

Developing a Gene Deletion Strategy for *Candida krusei*

Katelynn Perez, Biology Major, College of Biological Sciences

Mentor: Professor Dana Davis

Abstract:

Candida krusei is a pathogenic fungus that is a frequent cause of candidiasis in humans. In the United States, approximately 1.6% of fungal bloodstream infections and 4% of cancers, such as breast cancer and leukemia, are directly related to *C. krusei*. Although it is an important pathogenic species, *C. krusei* is not well studied because of the lack of genetic tools for its research. Here, we show a strategy for the targeted deletion of *CCC2* and *HOS2* from the genome of *C. krusei*. Copper-transporting P-type ATPase, encoded by *CCC2*, is required for iron uptake in *C. krusei*. In order to begin the deletion process of *CCC2*, DNA purification was performed. After purifying the *C. krusei* genome, this gene was amplified via PCR using *CCC2* detection primers. *CCC2* was replaced by a *NAT1* cassette, which offers cell resistance to nourseothricin, an antibiotic to which many organisms are sensitive. The *NAT1* cassette was obtained through PCR amplification using *CCC2* disruption primers. In amplifying the *NAT1* cassette, the disruption primers attached 60 nucleotides of flanking sequence on each side of the cassette. This flanking sequence is homologous to the region upstream and downstream of *CCC2* in the *C. krusei* genome. The targeted deletion of *HOS2*, which is a gene required for NAD dependent/independent histone deacetylase activity, was performed following a similar protocol for the deletion of *CCC2*. Alteration to this procedure included the use of *HOS2* detection primers and *HOS2* disruption primers. The genomes of the *C. krusei* transformants were purified, and amplified via PCR, using the gene-specific detection primers. The transformant DNA was then visualized through gel electrophoresis. The results obtained from this experiment lay the ground work for future research on *C. krusei* by addressing a strategy for targeted gene deletion in this species.

C. krusei: A Fungal Pathogen Necessary for the Fermentation of Chocolate

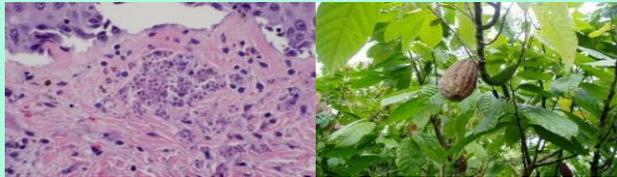


Figure 1. (Left) Papillary dermis tissue infected by *C. krusei* (dermatology.cdlib.org). (Right) This pathogenic fungal species is also necessary for the fermentation of chocolate from the Cacao bean (botany.wisc.edu).

Amplification of *NAT1* or *HygB* using Gene-Specific DR Primers

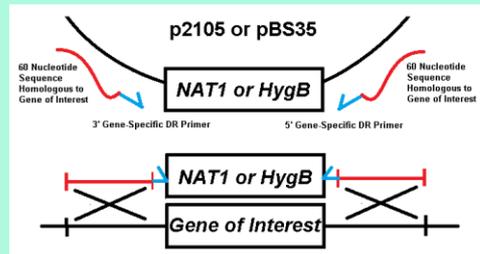


Figure 2. Amplification of *NAT1* or *HygB* using gene-specific disruption primers. A 60 nucleotide flank, which was homologous to the region upstream and downstream of the gene of interest, was added to *NAT1* and *HygB* during amplification.

Nourseothricin and Hygromycin Impede the Growth of Wild-Type *C. krusei*

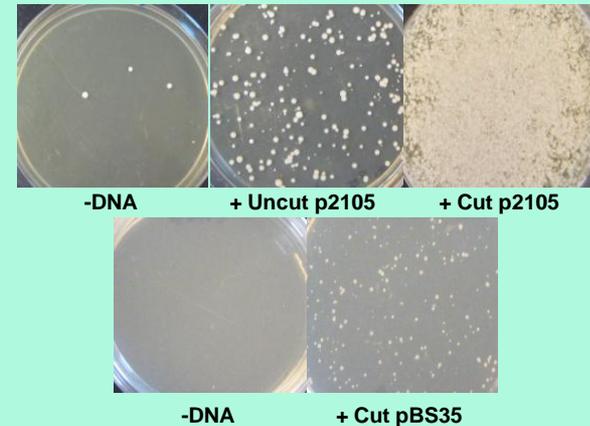


Figure 3. (Top) Chemical transformation of *C. krusei* with cut and uncut p2105, a plasmid containing the *NAT1* cassette. The cells transformed with cut p2105 exhibited the most growth, relative to the untransformed cells and the cells transformed with uncut plasmid. (Bottom) Chemical transformation of *C. krusei* with cut pBS35, a plasmid containing *HygB*. These results indicate that cut plasmids integrate most efficiently in *C. krusei*.

Chemical Transformation of *CCC2* Allele in *C. krusei* with the *NAT1* Cassette

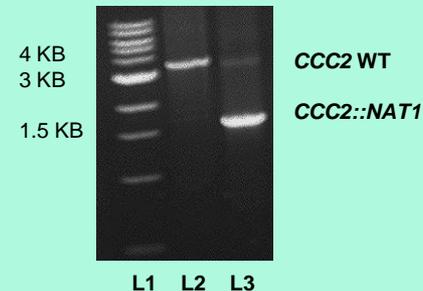


Figure 4. Heterozygous deletion of *CCC2* in *C. krusei* was achieved through chemical transformation using the *NAT1* cassette. The marker used was a 1KB ladder, as shown in L1. L2 shows *C. krusei* WT. L3 shows *C. krusei* as a diploid organism, indicated by the band at approximately 3.3 KB, which represents one intact *CCC2* allele. The band at approximately 1.6 KB represents the *NAT1* cassette, which replaced one *CCC2* allele.