hz, 1H), 7.63 (d, $J = 8$ Hz, 1H), 7.03 (s, 1H), 6.95 (d, $J = 8$ Hz, 1H), 6.40 (s, 1H), 3.78 (s, 3H), 3.55 (br, s, 1H), 1.71-1.53 (m, 5H), 1.27-1.09 (m, 6H)

^{13}C NMR (125 MHz, DMSO- d_6) δ 168.9, 162.4, 155.7, 132.4, 115.5, 107.0, 79.9, 55.9, 48.2, 32.9, 32.8, 25.8, 25.4

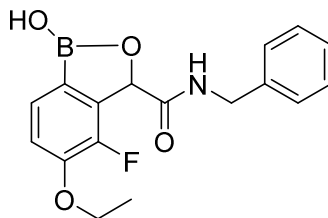


6f

***N*-(tert-butyl)-1-hydroxy-5-methoxy-1,3-dihydrobenzo[*c*][1,2]oxaborole-3-carboxamide 6f:** yield = 54%

^1H NMR (500MHz, DMSO- d_6) δ 9.28 (s, 1H), 7.63 (d, J = 8 Hz, 1H), 7.4 (s, 1H), 7.03 (s, 1H), 6.96 (dd, J = 2 Hz, 8, 1H), 5.34 (s, 1H), 3.79 (s, 3H), 1.28 (s, 9H)

^{13}C NMR (125 MHz, DMSO- d_6) δ 169.1, 162.4, 155.7, 132.4, 115.3, 107.1, 79.9, 55.9, 51.0, 29.1

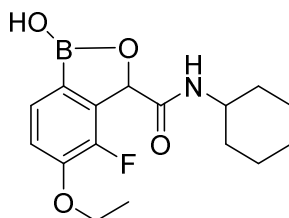


6g

***N*-benzyl-5-ethoxy-4-fluoro-1-hydroxy-1,3-dihydrobenzo[*c*][1,2]oxaborole-3-carboxamide 6g:** yield = 54%

^1H NMR (500MHz, DMSO- d_6) δ 9.45 (s, 1H), 8.86 (t, $J = 6$ Hz, 1H), 7.47 (d, $J = 8$ Hz, 1H), 7.33-7.25 (m, 6H), 5.68 (s, 1H), 4.39-4.07 (m, 4H), 1.36 4.19 (t, $J = 6.7$, 3H)

^{13}C NMR (125 MHz, DMSO- d_6) δ 168.7, 139.7, 129.0, 127.9, 127.6, 127.6, 116.2, 78.1, 65.4, 42.9, 15.3



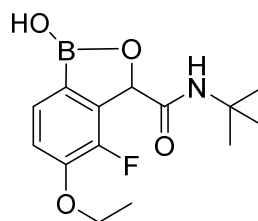
6h

***N*-cyclohexyl-5-ethoxy-4-fluoro-1-hydroxy-1,3-dihydrobenzo[*c*][1,2]**

oxaborole-3-carboxamide 6h: yield = 58%

^1H NMR (500MHz, DMSO- d_6) δ 9.38 (s, 1H), 8.31 (d, $J = 7.5$ Hz, 1H), 7.45 (d, $J = 7.5$ Hz, 1H), 7.18 (t, $J = 7.5$ Hz, 1H), 5.60 (s, 1H), 4.13 (q, $J = 7$, 2H), 3.53 (br, d, $J = 7.5$ Hz, 1H), 1.72-1.13 (m, 13H)

^{13}C NMR (125 MHz, DMSO- d_6) δ 167.4, 149.2, 149.1, 147.7, 145.7, 139.7, 127.5, 115.9, 77.8, 65.3, 48.3, 32.9, 31.4, 25.9, 25.2, 15.3



6i

***N*-(tert-butyl)-5-ethoxy-4-fluoro-1-hydroxy-1,3-dihydrobenzo[c][1,2]**

oxaborole-3-carboxamide 6i: yield = 49%

^1H NMR (500MHz, DMSO- d_6) δ 9.35 (s, 1H), 8.09 (s, 1H), 7.45 (d, J = 8 Hz, 1H), 7.2 (t, J = 8, 1H), 5.62 (s, 1H), 4.13 (q, J = 7 Hz, 1H), 1.36 (t, J = 7 Hz, 3H), 1.28 (s, 9H)

^{13}C NMR (125 MHz, DMSO- d_6) δ 167.7, 149.2, 149.0, 139.8, 127.5, 115.9, 78.0, 65.3, 51.1, 29.0, 15.3

References

1. (a) Miyaura, N.; Suzuki, A. *Chem. Rev.* **1995**, *95*, 2457; (b) Suzuki, A. *J. Organomet. Chem.* **1999**, *576*, 147.
2. Medicinal uses of Boronic acids: (a) Yang, W.; Gao, X.; Wang, B. *Med. Res. Rev.* **2003**, *23*, 346-368; (b) Dembitsky, V. M.; Srebnik, M. *Tetrahedron* **2003**, *59*, 579; (c) Coutts, S. J.; Kelly, T. A.; Snow, R. J.; Kennedy, C. A.; Barton, R. W.; Adams, J.; Krolikowski, D. A.; Freeman, D. M.; Campbell, S. J.; Ksiazek, J. F.; Bachovchin, W. W., *J. Med. Chem.* **1996**, *39*, 2087; (d) Kinder, D. H.; Frank, S. K.; Ames, M. M. *J. Med. Chem.* **1990**, *33*, 819; (e) Kettner, C. A.; Shenvi, A. B. *J. Biol. Chem.* **1984**, *259*, 15106.
3. Chauhan, D.; Catley, L.; Li, G.; Podar, K.; Hideshima, T.; Velanmkar, M.; Mitisades, C.; Mitsiades, N.; I Yasui, H.; Letai, A.; Ovaa, H.; Berkers, C.; Nicholson, B.; Chao, T.H.; Neuteboom, S. T. C.; Richardson, P.; Palladino, M. A.; Anderson, K. C. *Cancer Cell* **2005**, *8*, 407; (b) Chauhan, D.; Hideshima, T.; Anderson, K. C. *Brit. J. Cancer* **2006**, *95*, 961.
4. Barth, R. F.; Coderre, J. A.; Vicente, M. G.; Blue, T. E. *Clin Cancer Res.* **2005**, *11*, 3987-4002.
5. (a) Wozniack, A. A.; Cyranski, M. K.; Zubrowska, A.; Sporzynski, A. *J. Organomet. Chem.* **2009**, *694*, 3533 and references therein. (b) Ye, L.; Ding, D.; Feng, Y.; Xie, D.; Wu, P.; Guo, H.; Meng, Q.; Zhou, H. *Tetrahedron* **2009**, *65*, 8738-8744. (c) Tan, Y. L.; White, A. J. P.; Widdowson, D. A.; Wilhelm, R.; Williams, D. J. *J. Chem. Soc. Perkin Trans 1*, **2001**, 3269. (d) Nicolaou, K. C.; Li, H.; Boody, C. N. C.; Ramanjulu, J. M.; Yue, T. Y.; Natarajan, S.; Chu, X. J.; Brase, S.; Rubsam, F. *Chem. Eur. J.*, **1999**, *5*, 2584; (e) Nicolaou, K. C.; Natarajan, S.; Li, H.; Jain, N. F.; Hughes, R.; Solomon, M. E.; Ramanjulu, J. M.; Boody, C. N. C.; Takayanagi, M. *Angew. Chem. Int. Ed.*, **1998**, *37*, 2708.
6. (a) AN2690, AN2718, & AN2728 are being currently developed as therapeutic agents by Anacor Pharmaceuticals Inc., Palo Alto, CA, USA. (b) Akama, T.; Baker, S. J.; Zhang, Y. K.; Hernandez, V.; Zhou, H.; Sanders, V.; Freund, Y.; Kimura, R.; Maples, K. R.; Plattner, J. J. *Bioorg. Med. Chem. Lett.* **2009**, *19*, 2129. (c) Baker, S. J.; Zhang, Y. K.; Akama, T.; Lau, A.; Zhou, H.; Hernandez, V.; Mao W.; Alley, M. R. K.; Sanders, V.; Plattner, J. J. *J. Med. Chem.* **2006**, *49*, 4447.
7. (a) Alexander, C.; Smith, C. R.; Whitcombe, M. J.; Vulfson, E. N. *J. Am. Chem. Soc.* **1999**, *121*, 6640. (b) Russell, A. P.; Zepp, C. W. U.S. Patent, US5512246, **1996**. (c) Lowe, C. R.; Sartain, F. K.; Yang, X. P.C.T. Int. Appl.,

- WO2006079843, **2006**. (d) Austin, P.W.; Kneale, C. J.; Crowley, P. J.; Clough, J. M. P.C.T. Int. Appl., WO9533754, **1995**.
8. (a) Oshima, K.; Aoyama, Y. *J. Am. Chem. Soc.* **1999**, *121*, 2315. (b) Dowlut, M.; Hall, D. G. *J. Am. Chem. Soc.* **2006**, *128*, 4226. (c) Pal, A.; Berube, M.; Hall, D. G. *Angew. Chem. Int. Ed. Engl.* **2010**, *49*, 1492-1495.
9. (a) Torssell, *Ark. Kemi.*, 1957, *10*, 507 (b) Hawkins, R. T.; Lennarz, W.J.; Snyder, H.R.; *J. Am. Chem. Soc.*, **1960**, *82*, 3053 (c) Lennarz, J.; Snyder, H. R.; *J. Am. Chem. Soc.*, **1960**, *82*, 2172.
10. M.A. Grassberger, *Liebigs Ann. Chem.*, **1985**, 683
11. Tschampel, P.; Snyder, H.R.; *J. Org. Chem.*, **1964**, *29*, 2168.
12. Yamamoto, Y.; Ishii, J.; Nishiyama, H.; Itoh, K.; *J. Am. Chem. Soc.*, **2005**, *127*, 9625.
13. Ding, D.; Meng, Q.; Gao, G.; Zhao, Y.; Wang, Q.; Nare, B.; Jacobs, R.; Rock, F.; Alley, M. R. K.; Plattner, J. J.; Chen, G.; Li, D.; Zhou, H., *J. Med. Chem.* **2011**, *54*, 1276–1287.
14. Kim, H.; Kang, Y. J.; Kang, S.; Kim, K. T., *J. Am. Chem. Soc.*, **2012**, *134*, 4030–4033
15. Adamczyk-Wozniak, A.; Madura, I.; Velders, A. H.; Sporzynski, A., *Tetrahedron Letters*, **2010**, *51*, 6181–6185
16. Gunasekera, D. S.; Gerold, D. J.; Alderks, N. S.; Chandra, J. S.; Maanu, C. A.; Kiprof, P.; Zhdankin, V. V.; Reddy, M. V. R., *Tetrahedron*, **2007**, *63*, 9401-9405
17. Qiao, Z.; Wang, Q.; Zhang, F.; Wang, Z.; Bowling, T.; Nare, B.; Jacobs, R. T.; Zhang, J.; Ding, D.; Liu, Y.; Zhou, H., *J. Med. Chem.*, **2012**, *55*, 3553-3557
18. Zhang, Y.; Plattner, J. J.; Freund, Y. R.; Easom, E. E.; Zhou, Y.; Gut, J.; Rosenthal, P. J.; Waterson, D.; Gamo, F.; Angulo-Barturen, I.; Ge, M.; Li, Z.; Li, L.; Jian, Y.; Cui, H.; Wang, H.; Yang, J., *Bioorganic & Medicinal Chemistry Letters*, **2011**, *21*, 644–651.
19. Xia, Y.; Cao, K.; Zhou, Y.; Alley, M. R. K.; Rock, F.; Mohan, M.; Meewan, M.; Baker, S. J.; Lux, S.; Ding, C. Z.; Jia, G.; Kully, M.; Plattner, J. J., *Bioorganic & Medicinal Chemistry Letters*, **2011**, *21*, 2533–2536.
20. Ding, D.; Zhao, Y.; Meng, Q.; Xie, D.; Nare, B.; Chen, D.; Bacchi, C. J.; Yarlett, N.; Zhang, Y.; Hernandez, V.; Xia, Y.; Freund, Y.; Abdulla, M.; Ang, K.; Ratnam, J.; McKerrow, J. H.; Jacobs, R. T.; Zhou, H.; Plattner, J. J., *Med. Chem. Lett.*, **2010**, *1*, 165–169.

21. Akama, T.; Baker, S. J.; Zhang, Y.; Hernandez, V.; Zhou, H.; Sanders, V.; Freund, Y.; Kimura, R.; Maples, K. R.; Plattner, J. J., *Bioorganic & Medicinal Chemistry Letters*, **2009**, *19*, 2129–2132.
22. Nicolaou, K. C.; Ramanjulu, J. M.; Natarajan, S.; Brase, S.; Li, H.; Boddy, C. N. C.; Rubsam, F., *Chem. Commun.*, **1997**, *19*, 1899-1900.
23. Jacobs, R. T.; Plattner, J. J.; Nare, B.; Wring, S. A.; Chen, D.; Freund, Y.; Gaukel, E. G.; Orr, M. D.; Perales, J. B.; Jenks, M.; Noe, R. A.; Sligar, J. M.; Zhang, Y.; Bacchi, C. J.; Yarlett, N.; Don, R., *Future Med. Chem.*, **2011**, *3*, 1259-1278.
24. (a) Kumar, J. S.; Alam, M. A.; Gurrupu, S.; Nelson, G.; Williams, M.; Corsello, M. A.; Johnson, J. L.; Jonnalagadda, S. C.; Mereddy, V. R. *J. Heterocyclic Chem.* **2013**, *50*, 814-820. (b) Kumar, J. S.; Bashian, C. M.; Corsello, M. A.; Jonnalagadda, S. C.; Mereddy, V. R. *Tetrahedron Lett.* **2010**, *51*, 4482-4485. (c) Kumar, J. S.; Jonnalagadda, S. C.; Mereddy, V. R. *Tetrahedron Lett.* **2010**, *51*, 779-782. (d) Gunasekera, D. S.; Gerold, D. A.; Alderks, N. A.; Jonnalagadda, S. C.; Kiprof, P.; Zhdankin, V. V.; Reddy, M. V. R. *Tetrahedron* **2007**, *63*, 9401-9405.
25. For reviews on Baylis Hillman reaction, please refer: (a) Singh, V.; Batra, S. *Tetrahedron*, **2008**, *64*, 4511; (b) Basavaiah, D.; Rao, K. V.; Reddy, R. J. *Chem. Soc. Rev.* **2007**; *26*, 1581. (c) Basavaiah, D.; Rao, A. J.; Satyanarayana, T. *Chem. Rev.* **2003**; *103*, 811. (d) Ciganek, E. *Organic Reactions*, Ed. Paquette, L. A.; John Wiley & Sons. Inc., New York, **1997**, *51*, p. 201; (e) Basavaiah, D.; Rao P. D.; Hyma, R. S. *Tetrahedron*, **1996**, *52*, 8001; (f) Drewes, S. E.; Roos, G. H. P. *Tetrahedron*, **1988**, *44*, 4653.
26. Boric Acid General Fact Sheet-National Pesticide Information Center: <http://npic.orst.edu/factsheets/boricgen.pdf>
27. (a) Shteinberg, L. Y. *Russ. J. Appl. Chem.* **2009**, *82*, 613-617. (b) Kondaiah, G. C. M.; Reddy, L. A.; Babu, K. S.; Gurav, V. M.; Hüge, K. M.; Bandichhor, R.; Reddy, P. P.; Bhattacharya, A.; Anand, R. V. *Tetrahedron Lett.* **2008**, *49*, 106–109. (c) Nath, J.; Chaudhuri, M. K. *Green Chemistry Letters and Reviews*, **2008**, *1*, 223-230. (d) Tu, S-J.; Zhu, X-T.; Fang, F.; Zhang, X-J.; Zhu, S-L.; Li, T-J.; Shi, D-Q.; Wang, X-S.; Ji, S-J.; *Chin. J. Chem.* **2005**, *23*, 596-598. (e) Chaudhuri, M. K.; Hussain, S.; Kantam, M. L.; Neelima, B.; *Tetrahedron Lett.* **2005**, *46*, 8329-8331. (f) Maki, T.; Ishihara, K.; Yamamoto, H. *Org. Lett.* **2005**, *7*, 5047-5050. (g) Stapp, P. R. *J. Org. Chem.* **1973**, *38*, 1433-1434.
28. (a) Banfi, L.; Riva, R. *Organic Reactions* **2005**, *65*, 1-141. (b) Domling, A.; Ugi, I. *Angew Chem. Int. Ed.* **2000**, *39*, 3168-3210.

29. Schenck, H. A.; Lenkowski, P. W.; Mukherjee, I. C.; Kob, S. H.; Stables, J. P.; Patel, M. K.; Brown, M. L. *Bioorg. Med. Chem.* **2004**, *12*, 979-993. (b) Kuduk, S. D.; Chang, R. K.; DiPardo, R. M.; Di Marco, C. N.; Murphy, K. L.; Ransom, R. W.; Reiss, D. R.; Tang, C.; Prueksaritanont, T.; Pettibone, D. J.; Bock, M. G. *Bioorg. Med. Chem. Lett.* **2008**, *18*, 5107-5110.
30. (a) Yue T.; Wang M.-X.; Wang D.-X.; Masson G.; Zhu J. *Angew. Chem. Intl. Ed.* **2009**, *48*, 6717-21. (b) Yue, T.; Wang, M. X.; Wang, D. X.; Masson G.; Zhu, J. *J. Org. Chem.* **2009**, *74*, 8396-8399. (c) Mihara, H.; Xu, Yingjie; Shepherd, N. E.; Matsunaga, S.; Shibasaki, M. *J. Am. Chem. Soc.* **2009**, *131*, 8384-8385. (d) Wang, S.; Wang, M.-X.; Wang, D.-X.; Zhu, J. *Eur. J. Org. Chem.* **2007**, *24*, 4076-4080. (e) Wang, S.-X.; Wang, M.-X.; Wang, D.-X.; Zhu, J. *Org. Lett.* **2007**, *9*, 3615-3618. (f) Denmark, S. E.; Bui, T. *J. Org. Chem.* **2005**, *70*, 10190-10193. (g) Denmark, S. E.; Fan, Y. *J. Am. Chem. Soc.* **2003**, *125*, 7825-7827. (h) Seebach, D.; Adam, G.; Gees, T.; Schiess, M.; Weigand, W. *Chem. Ber.* **1988**, *121*, 507-17. (i) Hagedorn, I.; Eholzer, U. *Chem. Ber.* **1965**, *98*, 936-940.
31. (a) Nicolaou, K. C.; Natarajan, S.; Li, H.; Jain, N. F.; Hughes, R.; Solomon, M. E.; Ramanjulu, J. M.; Boody, C. N. C.; Takayanagi, M. *Angew. Chem. Intl. Ed.*, **1998**, *37*, 2708; (b) Nicolaou, K. C.; Li, H.; Boody, C. N. C.; Ramanjulu, J. M.; Yue, T. Y.; Natarajan, S.; Chu, X. J.; Brase, S.; Rubsam, F. *Chem. Eur. J.*, **1999**, *5*, 2584; (c) Tan, Y. L.; White, A. J. P.; Widdowson, D. A.; Wilhelm, R.; Williams, D. J. *J. Chem. Soc. Perkin Trans 1*, **2001**, 3269.
32. (a) Akama, T.; Baker, S. J.; Zhang, Y. K.; Hernandez, V.; Zhou, H.; Sanders, V.; Freund, Y.; Kimura, R.; Maples, K. R.; Plattner, J. J. *Bioorg. Med. Chem. Lett.* **2009**, *19*, 2129-2132. (b) Wozniack, A. A.; Cyranski, M. K.; Zubrowska, A.; Sporzynski, A. *J. Organomet. Chem.* **2009**, *694*, 3533-3541. (c) Baker, S. J.; Zhang, Y. K.; Akama, T.; Lau, A.; Zhou, H.; Hernandez, V.; Mao W.; Alley, M. R. K.; Sanders, V.; Plattner, J. J. *J. Med. Chem.* **2006**, *49*, 4447-4450.

APPENDIX

